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(21) International Application Number: PCT/US93/12586 (22) International Filing Date: 22 December 1993 (22.12.93) (30) Priority Data: 07/996,772 24 December 1992 (24.12.92) US (60) Parent Application or Grant (63) Related by Continuation US 07/996,772 (CIP) Filed on 24 December 1992 (24.12.92) (71) Applicant (for all designated States except US): SYNAPTIC PHARMACEUTICAL CORPORATION [US/US]; 215 College Road, Paramus, NJ 07652 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): GERALD, Christophe [FR/US]; 204-B, Union Street, Ridgewood, NJ 07450 (US). HARTIG, Paul [US/US]; 19 Pheasant Run, Kinnelon, NJ 07405 (US). BRANCHEK, Theresa, A. [US/US]; 541 Martense Avenue, Teaneck, NJ 07666 (US). WEINSHANK, Richard, L. [US/US]; 302 West 87th Street, New York, NY 10024 (US).		(74) Agent: COBERT, Robert, J.; Cooper & Dunham, 30 Rockefeller Plaza, New York, NY 10112 (US). (81) Designated States: AU, CA, FI, HU, JP, KR, NO, NZ, RU, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>Without international search report and to be republished upon receipt of that report.</i>
(54) Title: DNA ENCODING 5-HT ₄ SEROTONIN RECEPTORS AND USES THEREOF (57) Abstract <p>This invention provides an isolated nucleic acid molecule encoding a mammalian 5-HT₄ receptor and an isolated nucleic acid molecule encoding a human 5-HT₄ receptor, an isolated protein which is a mammalian 5-HT₄ receptor, an isolated protein which is a human 5-HT₄ receptor, vectors comprising an isolated nucleic acid molecule encoding a mammalian 5-HT₄ receptor, vectors comprising an isolated nucleic acid molecule encoding a human 5-HT₄ receptor, mammalian cells comprising such vectors, antibodies directed to the 5-HT₄ receptor, nucleic acid probes useful for detecting nucleic acid encoding a mammalian or human 5-HT₄ receptor, antisense oligonucleotides complementary to any sequences of a nucleic acid molecule which encodes a mammalian or human 5-HT₄ receptor, pharmaceutical compounds related to the human 5-HT₄ receptor, and nonhuman transgenic animals which express DNA encoding a normal or a mutant mammalian or human 5-HT₄ receptor. This invention further provides methods for determining ligand binding, detecting expression, drug screening, and treatments for alleviating abnormalities associated with a human 5-HT₄ receptor.</p>		

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5 DNA ENCODING 5-HT₁, SEROTONIN RECEPTORS AND USES THEREOFBackground of the Invention

10 Throughout this application various publications are referred to by partial citations within parenthesis. Full citations for these publications may be found at the end of the specification immediately preceding the claims. The disclosures of these publications, in their entireties, are hereby incorporated by reference
15 into this application in order to more fully describe the state of the art to which this invention pertains.

Molecular cloning efforts have provided primary amino acid sequence and signal transduction data for a large
20 collection of serotonin receptor subtypes. These include five cloned 5-HT₁-like receptors, three cloned 5-HT₂ receptors, and one 5-HT₃ receptor. The 5-HT₁ subfamily includes: 5-HT_{1A} (Fargin, 1988; Kobilka, 1989), 5-HT_{1B}/5-HT_{1D} (Weinshank et al., 1991; Demchyshyn et al., 1992; Jin et al., 1992; Adham et al., 1992; Maroteaux et al., 1992; Voight et al., 1991), 5-HT_{1D}
25 (Branchek et al. 1991; Hamblin and Metcalf, 1991; Weinshank et al., 1992), 5-HT_{1E} (Levy et al., 1992; McAllister et al., 1992; Zgombick et al., 1992) and 5-HT_{1F} (Adham et al., 1993). All five have been shown to couple to the inhibition of adenylate cyclase activity. The 5-HT₂ family includes the 5-HT₂ receptor (Pritchett et al., 1988), 5-HT_{2C} (Julius et al., 1989) and 5-HT_{2F}
30 (Rat Stomach Fundus; Foquet et al., 1992; Kursar et al., 1992). These receptors all couple to phosphoinositide hydrolysis. The 5-HT₃ receptor is a ligand-gated ion channel (Maricq et al., 1991).
35

Alth ough this work represents enormous success, the
40 absence of molecular biological information on the 5-

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HT₁ receptors, which have been shown in native tissues to couple to the activation of adenylate cyclase as a primary mode of signal transduction (Dumius et al., 1988; Bockaert et al., 1990), is apparent. In a
5 previous copending application (U.S. Serial No., 971,690, filed November 3, 1992), we reported the cloning of the first mammalian 5-HT receptor that couples to the stimulation of adenylate cyclase activity which we named 5-HT_{4B}. The 5-HT_{4B} receptor was
10 subsequently renamed to the "5-HT₇ receptor" by the "Serotonin Receptor Nomenclature Committee" of the IUPHAR. The pharmacological properties of this receptor indicated that it was similar to a series of functionally defined 5-HT receptors described in the
15 porcine vena cava (Trevethick et al., 1984), cat saphenous vein, coronary arteries (Cushing and Cohen, 1992), and several vascular dilatory effects (Mylecharane and Phillips, 1989). However, the classically defined 5-HT₁ receptor remained to be
20 cloned. We now report the cloning of the pharmacologically-defined 5-HT₁ receptor which we have previously called 5-HT_{4A} and now designate as the 5-HT₁ receptor. This receptor also stimulates adenylate
25 cyclase activity but unlike 5-HT_{4B}, is sensitive to a series of benzamide derivatives which act as agonists or partial agonists at this subtype. The presence of this subtype in the brain, particularly in areas such as the hippocampus, indicates a potential role in cognitive enhancement. In addition, the 5-HT₁ receptor
30 has been described functionally in the heart, adrenal, bladder, and alimentary canal indicating potential roles in achalasia, hiatal hernia, esophageal spasm, irritable bowel disease, postoperative ileus, diabetic gastroparesis, emesis and other diseases of the
35 gastrointestinal tract, as well as in cardiac, urinary, and endocrine function.

Summary of the Invention

This invention provides an isolated nucleic acid molecule encoding a mammalian 5-HT₁ receptor. In a preferred embodiment of this invention, the isolated nucleic acid encodes a human 5-HT₁ receptor. In another embodiment of this invention, the nucleic acid molecule encoding a human 5-HT₁ receptor comprises a plasmid designated pBluescript-hS10 (ATCC Accession No. 75392). In another embodiment of this invention a nucleic acid molecule encoding a mammalian 5-HT₁ receptor comprises a plasmid designated pcEXV-S10-87 (ATCC Accession No. 75390). In another embodiment of this invention a nucleic acid molecule encoding a mammalian 5-HT₁ receptor comprises a plasmid designated pcEXV-S10-95 (ATCC Accession No. 75391).

This invention provides a nucleic acid probe comprising a nucleic acid molecule of at least 15 nucleotides capable of specifically hybridizing with a unique sequence included within the sequence of a nucleic acid molecule encoding a mammalian 5-HT₁ receptor. This invention also provides a nucleic acid molecule of at least 15 nucleotides capable of specifically hybridizing with a sequence included within the sequence of a nucleic acid molecule encoding a human 5-HT₁ receptor.

This invention provides an antisense oligonucleotide having a sequence capable of binding specifically to an mRNA molecule encoding a mammalian 5-HT₁ receptor so as to prevent translation of the mRNA molecule. This invention also provides an antisense oligonucleotide having a sequence capable of binding specifically to an mRNA molecule encoding a human 5-HT₁ receptor so as to prevent translation of the mRNA molecule.

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This invention provides a monoclonal antibody directed to a mammalian 5-HT₁ receptor. This invention also provides a monoclonal antibody directed to a human 5-HT₁ receptor.

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This invention provides a pharmaceutical composition comprising an amount of a substance effective to alleviate the abnormalities resulting from overexpression of a mammalian 5-HT₁ receptor and a pharmaceutically acceptable carrier. This invention also provides a pharmaceutical composition comprising an amount of a substance effective to alleviate abnormalities resulting from underexpression of mammalian 5-HT₁ receptor and a pharmaceutically acceptable carrier.

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This invention provides a pharmaceutical composition comprising an amount of a substance effective to alleviate the abnormalities resulting from overexpression of a human 5-HT₁ receptor and a pharmaceutically acceptable carrier. This invention also provides pharmaceutical composition comprising an amount of a substance effective to alleviate abnormalities resulting from underexpression of a human 5-HT₁ receptor and a pharmaceutically acceptable carrier.

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This invention provides a transgenic, nonhuman mammal whose genome comprises DNA encoding a mammalian 5-HT₁ receptor so positioned within such genome as to be transcribed into antisense mRNA complementary to mRNA encoding the mammalian 5-HT₁ receptor and when hybridized to mRNA encoding the mammalian 5-HT₁ receptor, the complementary mRNA reduces the translation of the mRNA encoding the mammalian 5-HT₁ receptor.

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5 This invention also provides a transgenic, nonhuman mammal whose genome comprises DNA encoding a human 5-HT₁ so positioned within such genome as to be transcribed into antisense mRNA complementary to mRNA encoding the human 5-HT₁, and when hybridized to mRNA encoding the human 5-HT₁, the complementary mRNA reduces the translation of the mRNA encoding the human 5-HT₁.

10 This invention provides a transgenic, nonhuman mammal whose genome comprises DNA encoding a mammalian 5-HT₁ receptor so positioned within such genome as to be transcribed into antisense mRNA which is complementary to mRNA encoding the mammalian 5-HT₁ receptor and when
15 hybridized to mRNA encoding the 5-HT₁ receptor, the antisense mRNA thereby prevents the translation of mRNA encoding the 5-HT₁ receptor.

20 This invention also provides a transgenic, nonhuman mammal whose genome comprises DNA encoding a human 5-HT₁ receptor so positioned within such genome as to be transcribed into antisense mRNA which is complementary to mRNA encoding the human 5-HT₁ receptor and when
25 hybridized to mRNA encoding the human 5-HT₁ receptor, the antisense mRNA thereby prevents the translation of mRNA encoding the human 5-HT₁ receptor.

30 This invention also provides a method of determining the physiological effects of expressing varying levels of a mammalian 5-HT₁ receptor which comprises producing a transgenic nonhuman animal whose levels of mammalian 5-HT₁ receptor expression are varied by use of an inducible promoter which regulates mammalian 5-HT₁ receptor expression.

35 This invention also provides a method of determining the physiological effects of expressing varying levels

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of a human 5-HT₄ receptor which comprises producing a transgenic nonhuman animal whose levels of human 5-HT₄ receptor expression are varied by use of an inducible promoter which regulates human 5-HT₄ receptor expression.

This invention further provides a method of determining the physiological effects of expressing varying levels of mammalian 5-HT₄ receptor which comprises producing a panel of transgenic nonhuman animals each expressing a different amount of mammalian 5-HT₄ receptor.

This invention further provides a method of determining the physiological effects of expressing varying levels of human 5-HT₄ receptor which comprises producing a panel of transgenic nonhuman animals each expressing a different amount of human 5-HT₄ receptor.

This invention provides a method for determining whether a compound not known to be capable of specifically binding to a human 5-HT₄ receptor can specifically bind to the human 5-HT₄ receptor, which comprises contacting a mammalian cell comprising a plasmid adapted for expression in a mammalian cell which plasmid further comprises a DNA which expresses a human 5-HT₄ receptor on the cell's surface with the compound under conditions permitting binding of ligands known to bind to a human 5-HT₄ receptor, detecting the presence of any compound bound to the human 5-HT₄ receptor, the presence of bound compound indicating that the compound is capable of specifically binding to the human 5-HT₄ receptor.

This invention provides a method of screening drugs to identify drugs which interact with, and specifically bind to, a human 5-HT₄ receptor on the surface of a cell, which comprises contacting a mammalian cell which

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comprises a plasmid adapted for expression in a mammalian cell which plasmid further comprises DNA which expresses a human 5-HT₁ receptor on the cell's surface with a plurality of drugs, determining those drugs which bind to the human 5-HT₁ receptor expressed on the cell surface of the mammalian cell, and thereby identifying drugs which interact with, and specifically bind to, the human 5-HT₁ receptor.

This invention provides a method for identifying a compound which specifically binds to and activates or blocks the activation of a human 5-HT₁ receptor on the surface of a mammalian cell, which comprises contacting the mammalian cell which comprises a plasmid adapted for expression in the mammalian cell such plasmid further comprising DNA which expresses the human 5-HT₁ receptor on the cell surface of the mammalian cell with the compound, determining whether the compound activates or blocks the activation of the human 5-HT₁ receptor and thereby identifying the compound as a compound which binds to, and activates or blocks the activation of the human 5-HT₁ receptor.

This invention provides a method for diagnosing a predisposition to a disorder associated with the expression of a human 5-HT₁ receptor allele which comprises: a.) obtaining DNA of subjects suffering from the disorder; b.) performing a restriction digest of the DNA with a panel of restriction enzymes; c.) electrophoretically separating the resulting DNA fragments on a sizing gel; d.) contacting the resulting gel with a nucleic acid probe capable of specifically hybridizing to DNA encoding a 5-HT₁ receptor and labelled with a detectable marker; e.) detecting labelled bands which have hybridized to the DNA encoding a 5-HT₁ receptor labelled with a detectable marker to create a unique band pattern specific to the

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- 5 DNA of subjects suffering from the disorder; f.) preparing DNA obtained for diagnosis by steps a-e; and g.) comparing the unique band pattern specific to the DNA of subjects suffering from the disorder from step e and the DNA obtained for diagnosis from step f to determine whether the patterns are the same or different and to diagnose thereby predisposition to the disorder if the patterns are the same.

Brief Description of Figures

5 **Figure 1:** Nucleotide and corresponding amino acid sequence of the S10-87 cDNA clone. Only partial 5' and 3' untranslated sequences are shown.

10 **Figure 2:** Nucleotide and corresponding amino acid sequence of the S10-95 cDNA clone. Only partial 5' and 3' untranslated sequences are shown.

Figure 3: Comparison of amino acid sequences between clones S10-87 (top row) and S10-95 (bottom row). The overall homology is 96.7%.

15 **Figure 4:** Comparison of the rat S10 receptor deduced amino acid sequences with those of other serotonin receptors and with the canine histamine H2 receptor. Solid bars, the seven putative membrane-spanning domains (TM I-VII). Shading, homologies between the
20 S10 receptors and other receptors. Hp78, 5-HT4B or hp78a receptor (U.S. Serial No., 971,960, filed, November 3, 1992, copending).

25 **Figure 5:** Nucleotide and amino acid sequences of the human S10 PCR clone. The numbering is given according to the rat S10-95 clone.

30 **Figure 6:** Comparison of nucleotide sequences between the human PCR S10 clone and the rat S10 cDNA clone. Top row: human sequence, the numbering is given according to the rat S10 nucleotide sequence. The bottom row outlines differences in the rat sequence (overall homology: 90.7%).

35 **Figure 7:** Comparison of deduced amino acid sequences between the Human S10 PCR clone and the rat S10 cDNA clone. Top row: human S10 sequence, the numbering is

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given according to the rat S10 amino acid sequence. Th bott m r w outlines differences in the rat sequence (overall homology: 92.3 %).

5 **Figure 8:** Comparison of binding affinities of key compounds at the S10 clone with adenylate cyclase functional responses obtained with mouse collicular neurons. A correlation plot was constructed between affinity constants of drugs for the S10 receptor with
10 those obtained at a pharmacologically defined 5-HT₁ receptor. Binding values for the correlation were taken from table 1 and were expressed as the negative logarithm. Functional data were taken from Dumuis et al. (1988). The correlation coefficient calculated by
15 linear regression was 0.96 indicating that the rank order of potency for the compounds was similar in both preparations.

20 **Figure 9:** Stimulation of cAMP production by 5-HT in transiently transfected Cos-7 cells expressing the cloned rat 5-HT₁ (CG-7) receptor and antagonism by ICS 205930. cAMP measurements on intact cells were as described under Methods and Materials. Each data point represents the mean of triplicates from a single
25 experiment representative of at least 2 others. The vertical bars indicate S.E.M. Data are presented as percent maximum cAMP released by 5-HT (basal cAMP release: 0.020 ± 0.002 pmol/ml/10 min; maximum cAMP release: 0.42 ± 0.03 pmol/ml/10 min).

30 **Figure 10:** Stimulation of cAMP production by 5-HT in transiently transfected Cos-7 cells expressing the cloned rat 5-HT₁ (CG-8) receptor and antagonism by ICS 205930. cAMP measurements on intact cells were as
35 described under Methods and Materials. Each data point represents the mean of triplicates from a single experiment representative of at least two others. The

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vertical bars indicate S.E.M. Data are presented as percent maximum CAMP released by 5-HT (basal CAMP release: 0.023 ± 0.004 pmol/ml/10 min; maximum CAMP release: 0.57 ± 0.04 pmol/ml/10 min).

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Figure 11A: Nucleotide sequence of the partial human S10-87 clone. Only partial 3' untranslated sequences are shown (SEQ. ID NO. 14).

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Figure 11B: Deduced amino acid sequence encoded by the nucleotide sequence of Figure 11A of the partial human S10-87 clone (SEQ. ID NO. 15).

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Figure 12: Comparison of the nucleotide sequences between the human (top row) and the rat S10-87 (bottom row) cDNA clones. The overall identity is 90.8%.

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Figure 13: Comparison of the deduced amino acid sequences between the human (top row) and the rat (bottom row) S10-87 receptors. The overall identity is 93.9%.

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Figure 14A: Nucleotide sequence of the full length human S10-95 clone (SEQ. ID NO. 7).

Figure 14B: Deduced amino acid sequence encoded by the nucleotide sequence of Figure 14A (SEQ. ID NO. 8).

30

Figure 15: Comparison of the nucleotide sequences between the human (top row) and the rat (bottom row) S10-95 cDNA clones. The overall identity is 90.7%.

35

Figure 16: Comparison of the deduced amino acid sequences between the human (top row) and the rat (bottom row) S10-95 receptors. The overall identity is 93.8%.

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Figure 17: Comparison of the nucleotide sequences corresponding to the available coding regions between the two human isoforms (top row S10-95; bottom row S10-87) of the 5-HT₂ receptor. The overall identity is 92%.

Figure 18: Comparison of the deduced amino acid sequences between the two human isoforms (top row S10-95; bottom row S10-87) of the 5-HT₂ receptor. The overall identity is 90%.

Figure 19: Inhibition of [³H]GR11380 binding on the cloned rat CG-8 receptor by 5-HT, in the absence and presence of Gpp(NH)p (100 μM). Membranes harvested from transient transfectants (COS-7 cells) were incubated with [³H]GR11380 (0.2-0.4 nM) for 30 min at 37°C. Nonspecific binding was defined by 50 μM unlabelled 5-HT. Data are from a single experiment. Data were analyzed by computer-assisted nonlinear regression analysis (Accufit; Lunden Software).

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Detail d Descripti n f th Inventi n

This invention provides an isolated nucleic acid molecule encoding a mammalian 5-HT₁ receptor. This invention further provides an isolated nucleic acid molecule encoding a human 5-HT₁ receptor. As used herein, the term "isolated nucleic acid molecule" means a non-naturally occurring nucleic acid molecule that is, a molecule in a form which does not occur in nature. Examples of such an isolated nucleic acid molecule are an RNA, cDNA, or isolated genomic DNA molecule encoding a mammalian 5-HT₁ receptor or a human 5-HT₁ receptor. As used herein, "5-HT₁ receptor" means a molecule which, under physiologic conditions, is substantially specific for the neurotransmitter serotonin, is saturable, of high affinity for serotonin and the activation of which is coupled to the activation of adenylate cyclase and the "5-HT₁ receptor" is also sensitive to benzamide derivatives which act as agonists and partial agonists at this receptor subtype. One embodiment of this invention is an isolated nucleic acid molecule encoding a mammalian 5-HT₁ receptor. Such a molecule may have coding sequences substantially the same as the coding sequences shown in Figures 1 and 2 and 5 (SEQ ID NOS. 1, 3 and 5). A preferred embodiment is an isolated nucleic acid molecule encoding a human 5-HT₁ receptor. Such a molecule may have a coding sequence substantially the same as the coding sequence shown in Figure 5 (SEQ ID NO. 5). The DNA molecules of Figures 1, 2 and 5 (Seq ID NOS. 1, 3 and 5) encode the sequence of mammalian 5-HT₁ receptors. The DNA molecule of Figure 5 (Seq ID No. 5) encodes a human 5-HT₁ receptor. This invention further provides isolated DNA molecules encoding mammalian 5-HT₁ receptors having the sequence H₂N-Y-X-COOH wherein Y is the amino acid sequence beginning at amino acid 1 and ending at amin

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acid 359 of Figure 1 (SEQ ID NOS. 1 and 2) and wherein X is an amino acid sequence encoding the carboxy terminal region of the receptor. The nucleic acid molecules of Figures 1 and 2 (SEQ ID NOS 1-4) encode

5 5-HT₁ receptors having an identical sequence Y and differing only in their carboxy terminal region X beginning at amino acid 360. One means of isolating a nucleic acid molecule encoding a mammalian 5-HT₁ receptor is to probe a mammalian genomic library with

10 a natural or artificially designed DNA probe, using methods well known in the art. In the preferred embodiment of this invention, the mammalian 5-HT₁ receptor is a human protein and the nucleic acid molecule encoding the human 5-HT₁ receptor is isolated

15 from human cDNA. Degenerate oligonucleotide primers derived from transmembrane (TM) domains of 5-HT_{1A}, 5-HT_{1C}, 5-HT₂ and 5-HT_{10A/8} receptors are useful for identifying cDNA containing a nucleic acid molecule encoding a 5-HT₁ receptor, obtaining a probe specific

20 to a mammalian 5-HT₁ receptor and for isolating a nucleic acid molecule encoding a mammalian 5-HT₁ receptor.

DNA and cDNA molecules which encode a mammalian 5-HT₁ receptor are used to obtain complementary genomic DNA,

25 cDNA or RNA from human, mammalian or other animal sources, or to isolate related cDNA or genomic clones by the screening of cDNA or genomic libraries, by methods described in more detail below.

30 Transcriptional regulatory elements from the 5' untranslated region of the isolated clone, and other stability, processing, transcription, translation, and tissue specificity determining regions from the 3' and 5' untranslated regions of the isolated gene are

35 thereby obtained.

This invention provides an isolated nucleic acid

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molecule which has a nucleic acid sequence which differs from the sequence of a nucleic acid molecule encoding a 5-HT₁ receptor at one or more nucleotides and which does not encode a protein having 5-HT₁ receptor activity. As used herein, "5-HT₁ receptor activity" means the capability of receptor to specifically bind the neurotransmitter, serotonin under physiological conditions and the capability of the receptor to activate adenylate cyclase when the receptor is coupled to adenylate cyclase. An example of a isolated nucleic acid molecule provided by this invention is a nucleic acid molecule which has an in-frame stop codon inserted into the coding sequence such that the transcribed RNA is not translated into protein.

This invention further provides a cDNA molecule encoding a mammalian 5-HT₁ receptor, wherein the cDNA molecule has a coding sequence substantially the same as the coding sequence shown in Figures 1, 2 and 5 (SEQ ID NOS. 1, 3 and 5). This invention provides a cDNA molecule encoding a human 5-HT₁ receptor, wherein the cDNA molecule has a coding sequence substantially the same as the coding sequence shown in Figure 5 (SEQ ID NO. 5). These molecules and their equivalents were obtained by the means described above.

This invention also provides an isolated protein which is a mammalian 5-HT₁ receptor. In a preferred embodiment of this invention, the protein is a human 5-HT₁ receptor protein having an amino acid sequence substantially similar to the amino acid sequence shown in Figures 1, 2 and 5 (SEQ ID Nos. 1-6). In another embodiment of this invention, the protein is a murine 5-HT₁ receptor protein having an amino acid sequence substantially similar to the amino acid sequence shown in Figures 1, 2 and 5 (SEQ ID NOS. 1-6). As used

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herein, the term "isolated protein" is intended to encompass a protein molecule free of other cellular components. One means for obtaining an isolated mammalian 5-HT₁ receptor protein is to express DNA encoding the 5-HT₁ receptor in a suitable host, such as a bacterial, yeast, insect, or mammalian cell, using methods well known to those skilled in the art, and recovering the receptor protein after it has been expressed in such a host, again using methods well known in the art. The receptor may also be isolated from cells which express it, in particular from cells which have been transfected with the expression vectors described below in more detail.

This invention provides a vector comprising DNA, RNA, or cDNA, encoding a mammalian 5-HT₁ receptor. This invention further provides a vector comprising DNA, RNA, or cDNA, encoding a human 5-HT₁ receptor. Examples of vectors are viruses such as bacteriophages (such as phage lambda), cosmids, plasmids (such as pUC18, available from Pharmacia, Piscataway, NJ), and other recombination vectors. Nucleic acid molecules are inserted into vector genomes by methods well known to those skilled in the art. Examples of such plasmids are plasmids comprising DNA having a coding sequence substantially the same as the coding sequence shown in Figures 1, 2 and 5 (SEQ ID NOS. 1, 3 and 5) and designated pcEXV-S10-87 (ATCC Accession No. 75390), pcEXV-S10-95 (ATCC Accession No. 75391) and pBLuescript-hS10 (ATCC No. 75392).

Alternatively, to obtain these vectors, insert and vector DNA can both be exposed to a restriction enzyme to create complementary ends on both molecules which base pair with each other and are then ligated together with a ligase. Alternatively, linkers can be ligated to the insert DNA which correspond to a restriction

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site in the vector DNA, which is then digested with the restriction enzyme which cuts at that site. Other means are also available.

5 This invention also provides vectors comprising a DNA or cDNA encoding a mammalian 5-HT₁ receptor and vectors comprising a DNA or cDNA encoding a human 5-HT₁ receptor, adapted for expression in a bacterial cell, a yeast cell, insect cell or a mammalian cell which
10 additionally comprise the regulatory elements necessary for expression of the DNA or cDNA encoding a mammalian 5-HT₁ receptor or the DNA or cDNA encoding a human 5-HT₁ receptor in the bacterial, yeast, insect or mammalian cells operatively linked to the DNA or cDNA encoding
15 the 5-HT₁ receptor as to permit expression thereof. DNA or cDNA having coding sequence substantially the same as the coding sequence shown in Figures 1 and 2 (SEQ ID NOs. 1 and 3) may be usefully inserted into these vectors to express a mammalian 5-HT₁ receptor.
20 DNA or cDNA having a coding sequence substantially the same as the coding sequence shown in Figure 5 (SEQ ID NO. 5) may be usefully inserted into these vectors to express the human 5-HT₁ receptor. Regulatory elements required for expression include promoter sequences to
25 bind RNA polymerase and transcription initiation sequences for ribosome binding. For example, a bacterial expression vector includes a promoter such as the lac promoter and for transcription initiation the Shine-Dalgarno sequence and the start codon AUG
30 (Maniatis, et al., 1982). Similarly, a eukaryotic expression vector includes a heterologous or homologous promoter for RNA polymerase II, a downstream polyadenylation signal, the start codon AUG, and a termination codon for detachment of the ribosome.
35 Furthermore, an insect expression vector, such as recombinant Baculovirus, uses the polyhedrin gene expression signals for expression of the inserted gene

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in insect cells. Such vectors may be obtained commercially or assembled from the sequences described by methods well known in the art, for example the methods described above for constructing vectors in general. Expression vectors are useful to produce cells that express receptors. Certain uses for such cells are described in more detail below.

In one embodiment of this invention a plasmid is adapted for expression in a bacterial, yeast, insect, or, in particular, a mammalian cell wherein the plasmid comprises DNA or cDNA encoding a mammalian 5-HT₁ receptor or DNA or cDNA encoding a human 5-HT₁ receptor and the regulatory elements necessary for expression of the DNA in the bacterial, yeast, insect, or mammalian cell operatively linked to the DNA or cDNA encoding a mammalian 5-HT₁ receptor or to the DNA or cDNA encoding a human 5-HT₁ receptor as to permit expression thereof. Suitable plasmids may include, but are not limited to plasmids adapted for expression in a mammalian cell, e.g., EVJB, EXV-3. An example of such a plasmid adapted for expression in a mammalian cell is a plasmid comprising cDNA having coding sequences substantially the same as the coding sequence shown in Figures 1, 2 and 5 (SEQ ID NOS. 1, 3 and 5) and the regulatory elements necessary for expression of the DNA in the mammalian cell. These plasmids have been designated pcEXV-S10-87 deposited under ATCC Accession No. 75390, pcEXV-S10-95 deposited under ATCC Accession No. 75391, and pBluescript-hS10, deposited under ATCC Accession No. 75392. Those skilled in the art will readily appreciate that numerous plasmids adapted for expression in a mammalian cell which comprise DNA encoding a mammalian or human 5-HT₁ receptor and the regulatory elements necessary to express such DNA in the mammalian cell may be constructed utilizing existing plasmids and adapted as appropriate to contain

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th regulatory elements necessary to express the DNA in the mammalian cell. The plasmids may be constructed by th meth ds described above for expression vectors and vectors in general, and by other methods well known in the art.

Deposit discussed supra were made pursuant to, and in satisfaction of, the provisions of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland 20852.

This invention provides a mammalian cell comprising a DNA or cDNA molecule encoding a mammalian 5-HT₁ receptor, such as a mammalian cell comprising a plasmid adapted for expression in a mammalian cell, said plasmid further comprises DNA or cDNA encoding a mammalian 5-HT₁ receptor and the regulatory elements necessary for expression of the DNA or cDNA in the mammalian cell operatively linked to the DNA or cDNA encoding a mammalian 5-HT₁ receptor as to permit expression thereof. This invention provides a mammalian cell comprising a DNA or cDNA molecule encoding a human 5-HT₁ receptor, such as a mammalian cell comprising a plasmid adapted for expression in a mammalian cell, said plasmid further comprises a DNA or cDNA molecule encoding a human 5-HT₁ receptor and the regulatory elements necessary for expression of the DNA or cDNA in the mammalian cell operatively linked to the DNA or cDNA encoding a human 5-HT₁ receptor as to permit expression thereof. Numerous mammalian cells may be used as hosts, including, but not limited to, the mouse fibroblast cell NIH3T3, CHO cells, HeLa cells, LM (tk-) cells, Cos-7 cells, etc. Expression plasmids such as that described supra may be used to transfect mammalian cells by methods well known in the art such as calcium

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phosphate precipitation, or DNA or cDNA encoding a human or mammalian 5-HT₁ receptor may be otherwise introduced into mammalian cells, e.g., by microinjection, to obtain mammalian cells which
5 comprise DNA, e.g., cDNA or a plasmid, encoding a human or mammalian 5-HT₁ receptor.

This invention provides a nucleic acid probe comprising a nucleic acid molecule of at least 15 nucleotides
10 capable of specifically hybridizing with an unique sequence included within the sequence of a nucleic acid molecule encoding a human 5-HT₁ receptor, for example with a coding sequence included within the sequences shown in Figure 5 (SEQ ID NO. 5). This invention
15 further provides a nucleic acid probe comprising a nucleic acid molecule of at least 15 nucleotides capable of specifically hybridizing with a sequence included within the sequence of a nucleic acid molecule encoding a mammalian 5-HT₁ receptor, for example with
20 a coding sequence included within the sequences shown in Figure 1 and Figure 2 (SEQ ID NOs. 1 and 3). As used herein, the phrase "specifically hybridizing" means the ability of a nucleic acid molecule to recognize a nucleic acid sequence complementary to its
25 own and to form double-helical segments through hydrogen bonding between complementary base pairs. As used herein, the phrase "unique sequence" means a nucleic acid molecule sequence specific to only the nucleic acid molecule encoding a mammalian 5-HT₁
30 receptor. Nucleic acid probe technology is well known to those skilled in the art who will readily appreciate that such probes may vary greatly in length and may be labeled with a detectable label, such as a radioisotope or fluorescent dye, to facilitate detection of the
35 probe. Detection of nucleic acid encoding a human 5-HT₁ receptor is useful as a diagnostic test for any disease process in which levels of expression of the 5-

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HT₁ receptor are altered. DNA probe molecules are produced by insertion of a DNA molecule which encodes a 5-HT₁ receptor or fragments thereof into suitable vectors, such as plasmids or bacteriophages, followed by insertion into suitable bacterial host cells and replication and harvesting of the DNA probes, all using methods well known in the art. For example, the DNA may be extracted from a cell lysate using phenol and ethanol, digested with restriction enzymes corresponding to the insertion sites of the DNA into the vector (discussed above), electrophoresed, and cut out of the resulting gel. An example of such DNA molecules is shown in Figures 1, 2 and 5 (SEQ ID NOS. 1, 3, and 5). The probes are useful for 'in situ' hybridization or in order to locate tissues which express this gene family, or for other hybridization assays for the presence of these genes or their mRNA in various biological tissues. In addition, synthesized oligonucleotides (produced by a DNA synthesizer) complementary to the sequence of a DNA molecule which encode a mammalian 5-HT₁ receptor or complementary to the sequence of a DNA molecule which encodes a human 5-HT₁ receptor are useful as probes for these genes, for their associated mRNA, or for the isolation of related genes by homology screening of genomic or cDNA libraries, or by the use of amplification techniques such as the polymerase chain reaction.

This invention also provides a method of detecting expression of a human 5-HT₁ receptor on the surface of a cell by detecting the presence of mRNA coding for a 5-HT₁ receptor. This invention further provides a method of detecting expression of a mammalian 5-HT₁ receptor on the surface of the cell by detecting the presence of mRNA coding for a mammalian 5-HT₁ receptor. These methods comprise obtaining total mRNA from the cell using methods well known in the art and contacting

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the mRNA so obtained with a nucleic acid probe as described hereinabove, under hybridizing conditions, detecting the presence of mRNA hybridized to the probe, and thereby detecting the expression of the receptor by the cell. Hybridization of probes to target nucleic acid molecules such as mRNA molecules employs techniques well known in the art. However, in one embodiment of this invention, nucleic acids are extracted by precipitation from lysed cells and the mRNA is isolated from the extract using a column which binds the poly-A tails of the mRNA molecules (Maniatis et al., 1982). The mRNA is then exposed to radioactively labelled probe on a nitrocellulose membrane, and the probe hybridizes to and thereby labels complementary mRNA sequences. Binding may be detected by autoradiography or scintillation counting. However, other methods for performing these steps are well known to those skilled in the art, and the discussion above is merely an example.

This invention provides an antisense oligonucleotide having a sequence capable of binding specifically with any sequences of an mRNA molecule which encodes a human 5-HT₁ receptor so as to prevent translation of the human 5-HT₁ receptor. The antisense oligonucleotide may have a sequence capable of binding specifically with any sequences of the cDNA molecule whose sequence is shown in Figure 5 (SEQ ID NO. 5). This invention also provides an antisense oligonucleotide having a sequence capable of binding specifically with any sequences of an mRNA molecule which encodes a mammalian 5-HT₁ receptor so as to prevent translation of the mammalian 5-HT₁ receptor. The antisense oligonucleotide may have a sequence capable of binding specifically with any sequences of the cDNA molecule whose sequence is shown in Figures 1 and 2 (SEQ ID NOS. 1 and 3). As used herein, the phrase "binding specifically" means the

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ability of an antisense ligonucleotide to recognize a nucleic acid sequence complementary to its own and to form double-helical segments through hydrogen bonding between complementary base pairs. A particular example of an antisense oligonucleotide is an antisense oligonucleotide comprising chemical analogues of nucleotides.

This invention also provides a pharmaceutical composition comprising an effective amount of the oligonucleotide described above effective to reduce expression of a human 5-HT₂ receptor by passing through a cell membrane and binding specifically with mRNA encoding the 5-HT₂ receptor in the cell so as to prevent its translation and a pharmaceutically acceptable hydrophobic carrier capable of passing through a cell membrane. This invention further provides a pharmaceutical composition comprising an effective amount of the oligonucleotide described above effective to reduce expression of a mammalian 5-HT₂ receptor by passing through a cell membrane and binding specifically with mRNA encoding a mammalian 5-HT₂ receptor in the cell so as to prevent its translation and a pharmaceutically acceptable hydrophobic carrier capable of passing through a cell membrane. As used herein, the term "pharmaceutically acceptable carrier" encompasses any of the standard pharmaceutical carriers, such as a phosphate buffered saline solution, water, and emulsions, such as an oil/water or water/oil emulsion, and various types of wetting agents. The oligonucleotide may be coupled to a substance which inactivates mRNA, such as a ribozyme. The pharmaceutically acceptable hydrophobic carrier capable of passing through cell membranes may also comprise a structure which binds to a transporter specific for a selected cell type and is thereby taken up by cells of the selected cell type. The structure may be part of

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a protein known to bind a cell-type specific transporter, for example an insulin molecule, which would target pancreatic cells. DNA molecules having a coding sequence substantially the same as the coding sequences shown in Figure 5 (SEQ ID No. 5) may be used as the oligonucleotides of the pharmaceutical composition.

This invention also provides a pharmaceutical composition comprising an effective amount of the oligonucleotide described above effective to reduce expression of a mammalian 5-HT₂ receptor by passing through a cell membrane and binding specifically with mRNA encoding the 5-HT₂ receptor in the cell so as to prevent its translation and a pharmaceutically acceptable hydrophobic carrier capable of passing through a cell membrane. DNA molecules having a coding sequence substantially the same as the coding sequences shown in Figures 1 and 2 (SEQ ID NOs. 1 and 3) may be used as the oligonucleotides of the pharmaceutical composition.

This invention provides a method of treating abnormalities which are alleviated by reduction of expression of 5-HT₂ receptor. This method comprises administering to a subject an effective amount of the pharmaceutical composition described above effective to reduce expression of the 5-HT₂ receptor by the subject. This invention further provides a method of treating an abnormal condition related to 5-HT₂ receptor activity which comprises administering to a subject an amount of the pharmaceutical composition described above effective to reduce expression of the 5-HT₂ receptor by the subject. Examples of such abnormal conditions are irritable bowel disease, postoperative ileus, diabetic gastroparesis, emesis, achalasia, hiatal hernia, esophageal spasm and other diseases of the

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gastrointestinal tract, as well as in cardiac, urinary, and endocrine function.

Antisense oligonucleotide drugs inhibit translation of mRNA encoding 5-HT₄ receptor. Synthetic antisense oligonucleotides, or other antisense chemical structures are designed to bind to mRNA encoding the 5-HT₄ receptor and inhibit translation of mRNA and are useful as drugs to inhibit expression of 5-HT₄ receptor genes in patients. This invention provides a means to therapeutically alter levels of expression of a human or mammalian 5-HT₄ receptor by the use of a synthetic antisense oligonucleotide drug (SAOD) which inhibits translation of mRNA encoding the 5-HT₄ receptor. Synthetic antisense oligonucleotides, or other antisense chemical structures designed to recognize and selectively bind to mRNA, are constructed to be complementary to portions of the nucleotide sequence shown in Figure 1, 2 and 5 (SEQ ID NOS. 1, 3 and 5) of DNA, RNA or of chemically modified, artificial nucleic acids. The SAOD is designed to be stable in the blood stream for administration to patients by injection, or in laboratory cell culture conditions, for administration to cells removed from the patient. The SAOD is designed to be capable of passing through cell membranes in order to enter the cytoplasm of the cell by virtue of physical and chemical properties of the SAOD which render it capable of passing through cell membranes (e.g., by designing small, hydrophobic SAOD chemical structures) or by virtue of specific transport systems in the cell which recognize and transport the SAOD into the cell. In addition, the SAOD can be designed for administration only to certain selected cell populations by targeting the SAOD to be recognized by specific cellular uptake mechanisms which bind and take up the SAOD only within certain selected cell populations. For example, the SAOD may be designed to

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bind to transporter found only in a certain cell type, as discussed above. The SAOD is also designed to recognize and selectively bind to the target mRNA sequence, which may correspond to a sequence contained within the sequence shown in Figure 1, 2 and 5 (SEQ ID NOS. 1, 3, and 5) by virtue of complementary base pairing to the mRNA. Finally, the SAOD is designed to inactivate the target mRNA sequence by any of three mechanisms: 1) by binding to the target mRNA and thus inducing degradation of the mRNA by intrinsic cellular mechanisms such as RNase I digestion, 2) by inhibiting translation of the mRNA target by interfering with the binding of translation-regulating factors or of ribosomes, or 3) by inclusion of other chemical structures, such as ribozyme sequences or reactive chemical groups, which either degrade or chemically modify the target mRNA. Synthetic antisense oligonucleotide drugs have been shown to be capable of the properties described above when directed against mRNA targets (Cohen, J.S., 1989; Weintraub, H.M., 1990). In addition, coupling of ribozymes to antisense oligonucleotides is a promising strategy for inactivating target mRNA (N. Sarver et al., 1990). An SAOD serves as an effective therapeutic agent if it is designed to be administered to a patient by injection, or if the patient's target cells are removed, treated with the SAOD in the laboratory, and replaced in the patient. In this manner, an SAOD serves as a therapy to reduce 5-HT₂ receptor expression in particular target cells of a patient, in any clinical condition which may benefit from reduced expression of 5-HT₂ receptor.

This invention provides an antibody directed to the human 5-HT₂ receptor. This invention also provides an antibody directed to the mammalian 5-HT₂ receptor. This antibody may comprise, for example, a monoclonal

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antibody directed to an epitope of a human 5-HT₁ receptor present on the surface of a cell, the epitope having an amino acid sequence substantially the same as an amino acid sequence for a cell surface epitope of the human 5-HT₁ receptor included in the amino acid sequence shown in Figure 5. Amino acid sequences may be analyzed by methods well known to those skilled in the art to determine whether they produce hydrophobic or hydrophilic regions in the proteins which they build. In the case of cell membrane proteins, hydrophobic regions are well known to form the part of the protein that is inserted into the lipid bilayer which forms the cell membrane, while hydrophilic regions are located on the cell surface, in an aqueous environment. Therefore antibodies to the hydrophilic amino acid sequences shown in Figure 5 will bind to a surface epitope of a 5-HT₁ receptor as described. Antibodies directed to a human or mammalian 5-HT₁ receptor may be serum-derived or monoclonal and are prepared using methods well known in the art. For example, monoclonal antibodies are prepared using hybridoma technology by fusing antibody producing B cells from immunized animals with myeloma cells and selecting the resulting hybridoma cell line producing the desired antibody. Cells such as NIH3T3 cells or LM (tk⁻) cells may be used as immunogens to raise such an antibody. Alternatively, synthetic peptides may be prepared using commercially available machines and the amino acid sequence shown in Figures 1, 2, and 5 (SEQ ID NOs. 1-6). As a still further alternative, DNA, such as a cDNA or a fragment thereof, may be cloned and expressed and the resulting polypeptide recovered and used as an immunogen. These antibodies are useful to detect the presence of 5-HT₁ receptor encoded by the isolated DNA, or to inhibit the function of the 5-HT₁ receptor in living animals, in humans, or in biological tissues or fluids isolated from animals or humans.

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This invention also provides a pharmaceutical composition which comprises an effective amount of an antibody directed to an epitope of the human 5-HT₂ receptor, effective to block binding of naturally occurring substrates to the 5-HT₂ receptor, and a pharmaceutically acceptable carrier. A monoclonal antibody directed to an epitope of a human 5-HT₂ receptor present on the surface of a cell which has an amino acid sequence substantially the same as an amino acid sequence for a cell surface epitope of the human 5-HT₂ receptor included in the amino acid sequence shown in Figure 5 (SEQ ID NOS. 5 and 6) is useful for this purpose.

This invention also provides a pharmaceutical composition which comprises an effective amount of an antibody directed to an epitope of a mammalian 5-HT₂ receptor, effective to block binding of naturally occurring substrates to the 5-HT₂ receptor, and a pharmaceutically acceptable carrier. A monoclonal antibody directed to an epitope of a mammalian 5-HT₂ receptor present on the surface of a cell which has an amino acid sequence substantially the same as an amino acid sequence for a cell surface epitope of a mammalian 5-HT₂ receptor included in the amino acid sequence shown in Figures 1 and 2 (SEQ ID NOS. 1-4) is useful for this purpose.

This invention also provides a method of treating abnormalities in a subject which are alleviated by reduction of expression of a human or mammalian 5-HT₂ receptor which comprises administering to the subject an effective amount of the pharmaceutical composition described above effective to block binding of naturally occurring substrates to the receptor and thereby alleviate abnormalities resulting from overexpression of a human or mammalian 5-HT₂ receptor. Binding of the antibody to the receptor prevents the receptor from

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functioning, thereby neutralizing the effects of over expression. The monoclonal antibodies described above are useful for this purpose. This invention additionally provides a method of treating an abnormal condition related to an excess of 5-HT₂ receptor activity which comprises administering to a subject an amount of the pharmaceutical composition described above effective to block binding of naturally occurring substrates to the 5-HT₂ receptor and thereby alleviate the abnormal condition. Some examples of abnormal conditions associated with excess 5-HT₂ receptor activity are irritable bowel disease, postoperative ileus, diabetic gastroparesis, emesis, achalasia, hiatal hernia, esophageal spasm and other diseases of the gastrointestinal tract, as well as in cardiac, urinary, and endocrine function.

This invention provides methods of detecting the presence of a 5-HT₂ receptor on the surface of a cell which comprises contacting the cell with an antibody directed to the 5-HT₂ receptor, under conditions permitting binding of the antibody to the receptor, detecting the presence of the antibody bound to the cell, and thereby the presence of the 5-HT₂ receptor on the surface of the cell. Such methods are useful for determining whether a given cell is defective in expression of 5-HT₂ receptors. Bound antibodies are detected by methods well known in the art, for example by binding fluorescent markers to the antibodies and examining the cell sample under a fluorescence microscope to detect fluorescence on a cell indicative of antibody binding. The monoclonal antibodies described above are useful for this purpose.

This invention provides a transgenic nonhuman mammal expressing DNA encoding a human 5-HT₂ receptor and a transgenic nonhuman mammal expressing DNA encoding a

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mammalian 5-HT₂ receptor. This invention also provides a transgenic nonhuman mammal expressing DNA encoding a human or mammalian 5-HT₂ receptor so mutated as to be incapable of normal receptor activity, and not expressing native 5-HT₂ receptor. This invention further provides a transgenic nonhuman mammal whose genome comprises DNA encoding a human 5-HT₂ receptor so placed as to be transcribed into antisense mRNA which is complementary to mRNA encoding a human 5-HT₂ receptor and which hybridizes to mRNA encoding a 5-HT₂ receptor thereby reducing its translation and a transgenic nonhuman mammal whose genome comprises DNA encoding a mammalian 5-HT₂ receptor so placed as to be transcribed into antisense mRNA which is complementary to mRNA encoding a mammalian 5-HT₂ receptor and which hybridizes to mRNA encoding a mammalian 5-HT₂ receptor thereby reducing its translation. The DNA may additionally comprise an inducible promoter or additionally comprise tissue specific regulatory elements, so that expression can be induced, or restricted to specific cell types. Examples of DNA are DNA or cDNA molecules having a coding sequence substantially the same as the coding sequences shown in Figure 1, 2 and 5 (SEQ ID NOS. 1, 3, and 5). An example of a transgenic animal is a transgenic mouse. Examples of tissue specificity-determining regions are the metallothionein promoter (Low et al., 1986) and the L7 promoter (Oberdick et al., 1990).

Animal model systems which elucidate the physiological and behavioral roles of mammalian receptors are produced by creating transgenic animals in which the expression of a receptor is either increased or decreased, or the amino acid sequence of the expressed receptor protein is altered, by a variety of techniques. Examples of these techniques include, but are not limited to: 1) Insertion of normal or mutant

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versions of DNA encoding a human 5-HT₁ receptor or homologous animal versions of this gene, by microinjection, retroviral infection or other means well known to those skilled in the art, into appropriate fertilized embryos in order to produce a transgenic animal (Hogan et al., 1986) or, 2) Homologous recombination (Capecchi M.R., 1989; Zimmer A, and Gruss, P., 1989) of mutant or normal, human or animal versions of these genes with the native gene locus in transgenic animals to alter the regulation of expression or the structure of the receptor. The technique of homologous recombination is well known in the art. It replaces the native gene with the inserted gene and so is useful for producing an animal that cannot express native receptor but does express, for example, an inserted mutant receptor, which has replaced the native receptor in the animal's genome by recombination, resulting in underexpression of the receptor. Microinjection adds genes to the genome, but does not remove them, and so is useful for producing an animal which expresses its own and added receptors, resulting in overexpression of the receptor.

One means available for producing a transgenic animal, with a mouse as an example, is as follows: Female mice are mated, and the resulting fertilized eggs are dissected out of their oviducts. The eggs are stored in an appropriate medium such as M2 medium (Hogan, B. et al. 1986). DNA or cDNA encoding a receptor is purified from a vector (such as plasmids pcEXV-S10-87, pcEXV-S10-95 and pBluescript-hS10 described above) by methods well known in the art. Inducible promoters may be fused with the coding region of the DNA to provide an experimental means to regulate expression of the trans-gene. Alternatively or in addition, tissue specific regulatory elements may be fused with the coding region to permit tissue-specific expression of

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the trans-gene. The DNA, in an appropriately buffered solution, is put into a microinjection needle (which may be made from capillary tubing using a pipet puller) and the egg to be injected is put in a depression slide. The needle is inserted into the pronucleus of the egg, and the DNA solution is injected. The injected egg is then transferred into the oviduct of a pseudopregnant mouse (a mouse stimulated by the appropriate hormones to maintain pregnancy but which is not actually pregnant), where it proceeds to the uterus, implants, and develops to term. As noted above, microinjection is not the only method for inserting DNA into the egg cell, and is used here only for exemplary purposes.

Since the normal action of receptor-specific drugs is to activate or to inhibit the receptor, the transgenic animal model systems described above are useful for testing the biological activity of drugs directed against the receptors even before such drugs become available. These animal model systems are useful for predicting or evaluating possible therapeutic applications of drugs which activate or inhibit receptors by inducing or inhibiting expression of the native or trans-gene and thus increasing or decreasing expression of normal or mutant receptors in the living animal. Thus, a model system is produced in which the biological activity of drugs directed against the receptors are evaluated before such drugs become available. The transgenic animals which over or under produce the receptor indicate by their physiological state whether over or under production of the receptor is therapeutically useful.

It is therefore useful to evaluate drug action based on the transgenic model system. One use is based on the fact that it is well known in the art that a drug such as an antidepressant acts by blocking neurotransmitter

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uptake, and thereby increases the amount of neurotransmitter in the synaptic cleft. The physiological result of this action is to stimulate the production of less receptor by the affected cells, leading eventually to underexpression. Therefore, an animal which underexpresses receptor is useful as a test system to investigate whether the actions of such drugs which result in under expression are in fact therapeutic. Another use is that if overexpression is found to lead to abnormalities, then a drug which down-regulates or acts as an antagonist to the receptor is indicated as worth developing, and if a promising therapeutic application is uncovered by these animal model systems, activation or inhibition of the 5-HT₁ receptor is achieved therapeutically either by producing agonist or antagonist drugs directed against the 5-HT₁ receptor or by any method which increases or decreases the expression of this receptor in man.

Further provided by this invention is a method of determining the physiological effects of expressing varying levels of human or mammalian 5-HT₁ receptors which comprises producing a transgenic nonhuman animal whose levels of human or mammalian 5-HT₁ receptor expression are varied by use of an inducible promoter which regulates receptor expression. This invention also provides a method of determining the physiological effects of expressing varying levels of human or mammalian 5-HT₁ receptor which comprises producing a panel of transgenic nonhuman animals each expressing a different amount of human or mammalian 5-HT₁ receptor. Such animals may be produced by introducing different amounts of DNA encoding a human or mammalian 5-HT₁ receptor into the oocytes from which the transgenic animals are developed.

This invention also provides a method for identifying

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a substance capable of alleviating abnormalities resulting from overexpression of a human or mammalian 5-HT₁ receptor comprising administering the substance to a transgenic nonhuman mammal expressing at least one artificially introduced DNA molecule encoding a human or mammalian 5-HT₁ receptor and determining whether the substance alleviates the physical and behavioral abnormalities displayed by the transgenic nonhuman mammal as a result of overexpression of a human or mammalian 5-HT₁ receptor. As used herein, the term "substance" means a compound or composition which may be natural, synthetic, or a product derived from screening. Examples of DNA molecules are DNA or cDNA molecules having a coding sequence substantially the same as the coding sequences shown in Figures 1, 2, and 5 (SEQ ID NOS. 1, 3, and 5).

This invention provides a pharmaceutical composition comprising an amount of the substance described supra effective to alleviate the abnormalities resulting from overexpression of 5-HT₁ receptor and a pharmaceutically acceptable carrier.

This invention further provides a method for treating the abnormalities resulting from overexpression of a human or mammalian 5-HT₁ receptor which comprises administering to a subject an amount of the pharmaceutical composition described above effective to alleviate the abnormalities resulting from overexpression of a human or mammalian 5-HT₁ receptor.

This invention provides a method for identifying a substance capable of alleviating the abnormalities resulting from underexpression of a human or mammalian 5-HT₁ receptor comprising administering the substance to the transgenic nonhuman mammal described above which expresses only nonfunctional human or mammalian 5-HT₁

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receptor and determining whether the substance alleviates the physical and behavioral abnormalities displayed by the transgenic nonhuman mammal as a result of underexpression of a human or mammalian 5-HT₁ receptor.

This invention also provides a pharmaceutical composition comprising an amount of a substance effective to alleviate abnormalities resulting from underexpression of a human or mammalian 5-HT₁ receptor and a pharmaceutically acceptable carrier.

This invention further provides a method for treating the abnormalities resulting from underexpression of a human or mammalian 5-HT₁ receptor which comprises administering to a subject an amount of the pharmaceutical composition described above effective to alleviate the abnormalities resulting from underexpression of a human or mammalian 5-HT₁ receptor.

This invention provides a method for diagnosing a predisposition to a disorder associated with the expression of a human or mammalian 5-HT₁ receptor allele which comprises: a) obtaining DNA of subjects suffering from the disorder; b) performing a restriction digest of the DNA with a panel of restriction enzymes; c) electrophoretically separating the resulting DNA fragments on a sizing gel; d) contacting the resulting gel with a nucleic acid probe capable of specifically hybridizing to DNA encoding a human or mammalian 5-HT₁ receptor and labelled with a detectable marker; e) detecting labelled bands which have hybridized to the DNA encoding a human or mammalian 5-HT₁ receptor labelled with a detectable marker to create a unique band pattern specific to the DNA of subjects suffering from the disorder; f) preparing DNA obtained for diagnosis by steps a-e; and

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g) comparing the unique band pattern specific to the DNA of subjects suffering from the disorder from step e and the DNA obtained for diagnosis from step f to determine whether the patterns are the same or different and thereby to diagnose predisposition to the disorder if the patterns are the same. This method may also be used to diagnose a disorder associated with the expression of a specific human 5-HT₁ receptor allele or mammalian 5-HT₁ receptor allele.

This invention provides a method of preparing the isolated 5-HT₁ receptor which comprises inducing cells to express receptor, recovering the receptor from the resulting cells, and purifying the receptor so recovered. An example of a 5-HT₁ receptor is an isolated protein having substantially the same amino acid sequence as the amino acid sequence shown in Figure 5. For example, cells can be induced to express receptors by exposure to substances such as hormones. The cells can then be homogenized and the receptor isolated from the homogenate using an affinity column comprising, for example serotonin or another substance which is known to bind to the 5-HT₁ receptor. The resulting fractions can then be purified by contacting them with an ion exchange column, and determining which fraction contains 5-HT₁ receptor activity or binds anti-receptor antibodies.

This invention provides a method of preparing an isolated human 5-HT₁ receptor which comprises inserting nucleic acid encoding the human 5-HT₁ receptor in a suitable vector, inserting the resulting vector in a suitable host cell, recovering the receptor produced by the resulting cell, and purifying the receptor so recovered. An example of an isolated human 5-HT₁ receptor is an isolated protein having substantially the same amino acid sequence as the amino acid sequence

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sh wn in Figure 5 (SEQ ID NOS. 5 and 6). This invention provides a m thod f preparing an isolated mammalian 5-HT₁ receptor which compris s inserting nucleic acid encoding the mammalian 5-HT₁ receptor in a suitable vector, inserting the resulting vector in a suitable host cell, recovering the receptor produced by the resulting cell, and purifying the receptor so recovered. An example of an isolated mammalian 5-HT₁ receptor is an isolated protein having substantially the same amino acid sequence as the amino acid sequence shown in Figures 1 and 2 (SEQ ID NOS. 1- 2 and Seq I. D. Nos. 3-4, respectively). These methods for preparing 5-HT₁ receptor uses recombinant DNA technology methods well known in the art. For example, isolated nucleic acid encoding 5-HT₁ receptor is inserted in a suitable vector, such as an expression vector. A suitable host cell, such as a bacterial cell, insect cell, or a eukaryotic cell such as a yeast cell, is transfected with the vector. 5-HT₁ receptor is isolated from the culture medium by affinity purification or by chromatography or other methods well known in the art.

This invention provides a method for determining whether a compound not known to be capable of specifically binding to a human 5-HT₁ receptor can specifically bind to the human 5-HT₁ receptor, which comprises contacting a mammalian cell comprising a plasmid adapted for expression in a mammalian cell which plasmid further comprises a DNA which expresses a human 5-HT₁ receptor on the cell's surface with the compound under conditions permitting binding of ligands known to bind to a human 5-HT₁ receptor, detecting the presence of any compound bound to the human 5-HT₁ receptor, the presence of bound compound indicating that the compound is capable of specifically binding to the human 5-HT₁ receptor.

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This invention provides a method for determining whether a compound not known to be capable of specifically binding to a mammalian 5-HT₁ receptor can specifically bind to the mammalian 5-HT₁ receptor, which comprises contacting a mammalian cell comprising a plasmid adapted for expression in a mammalian cell which plasmid further comprises a DNA which expresses a mammalian 5-HT₁ receptor on the cell's surface with the compound under conditions permitting binding of ligands known to bind to a mammalian 5-HT₁ receptor, detecting the presence of any compound bound to the human 5-HT₁ receptor, the presence of bound compound indicating that the compound is capable of specifically binding to the mammalian 5-HT₁ receptor.

This invention provides a method for identifying a compound which is not known to be capable of binding to a human 5-HT₁ receptor can functionally activate the human 5-HT₁ receptor on the surface of a mammalian cell or prevent a ligand which does so, which comprises contacting the mammalian cell which cell comprises a plasmid adapted for expression in the mammalian cell such plasmid further comprising DNA which expresses the human 5-HT₁ receptor on the surface of the mammalian cell with the compound, determining whether the compound activates the human 5-HT₁ receptor or prevents a ligand which does so, and thereby identifying the compound as a compound which binds to and functionally activates the human 5-HT₁ receptor or prevents the functional activation of the human 5-HT₁ receptor by a ligand which does so.

The DNA in the cell may have a coding sequence substantially the same as the coding sequence shown in Figure 5 (SEQ ID No. 5).

This invention provides a method for identifying a compound which is not known to be capable of binding to

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a mammalian 5-HT₁ receptor can functionally activate the mammalian 5-HT₁ receptor on the surface of a mammalian cell or prevent a ligand which does so, which comprises contacting the mammalian cell which cell
5 comprises a plasmid adapted for expression in the mammalian cell such plasmid further comprising DNA which expresses the mammalian 5-HT₁ receptor on the surface of the mammalian cell with the compound, determining whether the compound activates the
10 mammalian 5-HT₁ receptor or prevents a ligand which does so, and thereby identifying the compound as a compound which binds to and functionally activates the mammalian 5-HT₁ receptor or prevents the functional activation of the mammalian 5-HT₁ receptor by a ligand
15 which does so. The DNA in the cell may have a coding sequence substantially the same as the coding sequence shown in Figures 1 and 2 (SEQ ID NOS. 1 and 3).

The activation or blockade of the functional response is detected by means of a bioassay from the mammalian
20 cell such as a second messenger response, and thereby determining whether the compound activates or prevents the activation of the 5-HT₁ receptor functional output. Preferably, the mammalian cell is nonneuronal in
25 origin. An example of a nonneuronal mammalian cell is an LM (tk-) cell. Another example of a non-neuronal mammalian cell to be used for functional assays is a murine fibroblast cell line, specifically the NIH3T3 cell. The preferred method for determining whether a
30 compound is capable of binding to the 5-HT₁ receptor comprises contacting a transfected nonneuronal mammalian cell (i.e. a cell that does not naturally express any type of 5-HT or G-protein coupled receptor, thus will only express such a receptor if it is
35 transfected into the cell) expressing a 5-HT₁ receptor on its surface, or contacting a membrane preparation derived from such a transfected cell, with the compound

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under conditions which are known to prevail, and thus to be associated with, in vivo binding of ligands to a 5-HT₁ receptor, detecting the presence of any of the compound being tested bound to the 5-HT₁ receptor on the surface of the cell, and thereby determining whether the compound binds to, and activates or prevents the activation of the 5-HT₁ receptor. This response system is obtained by transfection of isolated DNA into a suitable host cell containing the desired second messenger system such as phosphoinositide hydrolysis, adenylate cyclase, guanylate cyclase or ion channels. Such a host system is isolated from pre-existing cell lines, or can be generated by inserting appropriate components of second messenger systems into existing cell lines. Such a transfection system provides a complete response system for investigation or assay of the activity of human 5-HT₁ receptor with compounds as described above.

Transfection systems are useful as living cell cultures for competitive binding assays between known or candidate drugs and ligands which bind to the receptor and which are labeled by radioactive, spectroscopic or other reagents. Membrane preparations containing the receptor isolated from transfected cells are also useful for these competitive binding assays. Functional assays of second messenger systems or their sequelae in transfection systems act as assays for binding affinity and efficacy in the activation of receptor function. A transfection system constitutes a "drug discovery system" useful for the identification of natural or synthetic compounds with potential for drug development that can be further modified or used directly as therapeutic compounds to activate or inhibit the natural functions of the human 5-HT₁ receptor. The transfection system is also useful for determining the affinity and efficacy of known drugs at

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human 5-HT₄ receptor sites.

This invention also provides a method of screening compounds to identify drugs which interact with, and specifically bind to, a human 5-HT₄ receptor on the surface of a cell, which comprises contacting a mammalian cell which comprises a plasmid adapted for expression in a mammalian cell which plasmid further comprises DNA which expresses a human 5-HT₄ receptor on the cell's surface with a plurality of compounds, determining those compounds which bind to the human 5-HT₄ receptor expressed on the cell surface of the mammalian cell, and thereby identifying compounds which interact with, and specifically bind to, the human 5-HT₄ receptor. The DNA in the cell may have a coding sequence substantially the same as the coding sequence shown in Figure 5 (SEQ ID NO. 5). This invention also provides a method of screening compounds to identify drugs which interact with, and specifically bind to, a mammalian 5-HT₄ receptor on the surface of a cell, which comprises contacting a mammalian cell which comprises a plasmid adapted for expression in a mammalian cell which plasmid further comprises DNA which expresses a mammalian 5-HT₄ receptor on the cell's surface with a plurality of compounds, determining those compounds which bind to the mammalian 5-HT₄ receptor expressed on the cell surface of the mammalian cell, and thereby identifying compounds which interact with, and specifically bind to, the mammalian 5-HT₄ receptor. The DNA in the cell may have a coding sequence substantially the same as the coding sequence shown in Figures 1 and 3 (SEQ ID NOs. 1 and 2). Various methods of detection may be employed. The compounds may be "labeled" by association with a detectable marker substance (e.g., radiolabel or a non-is topic label such as biotin). Preferably, the mammalian cell is nonneuronal in origin. An example of

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a nonneuronal mammalian cell is a Cos-7 cell. Drug candidates are identified by choosing chemical compounds which bind with high affinity to the expressed 5-HT₁ receptor protein in transfected cells, using radioligand binding methods well known in the art, examples of which are shown in the binding assays described herein. Drug candidates are also screened for selectivity by identifying compounds which bind with high affinity to one particular receptor but do not bind with high affinity to any other receptor subtypes or to any other known receptor. Because selective, high affinity compounds interact primarily with the target 5-HT₁ receptor site after administration to the patient, the chances of producing a drug with unwanted side effects are minimized by this approach.

This invention provides a pharmaceutical composition comprising a drug identified by the method described above and a pharmaceutically acceptable carrier. As used herein, the term "pharmaceutically acceptable carrier" encompasses any of the standard pharmaceutical carriers, such as a phosphate buffered saline solution, water, and emulsions, such as an oil/water or water/oil emulsion, and various types of wetting agents. Once the candidate drug has been shown to be adequately bioavailable following a particular route of administration, for example orally or by injection (adequate therapeutic concentrations must be maintained at the site of action for an adequate period to gain the desired therapeutic benefit), and has been shown to be non-toxic and therapeutically effective in appropriate disease models, the drug may be administered to patients by that route of administration determined to make the drug bioavailable, in an appropriate solid or solution formulation, to gain the desired therapeutic benefit.

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Applicants have identified a novel 5-HT receptor subtype protein, designated 5-HT₄, and have described methods for the identification of pharmacological compounds for therapeutic treatments. Pharmacological compounds which are directed against specific receptor subtypes provide effective new therapies with minimal side effects.

Elucidation of the molecular structures of the neuronal serotonin receptors is an important step in the understanding of serotonergic neurotransmission. This disclosure reports the isolation and amino acid sequence of a novel cDNA which encodes a human 5-HT₄ receptor. This disclosure reports the isolation, amino acid sequence, and functional expression of a two novel cDNAs which encode mammalian 5-HT₄ receptors. The identification of 5-HT receptor subtypes play a pivotal role in elucidating the molecular mechanisms underlying serotonergic transmission, and should also aid in the development of novel therapeutic agents.

A complementary DNA clone (designated pBluescript-hS10) encoding a serotonin receptor subtype, 5-HT₄, has been isolated from human brain, human heart and human retina. Additionally, two complementary DNA clones encoding the serotonin 5-HT₄ receptor subtype have been isolated from mammalian brain and their functional properties have been examined in mammalian cells. Analysis of 5-HT₄ structure and function provides a model for the development of drugs useful for the treatment of gastrointestinal conditions including irritable bowel disease, postoperative ileus, diabetic gastroparesis, emesis, achalasia, hiatal hernia, and esophageal spasm. In addition, 5-HT₄ receptors have been described functionally in the heart (Kaumann,

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1992), adrenal (Lefebvre et al., 1992), and bladder (Corsi et al., 1991) indicating possible roles in cardiac rate and force of contraction, endocrine control of cortisol secretion, and urinary incontinence or spasticity. 5-HT₂ receptors have also been described in the brain, particularly in areas such as the hippocampus, in which we have localized the gene encoding 5-HT₂ receptors (S10-95), indicating a potential role in cognitive enhancement (Bockaert et al., 1992).

This invention identifies a mammalian serotonin receptor, its amino acid sequence, and its mammalian gene, the activation of which is coupled to activation of adenylate cyclase. The information and experimental tools provided by this discovery are useful to generate new therapeutic agents, and new therapeutic or diagnostic assays for this receptor protein, its associated mRNA molecule or its associated genomic DNA. The information and experimental tools provided by this discovery will be useful to generate new therapeutic agents, and new therapeutic or diagnostic assays for this new serotonin receptor subtype, its associated mRNA molecule, or its associated genomic DNA.

Specifically, this invention relates to the isolation of human cDNA clone and mammalian cDNA clones encoding a new serotonin receptor, designated 5-HT₂. In addition, the mammalian 5-HT₂ receptors have been expressed in COS-7 cells by transfecting the cells with the plasmids pcEXV-S10-87 and pcEXV-S10-95. The pharmacological binding properties of the encoded 5-HT₂ receptor have been determined, and the binding properties classify this receptor as a novel serotonin receptor. Mammalian cell lines expressing the mammalian 5-HT₂ receptor on the cell surface have been constructed, thus establishing the first well-defined,

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cultured cell lines with which to study the novel 5-HT₂ receptor.

5 The invention will be better understood by reference to the Experimental Details which follow, but those skilled in the art will readily appreciate that the specific experiments detailed are only illustrative, and are not meant to limit the invention as described herein, which is defined by the claims which follow

10 thereafter.

Methods and Materials

PCR Amplification: The third (III) and fifth (V) transmembrane (TM) domains of the following receptors were aligned and used to synthesize a pair of degenerate primers: 5-HT_{1A}, 5-HT_{1C}, 5-HT₂ and the 5-HT_{10a/β} receptors. Primers 3.17 and 5.5 ([5'-TGGAATTCTG(C/T)G(C/T)IAT(A/C/T)(G/T)CICTGGA(C/T)(A/C)G(C/G)TA-3'] (SEQ ID No. 9), [5'-CATIA(G/C/A)I(G/A)IIA(G/A)IGG(T/G/A)AT(G/A)(T/A)A(G/A)AAIGC-3']) (SEQ ID No. 10) were used to amplify 5 μg of poly (A+) RNA from rat brain that was reverse transcribed by avian myeloblastosis virus reverse transcriptase (AMV). PCR was performed on single-stranded cDNA under the following conditions: 94°C for 1 min, 50°C for 2 min and 72°C for 3 min for 40 cycles. Following PCR, 90 μl of the reaction was phenol:chloroform extracted and precipitated; 10 μl was visualized on a gel using ethidium bromide staining. After precipitation the sample was treated with T4 DNA polymerase and digested with EcoR1 prior to separation on a 1% agarose gel. The DNA fragments (200 to 400 base pairs) were isolated from the gel, kinased and cloned into pBluescript. Recombinant clones were analyzed by sequencing. One fragment 270 base pairs in length, named S10, was identified. This sequence contained a "TM IV" like domain and represented a potentially new serotonin receptor. The corresponding full length cDNA was isolated from a rat brain cDNA library.

Rat PCR primers (from TM3 to TM7) were used to amplify single-stranded cDNA prepared from human heart, brain and retina, as described above. Those human PCR DNA fragments were subcloned in pBluescript and sequenced.

cDNA Library Construction, Screening and Sequencing:

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Rat brains were dissected from adult male CD rats (Charles River Laboratories) and total RNA was prepared by the guanidine thiocyanate method (Chirgwin, J.W. et al.; 1979). Poly A⁺RNA was purified with a Fast track kit (Invitrogen Corp., San Diego, CA). Double stranded (DS) cDNA was synthesized from 5 µg of poly A⁺RNA according to Gubler and Hoffman (Gubler, U. and B. J. Hoffman, 1983). The resulting DS cDNA was ligated to BstXI/EcoRI adaptors (Invitrogen Corp.), the excess of adaptors was removed by chromatography on Sepharose CL 4B (Pharmacia LKB) and the DNA was then size selected on a Gen-Pak Fax HPLC column (Zhao, D. et al., 1992) (Waters, Millipore Corp., Milford, MA). High molecular weight fractions were ligated in pCDM8 cut by BstXI (Invitrogen Corp.). The ligated DNA was electroporated in E.Coli MC 1061 (Gene Pulser, Biorad). A total of 20×10^6 independent clones with an insert mean size of 1.9 kb could be generated. Before amplification, the library was divided into pools of 2.5 to 5×10^4 independent clones. After 18 hours amplification, the pools were stored at -85°C in 20% glycerol.

100 pools of the cDNA library, representing 3.2×10^6 primary clones, were screened using exact PCR primers derived from the S10 PCR clone sequence. 1 µl (4×10^6 bacteria) of each amplified pool was subjected directly to 40 cycles of PCR and the resulting products analyzed by agarose gel electrophoresis and Southern blotting. Two out of four positive pools were analyzed further and by sib selection and plating out, two individual full length cDNA clones, S10-87 and S10-95, were isolated. DS-DNA was sequenced with a sequanase kit (US Biochemical, Cleveland, OH) according to the manufacturer. Nucleotide and peptide sequences analysis were performed with GCG programs.

Genomic Cloning and Sequencing: A human fibroblast

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genomic library in λ dash II ($\approx 1.5 \times 10^6$ total recombinants; Stratagene, LaJolla, CA) was screened using a 45 nt. oligonucleotide probe derived from the rat S10-87 receptor gene, designed in the 3' end of the carboxyl terminal tail (from the anti-sense strand [n u c l e o t i d e 1 2 2 0 - 1 2 6 4) , 5 ' TCAAAAGCATGATTCCAGGGACTCTGGGTCATTGTGTATGGG CAA 3' (SEQ ID No. 11) (see Fig.1). The oligomer was labeled with [32 P] γ ATP by using polynucleotide kinase.

Hybridization was performed at medium stringency conditions: 45°C. in a solution containing 37.5% formamide, 5x SSC (1x SSC is 0.15M sodium chloride, 0.015M sodium citrate), 1x Denhardt's solution (0.02% polyvinylpyrrolindone, 0.02% Ficoll, 0.02% bovine serum albumin), and 200 μ g/ μ l sonicated salmon sperm DNA. The filters were washed at 45°C. in 0.1x SSC containing 0.1% sodium dodecyl sulfate and exposed at -70°C. to Kodak XAR film in the presence of an intensifying screen. Lambda phage clones hybridizing with the probe were plaque purified and DNA was prepared for Southern blot analysis (Southern, 1975; Sambrook et al., 1989). A 900 bp Hind2/SstI hybridizing fragment was subcloned into pUC18 (Pharmacia, Piscataway, NJ)). Nucleotide sequence analysis was accomplished by the Sanger dideoxy nucleotide chain termination method (Sanger et al., 1977) on denatured double-stranded plasmid templates, using Sequenase (US Biochemical Corp., Cleveland, OH).

PCR amplification of a partial length human S10-87 cDNA clone:

The 900 bp Hind2/SstI fragment contained sequence encoding the human S10-87 carboxy terminal tail, including the stop codon. This sequence was used to generate a 25 mer (reverse primer) containing the stop codon: 5' CCTCAATCAGAAGCATGATTCCAGG 3' (SEQ ID No. 12). As a forward primer we used the 5' end of the human PCR

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fragment previously identified (figure 6):
5'TTGGTCTATAGGAACAAGATGACCC 3' (SEQ ID No. 13). These
human PCR primers were used to amplify single stranded
cDNA prepared from human brain as previously described.
5 The amplified DNA was subcloned and sequenced as
described above.

Isolation of the full length human S10-95 cDNA clone:
20 pools of a human hippocampal cDNA library (3kb
10 average size insert, in pcEXV-3) representing 10^6
independent clones were screened by PCR with TM4-TM6
primers as previously described. Five positive pools
were identified. one of those pools was analyzed
further and by sib selection a 5kb cDNA clone, CG-17,
15 was isolated. Double Stranded-DNA was sequenced as
described above. Nucleotide and peptide sequence
analysis were performed with the Genetics Computer
Group sequence analysis software package.

20 DNA transfection: The full coding region of S10-87
(clone CG-5) and S10-95 (clones CG-6 and CG-17) were
subcloned in the correct orientation in the mammalian
expression vectors pCDNA1-Amp (Invitrogen Corp.), and
pcEXV-3 (Miller, J. and R. N. Germain, 1986) (CG-7 and
25 CG-8 respectively). For transient expression, Cos-7
cells were transfected by the DEAE-Dextran method,
using 1 μ g of DNA / 10^6 cells (Warden, D. and H.V.
Thorne, 1968).

30 Membrane Preparation: Membranes were prepared from
transiently transfected COS-7 cells which were grown to
100% confluence. The cells were washed twice with
phosphate-buffered saline, scraped from the culture
dishes into 5 ml of ice-cold phosphate-buffered saline,
35 and centrifuged at 200 x g for 5 min at 4°. The pellet
was resuspended in 2.5 ml of ice-cold Tris buffer (20
mM Tris -HCl, pH 7.4 at 23°, 5 mM EDTA), and

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homogenized by a Wheaton tissue grinder. The lysate was subsequently centrifuged at 200 x g for 5 min at 4° to pellet large fragments which were discarded. The supernatant was collected and centrifuged at 40,000 x g for 20 min at 4°. The pellet resulting from this centrifugation was washed once in ice-cold Tris wash buffer and finally resuspended in a final buffer containing 50 mM Tris-HCl and 0.5 mM EDTA, pH 7.4 at 23°. Membrane preparations were kept on ice and utilized within two hr for the radioligand binding assays. Protein concentrations were determined by the method of Bradford (1976) using bovine serum albumin as the standard.

Radioligand Binding: [³H]5-HT binding was performed using slight modifications of the 5-HT₁₀ assay conditions reported by Herrick-Davis and Titeler (1989) with the omission of masking ligands. Radioligand binding studies were achieved at 37° C in a total volume of 250 µl of buffer (50 mM Tris, 10 mM MgCl₂, 0.2 mM EDTA, 10 µM pargyline, 0.1 % ascorbate, pH 7.4 at 37° C) in 96 well microtiter plates. Saturation studies were conducted using [³H]5-HT at 10 different concentrations ranging from 1.0 nM to 100 nM. Displacement studies were performed using 10 nM [³H]5-HT. The binding profile of drugs in competition experiments was established using 7 concentrations of compound. Incubation times were 30 min for both saturation and displacement studies. Nonspecific binding was defined in the presence of 10 µM 5-HT. Binding was initiated by the addition of 50 µl membrane homogenates (10-20 µg). The reaction was terminated by rapid filtration through presoaked (0.5% polyethyleneimine) filters using 48R Cell Brandel Harvester (Gaithersburg, MD). Subsequently, filters were washed for 5 sec with ice cold buffer (50 mM Tris HCL, pH 7.4 at 4° C), dried and placed into vials

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containing 2.5 ml of Radi-Safe (Beckman, Fullerton, CA), and radioactivity was measured using a Beckman LS 6500C liquid scintillation counter. The efficiency of counting of [^3H]5-HT averaged between 45-50%. Binding data were analyzed by computer-assisted nonlinear regression analysis (Accufit and Accucomp, Lunden Software, Chagrin Falls, OH). IC_{50} values were converted to K_i values using the Cheng-Prusoff equation (1973).

[^3H]GR113808 binding was performed using slight modifications of the method of Waeber et al., 1993. Radioligand binding studies were achieved at 37°C in a total volume of 250 μl of buffer (50 mM Tris, 10 μM , 0.01% ascorbate, pH 7.4 at 37°C) in 96 well microtiter plates. Saturation studies were conducted using [^3H]GR113808 at 10-12 different concentrations ranging from 0.005-2.5 nM. Displacement studies were performed using 0.2-0.4 nM [^3H]GR113808. The binding profile of drugs in competition experiments was established using 10-12 concentrations of compound. Incubation times were 30 min for both saturation and displacement studies. Nonspecific binding was defined in the presence of 50 μM 5-HT. Binding was initiated and terminated as described for [^3H]5-HT binding (see above). Radioactivity was measured and data were analyzed as described above for [^3H]5-HT.

Measurement of cAMP Formation: The transiently transfected Cos-7 cells were incubated in Dulbecco's modified Eagle's medium, 5 mM theophylline, 10 mM Hepes (4-[2-Hydroxyethyl]-1-piperazineethanesulfonic acid), 10 μM pargyline, and/or appropriate concentrations of antagonists for 20 minutes at 37°C, 5% CO_2 . Serotonin or other agonists in the presence or absence of forskolin (FSK) (10 μM) were then added at appropriate concentrations and incubated for an additional 10

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minutes at 37°C, 5% CO₂. The media was aspirated and the reaction stopped by the addition of 100 mM HCl. The plates were stored at 4°C for 15 minutes, centrifuged for 5 minutes, 500 x g to pellet cellular debris, and the supernatant aliquotted and stored at -20°C prior to assessment of cAMP formation by radioimmunoassay (cAMP Radioimmunoassay kit, Advanced Magnetics, Cambridge, MA). Radioactivity was quantitated using a Packard COBRA Auto Gamma Counter equipped with data reduction software. Functional data was fitted to a four parameter logistic equation to obtain response parameters (EC₅₀, E_{max}, nH; Inplot, GraphPad, San Diego, CA).

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Drugs: [³H]5-HT (specific activity = 22.7 Ci/mmol) was obtained from New England Nuclear, Boston, MA. [³H]GR113808 (specific activity = 82 Ci/mmol) was obtained from Amersham International (Arlington Hts., IL). All other chemicals were obtained from commercial sources and were of the highest grade purity available.

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Experimental Results

A 270 bp DNA fragment (S10) was identified when rat brain cDNA was used as template in a PCR amplification with two degenerate oligonucleotide primers derived from well conserved regions among several serotonin receptors, in the third and fifth putative transmembrane domains. The peptide sequence corresponding to this S10 PCR clone contained a "transmembrane IV like" domain. Since we used "serotonin receptor specific" PCR primers, this S10 clone represented a potentially new serotonin receptor. The corresponding full length cDNA was isolated from a rat brain cDNA library. Since five amplified commercial phage cDNA libraries turned out to be negative, we split the plasmid cDNA library into small pools of 2.5 to 5 x 10⁴ independent clones before amplification to avoid a potential growth bias against the S10 cDNA clone. By direct PCR analysis of bacterial pools, subsequent sib selection and standard filter hybridization, two cDNA clones were identified, S10-87 (5.5 kb) and S10-95 (4.5 kb). The nucleotide and deduced amino acid sequences are shown in Figure 1 (S10-87) and Figure 2 (S10-95). Surprisingly the peptide sequences between those two clones are only 96.7% identical, diverging in the second half of the carboxy terminus tails, downstream of position 359 (Figure 3). In addition, the entire 3' untranslated regions are totally divergent. The longest open reading frame for S10-87 encodes a protein of 387 amino acids and 406 amino acids for S10-95. The hydrophobicity plot displayed seven hydrophobic, putative membrane spanning regions which when compared to other G protein-coupled receptors did not show any significant high homologies, even to other serotonin receptors (Table 1 and figure 4). It is interesting to note that the highest homology, overall or restricted

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to the 7 TM region, is exhibited by the human histamine H₂ receptor, which like the 5-HT₁ receptor, is coupled to stimulation of cAMP.

5 Both S10-87 and S10-95 proteins carry 4 potential N-glycosylation sites in positions 7, 180, 316, and 352. They also possess 3 potential phosphorylation sites for protein kinase C in positions 218, 248, 318 and 4 potential phosphorylation sites for casein kinase II in
10 positions 9, 97, 218 and 288. A potential palmitoylation site is present in both clones at the cysteine found in position 329. A large number of G protein-coupled receptors carry a cysteine in the same position and O'Dowd et al. have speculated that it
15 plays an important role in the functional coupling of the human β_2 -adrenergic receptor. In addition, clone S10-95 carries one more potential phosphorylation site for protein kinase C at position 400. This additional phosphorylation site could lead to differential
20 functional coupling between the S10-87 and S10-95 receptors.

Since we isolated two different S10 cDNA clones by screening a library made from an entire brain, we
25 checked for differential expression in seven different parts of the brain by PCR amplification using pairs of primers specific for each clone. The results are summarized in table 2. Clone S10-95 seems to be transcribed everywhere in the rat brain except in
30 cerebellum. Clone S10-87 is only expressed in striatum. It remains to be determined if only one or both receptors are expressed in rat cortex.

35 The partial human S10-87 nucleotide (Fig 11A) and deduced amino acid sequences (Fig 11B) are shown. The sequences are highly similar to the rat S10-87 homolog, 90.8 % at the nucleotide level and 93.8 % at the amino

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acid level (figures 12 and 13 respectively).

5 The full length human S10-95 nucleotide (Fig 14A) and deduced amino acid sequences (Fig 14B) are shown. Compared to the rat S10-95 sequence, it shows 90.7 % identity at the nucleotide level and 91.8 % identity at the amino acid level (figures 15 and 16 respectively). The human S10-95 nucleotide sequence contains one nucleotide insertion in position 1159. 10 This insertion creates a frame shift and introduces a stop codon in the reading frame 16 nucleotides downstream. The protein motifs are highly conserved between the rat and human homologs except for a casein kinase II potential phosphorylation site in position 15 288 which is lost in both human receptors. The human homologs both carry a potential cAMP/cGMP phosphorylation site in position 338 in their carboxy terminal tail which is absent in the rat homologs. A comparison of the amino acid sequence between the human 20 and the rat S10-95 clones beginning from the initiating methionine and ending with the stop codon of the human S10-95 clone, reveals 31 amino acid changes of which 11 are non conservative, including 2 in TM1, 1 in TM2 and 1 in TM4. Due to the nucleotide insertion and the 25 corresponding frame shift described above, the carboxy terminal tail of the human S10-95 receptor is 16 amino acid shorter than its rat homolog.

30 Identical to the rat homologs, both human clones are identical in the loops and transmembrane regions, differing only in the second half of their carboxy terminal tail (fig 17, nucleic acid sequence; fig 18, aa sequence).

35 The human PCR cDNA fragments (TM-3 to TM-7) are 100% identical between heart, brain and retina. The nucleotide and deduced amino acid sequences are shown

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in figure 5. The human s quence shows 90.7% homology with the rat nucleotide sequence (Figure 6) and 92.3% homology (Figur 7) with the rat amino acid sequenc .

5 The genes encoding the rat S10-87 and S10-95 receptors were transiently expressed in Cos-7 cells for pharmacological evaluation. Initial experiments using 5 nM [³H]5-HT indicated that both S10-87 and S10-95 were serotonergic sites as demonstrated by the degree
10 of specific binding and density of sites expressed in the transfected cells when compared against the mock transfected controls. Saturation analysis of S10-87 (CG-7) was performed using 10 concentrations of [³H]5-HT (1-100 nM) and yielded a Bmax of 1,938 ± 399 fmol/
15 mg of protein and a K_d for [³H]5-HT of 7.87 ± 0.06 nM. The degree of specific binding at concentrations of [³H]5-HT close to the K_d ranged from 70-84% throughout the experimental series (including saturation and competition studies). Although the use of [³H]5-HT as
20 a radioligand to label 5-HT₁ receptors in brain tissue is not efficient due to the nonselectivity of the ligand, it became clear in the present studies using a homogeneous receptor population that [³H]5-HT would label this particular receptor. In fact, [³H]5-HT
25 appears to be labelling the high affinity state of the 5-HT₁ receptor which is not unusual for the conditions upon which this receptor has been studied. Similar results using an agonist radioligand have been previously reported for the cloned 5-HT₂ receptor
30 (Branchek et al., 1990).

A pharmacological binding profile of S10-87 and S10-95 (CG-7 and CG-8) was performed and demonstrated that
35 this novel receptor was similar to the 5-HT₁ receptor as defined by functional assays in the literature (Bockart et al., 1992). This is clearly shown in table 3 where the binding affinities of various

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serotonergic agents are displayed for S10. Notably, 5-HT and the tryptamine derivative 5-methoxytryptamine possessed high affinity. Furthermore, as previously reported for the 5-HT₁ receptor, benzamide derivatives including cisapride, BRL 24924 and zacopride bound with fairly high affinity to receptors expressed from the S10 gene. ICS 205930, a tropanyl-indole derivative, which has been reported to be an antagonist at both 5-HT₁ and 5-HT₂ receptors (Bockaert et al., 1992), also bound to these receptor sites. Compounds such as 8-hydroxy-2-(di-n-propylamino)tetralin, ketanserin, sumatriptan and 5-carboxyamidotryptamine were of low affinity having K_i values estimated to be greater than 1 μ M. This data would rule out S10 from belonging to other serotonergic receptor subfamilies such as 5-HT₁ and 5-HT₂. Taken together, the complete pharmacological profile also differentiates S10 from the related subtype 5-HT₄ (U.S. Serial No., 971,960, filed, November 3, 1992, copending). Although some of the drugs tested also have good affinity for 5-HT₃ receptors, S10 is clearly a 5-HT₁ receptor based upon the binding data and the functional data demonstrating a positive-coupling to adenylate cyclase. Finally, a correlation plot for the binding affinities of 5-HT, cisapride, BRL 24924, zacopride, and ICS 205930 against their functional responses in adenylate cyclase assays from cultured mouse collicular neurons (Dumuis et al., 1989) yielded a correlation coefficient of 0.96 (Figure 8). Thus, the rank order of potency for these key compounds also provides conclusive evidence that S10 encodes a 5-HT₁ receptor.

To examine the ability of S10 clone to couple to adenylate cyclase, Cos-7 cells transiently expressing S10 were tested for the ability to exhibit an increase in basal cAMP release or a decrease in FSK-stimulated cAMP response. 5-HT (1 μ M) had no effect on either

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basal or FSK-stimulated adenylate cyclase activity in untransfected or mock-transfected Cos-7 cells (data not shown), indicating that endogenous α -coupled serotonin receptors are not present in untransfected cells. Preliminary studies were carried out by adding only one dose of various drugs in triplicate. Addition of 5-HT ($1 \mu\text{M}$) to this system caused stimulation of basal cAMP release (CG-7 = 0.020 ± 0.002 ; CG-8 = $0.023 \pm 0.004 \text{ pmol/ml/10min}$) by about 30 fold for each clone; no inhibition of either the basal or FSK-stimulated cAMP release was observed. On the contrary, addition of $10 \mu\text{M}$ FSK together with $1 \mu\text{M}$ 5-HT stimulated cAMP accumulation about 10-fold more than either agent alone (data not shown). For various compounds, full dose-response curves were determined for both clones and the data are summarized in table 4. 5-HT caused a concentration-dependent stimulation of basal adenylate cyclase activity with mean EC_{50} s of 26 ± 3 and $51 \pm 7 \text{ nM}$ and E_{max} s of 2,107 and 2,598 % basal cAMP release for CG-7 and CG-8 respectively (Figures 9 and 10). Among the tryptamine derivatives tested, 5-MeOT was approximately equipotent with 5-HT in both clones, whereas α -Me-5-HT and 5-CT were about 10 and 200 times respectively less potent than 5-HT at CG-7. On the other hand, the latter two compounds displayed approximately 20 and 30 fold lower affinity than 5-HT respectively for CG-8. The 2-methoxy-4-amino-5-chloro-substituted benzamides (cisapride, BRL-249245 and zacopride) were less potent agonists than 5-HT in stimulating basal cAMP release and displayed different rank order of potency for CG-7 and CG-8. As indicated in table 4 using CG-7, cisapride, BRL-24924 and zacopride exhibited approximately 10, 30 and 100 fold lower potency than 5-HT respectively, whereas at CG-8 these compounds had almost equal affinity. Thus, although not different in binding properties, these subtle differences in affinity in functional assays f

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the two "variants" (CG-7 and CG-8) indicate the potential to develop separate therapeutic entities for each separate target. All the agonists tested acted as full agonists with the exception of cisapride, BRL-24924 and zacopride, which also displayed antagonist activity and were therefore partial agonists at both clones, with intrinsic activities ranging between 0.85 and 1.4 (Table 4). ICS-205-930 (100 μ M) had similar effect at the two clones and was found to be a silent antagonist causing parallel dextral shifts in the concentration effect curve of 5-HT without altering the maximum response significantly. The estimated K_d value for ICS-205-930 was not significantly different between the two clones (CG-7 = 962 ± 244 nM; CG-8 = 607 ± 30 nM). Responses to 5-HT were not affected by spiperone or methiothepin (10 μ M) in either clone.

Saturation analysis of rat 5-HT_{4A} S10-87 (CG-7) and S10-95 (CG-8) clones and human 5-HT_{4A} clone CG-17 were performed using 10-12 concentrations of [³H]GR113808 (0.005-2.5 nM) and revealed a single saturable site of high affinity for both clones (CG-7: $K_d = 0.74$ nM, $B_{max} = 5.7$ pmol/mg membrane protein; CG-8: $K_d = 1.0$ nM, $B_{max} = 3.7$ pmol/mg membrane protein; CG-17: $K_d = 0.20$ nM, $B_{max} = 1.8$ pmol/mg membrane protein). These preliminary data indicate that although the rat clones (CG-7 and CG-8) have similar affinity for the antagonist [³H]GR113808, the human clone (CG-17) displays approximately 5-fold higher affinity than the rat clones for this ligand. For all three clones nonspecific binding increased linearly with increasing ligand concentration. The degree of specific binding at concentrations of [³H]GR113808 (0.4-0.5 nM) ranged from 80-90%.

The pharmacological binding profile of S10-87 and S10-95 (CG-7, CG-8, respectively) was investigated in

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displacement studies using [^3H]GR113808 and/or [^3H]5-HT. In order to compare CG-17 pharmacology with that previously obtained for the rat clones, CG-7 and CG-8, displacement studies on the human CG-17 clone were performed using [^3H]5-HT as the radioligand.

A range of 5-HT₂ receptor agonists and antagonists completely inhibited the specific binding of [^3H]GR113808 on both the rat CG-7 and CG-8 clones. Affinity values and Hill slopes derived from the curves using computer analysis are presented in Table 5. As previously observed using [^3H]5-HT as the radioligand, the rat CG-7 and CG-8 receptors had very similar pharmacology. Of the agonists tested, only those active in 5-HT₂ containing preparations (5-HT and 5-MeOT) potently inhibited [^3H]GR113808. By contrast, agonists for other 5-HT receptors, for example 5-HT_{1A} receptor agonist, 8-OH-DPAT, the 5-HT₁₀ receptor agonist, sumatriptan, the 5-HT₂ receptor antagonist, ketanserin, had no effect on [^3H]GR113808 binding at concentrations up to 1 μM . The substituted benzamides, cisapride, BRL-24924 and zacopride, partial agonists at 5-HT₂ receptor all potently inhibited [^3H]GR113808 binding. Specific [^3H]GR113808 binding was also inhibited by the 5-HT₂ receptor antagonist ICS-205930.

For both the rat CG-7 and CG-8 clones, Hill slopes for the inhibition of [^3H]GR113808 binding by 5-HT₂ receptor agonists but not the antagonist, ICS-205930, were shallow in the absence of Gpp(NH) with the exception of 5-CT, and α -Me-5-HT. For agonists that had shallow displacement curves, the binding was resolved into high and low affinity components and these are summarized in Table 5. The K_i values obtained for the high affinity state of the receptor using [^3H]GR113808 as the radioligand were compatible with the K_i values obtained previously using [^3H]5-HT as the radioligand which

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labels the high affinity state of the receptor (Table 5 and 6). Some differences were observed for the K_i values of high affinity state of CG-7 compared to CG-8 (Table 3) and their nH values. For example, although there were no differences in the K_i values of CG-7 and CG-8, the displacement curve obtained for 5-MeOT using CG-8 clone could not be resolved into two sites. Also the K_i value obtained for the high affinity state of CG-8 using cisapride was approximately 3-fold lower than that obtained for CG-7 previously using [3H]5-HT to directly label the high affinity state of the receptor. We are currently investigating these differences using [3H]5-HT to directly label the high affinity state.

In the presence of 100 μM Gpp(NH)p, competition binding curves for the agonists displaying shallow curves in the absence of Gpp(NH)p were shifted to the right and this is exemplified for 5-HT in Fig. The Hill slopes were increased.

Preliminary results obtained with the human clone (CG-17) using [3H]5-HT as the radioligand in displacement studies are summarized in Table 3. Similar to the rat CG-7 and CG-8 clones, 8-OH-DPAT, sumatriptan and ketanserin were inactive at the CG-17 clone for up to concentration of 1 μM . The differences observed between the human and the rat CG-8 clones were as follows. The biggest difference was observed with α -Me-5-HT which had approximately 100 fold higher affinity for the human CG-17 clone. Zacopride, 5-MeOT and cisapride had about 7-fold, 5-fold and 4-fold higher affinity, respectively for the human clone CG-17.

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DISCUSSION

W have identified two cDNA clones encoding the pharmacologically-defined 5-HT₁ receptor. This receptor is expressed at low levels in rat brain if we consider its frequency in the cDNA library ($\leq 1:10^6$). Surprisingly, two receptors differing in their carboxy-terminus regions have been isolated. Since the 3' untranslated nucleotide sequences are also different, these two receptors could be encoded by two different genes or could arise by alternative splicing of pre-mRNA. These two receptors (S10-87 and S10-95) are differentially expressed in rat brain and the physiological relevance of the S10-87 receptor being expressed only in striatum remains to be determined.

The pharmacology binding profile and the functional coupling obtained from cells expressing S10 clones indicate that these genes both encode a pharmacologically-defined 5-HT₁ receptor. The cloned rat CG-7 and CG-8 genes transiently expressed in Cos-7 cells coupled to stimulation of adenylate cyclase. The magnitude of this response (~20-25 fold) was large. With the exception of 5-MeOT, agonist potencies determined from functional assays were less than expected from K_i values obtained from binding assays using [³H]5-HT. Could this result be due to the possibility that the dose of [³H]5-HT used in binding assays measures only the high affinity site of agonists? This is not likely, as it would not account for the data obtained with ICS-205-930 which is a silent antagonist in the present system displaying approximately 6 (CG-8) and 10 (CG-7) fold lower affinity in the functional assay as compared to the binding experiments. It is more likely that differences in experimental conditions used in binding assays compared with those used in the functional

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assays (such as membrane vs. cells, buffers and extent of equilibrium achieved) accounts for these differences.

5 5-HT responses were resistant to blockade by methiothepin and spiperone (10 μ M). As the concentration of these agents exceed their equilibrium dissociation constants for their respective receptor sites by 10-100 fold, it seems that 5-HT₁-like (5-HT_{1A},
10 5-HT_{1B}, 5-HT_{1C}, 5-HT_{1D}, 5-HT_{1E}, 5-HT_{1F}), 5-HT₂ and 5-HT₄ receptors can be ruled out. In addition, the weak agonistic activity of 5-CT relative to 5-HT further supports the notion that 5-HT₁-like receptors are probably not involved (Bradley, 1986). The results
15 obtained with the indole agonists reflect those reported at the 5-HT₂ receptor in both the CNS and the periphery (Dumuis et al., 1988; Craig and Clarke, 1990; Eglen et al., 1990; Baxter, Craig and Clarke, 1991). The substituted benzamides, cisapride, BRL-24924 and
20 zacopride acted as partial agonists. Although the benzamides also possess affinity for 5-HT₃ receptors, they lack intrinsic efficacy (Schuurkes et al., 1985; Sanger and King, 1988). Furthermore, the affinity of ICS-205-930 for antagonism of 5-HT response at S10 is
25 1-3 orders of magnitude lower than that at 5-HT₃ receptors (Richardson et al., 1985) and therefore indicates a binding site different from 5-HT₃ receptor.

As was the case with the rat 5-HT₂ receptor, there are
30 two forms of the human homolog, most likely splice variants differing in the carboxyterminal end of the receptor. Non conservative amino acid substitutions, especially in transmembrane domains 1, 2 and 4 could provide the basis for the pharmacological differences
35 observed between the rat and the human 5-HT₂ receptors (see below). A difference in the functional response is observed between the two rat clones: although the

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CG-7 construct (S10-87) gives higher levels of receptor expression in COS-7 cells (Bmax of 5.7pmol/mg of protein versus 3.7 pmol/mg for S10-95), in the functional assay, the CG-8 construct (S10-95) reproducibly shows a higher level of cAMP stimulation (2598 \pm 154% of basal cAMP release versus 2107 \pm 52% for CG-7). This finding could be attributed to the different amino acid sequence in the carboxy terminal tail of the rat receptors, specially since the rat S10-95 isoform carries an additional potential phosphorylation site at position 400, absent in S10-87 (CG-7).

Since the human S10-95 homolog lacks the last 16 carboxy terminal amino acids which carry the phosphorylation site mentioned above in the rat homolog, it will be interesting to check for differences in the level of cAMP stimulation upon activation of the rat and human S10-95 homologs. In the same way, after we get the full length human S10-87, both human isoforms will be compared in binding and functional assays.

All the unique pharmacological characteristics described above define the S10 genes as adenylate cyclase stimulatory "5-HT₄" receptors reported in the literature that are expressed in the ileum (Craig and Clarke, 1990), hippocampus (Shenker et al., 1987), esophagus (Baxter et al., 1991), embryo colliculi neurons (Dumius et al., 1988), atrium (Kaumann et al., 1990), adrenal (Lefebvre et al., 1992) and bladder (Corsi et al., 1991), and distinguish these clones from all other cloned subtypes of 5-HT receptor. Although the binding profile of CG-7 and CG-8 were identical (Table 3), some differences in agonist potency (benzamides in particular) were observed between them in the functional assays. This is not surprising since

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the amino acid sequences of these two clones are identical, apart from the cytoplasmic carboxy tail, a region that is important for G protein-coupling, where the CG-8 receptor carries an extra phosphorylation site. Cisapride, BRL-24924 and zacopride had similar affinities at CG-8 whereas BRL-24924 and zacopride displayed approximately 4 and 15 fold lower affinity than cisapride at CG-7 clone for stimulation of adenylate cyclase. It is noteworthy that tissue differences in the potency of benzamides have been reported (Baxter et al., 1991) and whether this reflects a heterogeneity of 5-HT₄ receptors remains to be investigated.

The data obtained with the rat CG-7 and CG-8 clones and the human CG-17 clone using [³H]GR113808 are very similar to those reported by Grossmann et al. (1993) and Waeber et al. (1993) with this ligand using guinea pig and human brain tissues. Specific [³H]GR113808 binding readily saturated for all three clones (CG-7, CG-8 and CG-17). Scatchard analysis of specific binding in all three clones revealed the involvement of a single site. Curve fitting analysis showed an equilibrium dissociation of approximately 1 nM for both rat clones (CG-7 and CG-8) whereas this value was about 5-fold lower for the human CG-17 clone ($K_d = 0.2$ nM). The K_d value obtained for the human CG-17 clone is in excellent agreement with that reported by Waeber et al. (1993) using human brain (0.23-0.37 nM) and is very similar to that of the guinea pig brain tissue (0.13-0.2 nM; Waeber et al., 1993; Grossmann et al., 1993). The K_d value of [³H]GR113808 for rat brain tissue has not been reported, however, it is interesting that the affinity obtained for GR113808 from functional receptor assays in the rat oesophagus (Grossmann et al., 1993) is about 0.3 nM which indicates that this antagonist has similar affinities for the human and the rat tissue

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used. The discrepancies between our data and those reported may be due to methodology, or different subtype (brain vs. oesophagus), however, this remains to be investigated.

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The rank order of potency of compounds competing for specific [³H]GR113808 are very similar for both CG-7 and CG-8 and is cisapride> 5-HT> BRL-24924> 5-MeOT= ICS205930> zacopride> α-Me-5-HT> 5-CT. This order of potency is different from that observed with guinea pig caudate (Grossmann et al., 1993; cisapride> 5-HT> ICS205930> BRL-24924> zacopride> 5-MeOT> α-Me-5-HT> 5-CT) and human caudate (Waeber et al., 1993; cisapride> ICS205930> BRL-24924> 5-HT> 5-MeOT. Whether these differences are due to species or different population of high affinity state of the receptor in the various preparation, remains to be investigated. Interestingly, the displacement curves for most of the agonists competing for specific [³H]GR113808 were shallow and could be resolved to high and low affinity states. Gpp(NH)p shifted these curves to the right and in the case of 5-HT the Hill coefficient was increased to unity; however for some agonists the shift was not complete. Grossmann et al.(1993) using guinea pig caudate also observed shallow competition curves for some agonist that could be partially shifted by the addition of GTP (Grossmann et al., 1993). However, Waeber did not observe shallow binding curves with the human caudate tissue. These observations indicate that the G protein content of these preparations may be different which may reflect differences in the coupling of the receptor with a G protein.

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Using [³H]5-HT as the radioligand, the affinity values for the human CG-17 clone are in general comparable with that obtained by us previously for the rat CG-7 and CG-8 clones with few exceptions. The most striking

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differences appears to be for α -Me-5-HT which displays approximately 100 fold higher affinity for the human CG-17. However, it has to be pointed out that the data are compared with 2 different radioligands and this difference has to be further investigated using the same radioligand for all both the rat and human clones in parallel. 5-MeOT was approximately 3 fold less potent whereas zacopride was about 7 fold more potent at the human CG-17 as compared to the rat clones.

The cloning and expression of genes encoding 5-HT₄ receptors allows, for the first time, the ability to develop subtype selective drugs using radioligand binding assays. It will further provide definitive answers to whether there are significant species differences in the pharmacology of 5-HT₄ receptors. In addition, the intrinsic activity of drugs can be determined from measures of adenylate cyclase activation in these transfected cells. So far, native tissue preparations have shown great disparity in agonist activity. 5-HT₄ receptors have been implicated in a wide variety of functions. Existing drugs such as metaclopramide and cisapride appear to exert a large part of their action through 5-HT₄ receptors (Taniyama et al., 1991; Meulemans and Schurkes, 1992; Flynn et al., 1992). Experience with these agents indicates a clear therapeutic role for 5-HT₄ receptors in the gastrointestinal system for conditions including irritable bowel disease, postoperative ileus, diabetic gastroparesis, emesis, achalasia, hiatal hernia, and esophageal spasm. In addition, 5-HT₄ receptors have been described functionally in the heart (Kaumann, 1992), adrenal (Lefebvre et al., 1992), and bladder (Corsi et al., 1991) indicating possible roles in cardiac rate and force of contraction, endocrine control of cortisol secretion, and urinary incontinence or spasticity. 5-HT₄ receptors have also been

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described in the brain, particularly in areas such as the hippocampus, in which we have localized the gene encoding 5-HT₁ receptors (S10-87), indicating a potential role in cognitive enhancement (Bockaert et al., 1992). As more specific pharmacological tools are developed, additional therapeutic indications will certainly be uncovered.

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TABLE 2

Primers	Corte x	Cerebell um	Brain stem	Hippocamp us	Olfactory Bulb	Striatu m	Thalam us
TM3-5	+	-	+	+	+	+	+
S10-87	ND	-	-	-	-	+	-
S10-95	ND	-	+	+	+	+	+

Table 2: PCR localization of the S10 mRNA in 7 different part of the rat brain.

The TM3-5 primers do not differentiate between clones S10-87 and S10-95.

The S10-87 primers were designed from the nucleotide sequences coding for the TM 6 domain common to both receptors and for the carboxy terminus end specific to S10-87. In the same way, the S10-95 primers are specific for S10-95.

Table 3. Binding affinities of key ligands for the identification of S10 (CG-7 and CG-8) as a 5-HT₄ receptor. Affinity constants (K_i; nM) of drugs competing for S10 label d with 10 nM [³H]5-HT were determined to pharmacologically define the encoded receptor as 5-HT₄. K_i (nM) values were calculated using the Cheng-Prusoff equation or estimated to be >1000 nM based upon one point displacements using a drug concentration of 1 uM. Affinity constants are expressed as the mean \pm SEM (n \geq 2). K_i values estimated to be >1000 were determined according to one point displacements studies at a concentration of 1 uM. (n=2 except BRL 24924 tested at CG-8: n=1)

CHARACTERIZATION OF CLONE S-10

Saturation Analysis: K_d = 7.87 ± 0.06 nM

B_{max} = $1,938$ fmol/ mg prot

Pharmacological profile:

DRUG	CG-7	CG-8
5-HT	8.6 ± 0.6	6.4 ± 0.5
Cisapride	10.9 ± 0.3	ND
5-MeOT	27.5 ± 5	ND
BRL 24924	27.7 ± 5	21.1^*
ICS 205930	115 ± 12	138 ± 26
Zacopride	130 ± 10	136 ± 5
8OHDPAT	>1000	ND
Ketanserin	>1000	ND
Sumatriptan	>1000	ND
5-CT	>1000	ND

* n = 1

ND = not determined

Table 4. Pharmacological profile for the cAMP response using the human 5-HT_{1A} (CG-7 and CG-8) receptor transiently expressed in Cos-7 cells, comparison with the binding data obtained with CG-7 clone using [³H]5-HT. cAMP measurements on intact cells were as described under Methods and Materials. EC₅₀ values (concentration producing the half-maximal effect) were derived from the analysis of full dose-response curves. Maximum response produced by each drug was normalized to the 5-HT maximum response which is indicated as having an intrinsic activity of 1.0. Data are means ± S.E.M. of three separate experiments. The apparent dissociation constant of antagonist (K_B) (ICS-205930) was calculated according to the formula: $K_B = [B]/(A'/A - 1)$, where [B] is the concentration of antagonist, A' and A the EC₅₀ values of agonist measured respectively in the presence and in the absence of antagonist (Furchgott, 1972).

DRUG	EC ₅₀ or K _B (nM)		I. A. °		K _i (nM)	K _i (nM)
	CG-7	CG-8	CG-7	CG-8	CG-7	CG-8
5-MeOT	21 ± 6	31 ± 13	1.0	1.0	27 ± 5	ND
5-HT	26 ± 3	51 ± 7	1.0	1.0	8.6 ± 0.6	6.4 ± 0.5
Cisapride	191 ± 26	413 ± 199	1.2	1.4	11 ± 0.3	ND
α-Me-5-HT	250 ± 91	1,038 ± 31	0.90	1.0	ND	ND
BRL-24924	736 ± 129	250 ± 25	1.1	0.9	28 ± 5	21
Zacopride	2,740 ± 274	239 ± 33	1.1	1.0	130 ± 10	136 ± 5
5-CT	5,570 ± 808	1,411 ± 211	0.85	1.2	>1,000	ND
ICS-205930	962 ± 244	607 ± 30	0	0	115 ± 12	138 ± 26

ND, not determined.

Maximum response to 5-HT in Cos-7 cells transiently transfected with CG-7 and CG-8 genes was:

CG-7 = 2,107 ± 52 % of basal cAMP release

CG-8 = 2,598 ± 154 % of basal cAMP release

Cisapride, BRL-24924 and zacopride had no antagonist activity and ICS-205930 had no intrinsic agonist activity.

Table 5. The affinities of various compounds that compete for 0.2-0.4 nM [³H]GR113808 binding in membranes of COS-7 cells transiently transfected with rat clones CG-7 and CG-8.

COMPOUNDS	CG-7	CG-8
5-HT	237 $K_d=2.6$, $BH=22\%$ $K_i=357$ $nH=0.62$	116 $KH=2.5$, $BH=24\%$ $KL=197$ $nH=0.67$
5-CT	>10,000	>10,000
5-Me-OT	438 $KH=14$, $BH=17\%$ $KL=658$ $nH=0.57$	518 $KH=$, $BH=$ $KL=$ $nH=0.66$
BRL-24924	189 $KH=37$, $BH=34\%$ $KL=371$ $nH=0.85$	188 $KH=23$, $BH=32\%$ $KL=373$ $nH=0.81$
ZACOPRIDE	729 $KH=424$, $BH=68\%$ $KL=2,757$ $nH=0.80$	820 $KH=$, $BH=$ $KL=$ $nH=0.85$
d-LSD	>10,000	>10,000
CISAPRIDE	ND	84 $KH=2.7$, $BH=11\%$ $KL=105$ $nH=0.83$
ICS205930	ND	529
α -Me-5-HT	2,255	1,855

Affinity estimates are given as K_i values (nM) using the computer program "Accucomp" (Lundon S ftware). Values are from a single experiment. K_d and K_i values indicate the affinity in nM for the high affinity and low affinity state of the receptor, B_H is the percentage of high affinity sites and nH is the Hill coefficient.
ND, not determined.

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Tabl 6. Binding affinities of key ligands for the identification of the human CG-17 clone as a 5-HT_{4A} receptor.

COMPOUND	K _i (nM)
5-HT	4.2
5-MeOT	71
5-CT	>10,000
Cisapride	12
α -Me-5-HT	1.6
BRL-24924	21
Zacopride	17
Sumatriptan	>1,000
8-OH-DPAT	>1,000
Ketanserin	>1,000

Affinity constants (K_i, nM) of drugs competing for CG-17 cloned labeled with 10 nM [³H]5-HT were calculated using the Cheng-Prusoff equation or estimated to be greater than 1,000 nM based upon one point displacement using a drug concentration of 1 μ M. Values are from a single experiment.

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25 (1992).

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: SYNAPTIC PHARMACEUTICAL CORPORATION

(ii) TITLE OF INVENTION: DMA ENCODING 5-HT4 SEROTONIN RECEPTORS
AND USES THEREOF

(iii) NUMBER OF SEQUENCES: 15

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: COOPER & DUNHAM
(B) STREET: 30 ROCKEFELLER PLAZA
(C) CITY: NEW YORK
(D) STATE: NEW YORK
(F) ZIP: 10112

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.24

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(vii) ATTORNEY/AGENT INFORMATION:

(A) NAME: White, P., John
(B) REGISTRATION NUMBER: 28,678
(C) REFERENCE/DOCKET NUMBER: 42667-A-PCT/JPW/TEP

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (212) 977-9550
(B) TELEFAX: (212) 664-0525
(C) TELEX: 422523 COOP UI

(2) INFORMATION FOR SEQ ID NO:1:

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- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1642 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: N
- (iv) ANTI-SENSE: N
- (vi) ORIGINAL SOURCE:
 (F) TISSUE TYPE: brain
- (vii) IMMEDIATE SOURCE:
 (A) LIBRARY: rat brain
 (B) CLONE: S10-87
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 101..1261
 (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:																
AGCCTTGCCG	AGCCTGGCTT	GGTTGGAAGG	AGGAGGATGC	TCTGCGTGCC	CAGGGTCTCG											60
TGGGCACTGA	CATCCAACGT	ACTCATGCCC	ATTTCTGTGA	ATG GAC AGA CTT GAT												115
				Met Asp Arg Leu Asp												
				1	5											
GCT AAT GTG AGT TCC AAC GAG GGT TTC GGG TCT GTG GAG AAG GTC GTA																163
Ala Asn Val Ser Ser Asn Glu Gly Phe Gly Ser Val Glu Lys Val Val																
				10				15							20	
CTG CTC ACG TTC TTC GCA ATG GTT ATC CTG ATG GCC ATC CTG GCC AAC																211
Leu Leu Thr Phe Phe Ala Met Val Ile Leu Met Ala Ile Leu Gly Asn																
				25				30						35		
CTG CTG GTG ATG GTT GCT GTG TGC AGG GAC AGG CAG CTC AGG AAA ATA																259
Leu Leu Val Met Val Ala Val Cys Arg Asp Arg Gln Leu Arg Lys Ile																
				40				45						50		
AAA ACC AAT TAT TTC ATT GTG TCT CTT GCC TTT GCT GAT CTG CTG GTT																307

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Lys	Thr	Asn	Tyr	Phe	Ile	Val	Ser	Leu	Ala	Phe	Ala	Asp	Leu	Leu	Val	
	55					60					65					
TCG	GTG	CTG	GTG	AAT	GCC	TTC	GGT	GCC	ATT	GAG	TTG	GTC	CAA	GAC	ATC	355
Ser	Val	Leu	Val	Asn	Ala	Phe	Gly	Ala	Ile	Glu	Leu	Val	Gln	Asp	Ile	
70					75					80					85	
TGG	TTT	TAT	GGG	GAG	ATG	TTT	TGC	CTG	GTC	CGG	ACC	TCT	CTG	GAT	GTC	403
Trp	Phe	Tyr	Gly	Glu	Met	Phe	Cys	Leu	Val	Arg	Thr	Ser	Leu	Asp	Val	
				90					95					100		
CTA	CTC	ACC	ACA	GCA	TCA	ATT	TTT	CAC	CTC	TGC	TGC	ATT	TCC	CTG	GAT	451
Leu	Leu	Thr	Thr	Ala	Ser	Ile	Phe	His	Leu	Cys	Cys	Ile	Ser	Leu	Asp	
			105					110					115			
AGG	TAT	TAT	GCC	ATC	TGC	TGT	CAA	CCT	TTG	GTT	TAT	AGA	AAC	AAG	ATG	499
Arg	Tyr	Tyr	Ala	Ile	Cys	Cys	Gln	Pro	Leu	Val	Tyr	Arg	Asn	Lys	Met	
		120					125					130				
ACC	CCT	CTA	CGC	ATC	GCA	TTA	ATG	CTG	GGA	GGC	TGC	TGG	GTC	ATT	CCC	547
Thr	Pro	Leu	Arg	Ile	Ala	Leu	Met	Leu	Gly	Gly	Cys	Trp	Val	Ile	Pro	
	135					140					145					
ATG	TTT	ATA	TCT	TTT	CTC	CCC	ATA	ATG	CAA	GCC	TGG	AAC	AAC	ATC	GGC	595
Met	Phe	Ile	Ser	Phe	Leu	Pro	Ile	Met	Gln	Gly	Trp	Asn	Asn	Ile	Gly	
150					155					160					165	
ATA	GTT	GAT	GTG	ATA	GAG	AAA	AGG	AAA	TTC	AAC	CAC	AAC	TCT	AAC	TCT	643
Ile	Val	Asp	Val	Ile	Glu	Lys	Arg	Lys	Phe	Asn	His	Asn	Ser	Asn	Ser	
				170					175					180		
ACA	TTC	TGT	GTC	TTC	ATG	GTC	AAC	AAG	CCC	TAT	GCC	ATC	ACC	TGC	TCT	691
Thr	Phe	Cys	Val	Phe	Met	Val	Asn	Lys	Pro	Tyr	Ala	Ile	Thr	Cys	Ser	
			185				190						195			
GTG	GTG	GCC	TTC	TAC	ATC	CCG	TTT	CTC	CTC	ATG	GTG	CTG	GCC	TAT	TAC	739
Val	Val	Ala	Phe	Tyr	Ile	Pro	Phe	Leu	Leu	Met	Val	Leu	Ala	Tyr	Tyr	
		200				205						210				
CGT	ATC	TAT	GTC	ACT	GCT	AAG	GAG	CAT	GCC	CAG	CAG	ATC	CAG	ATG	TTA	787
Arg	Ile	Tyr	Val	Thr	Ala	Lys	Glu	His	Ala	Gln	Gln	Ile	Gln	Met	Leu	
	215					220					225					
CAA	CGG	GCA	GGA	GCC	ACC	TCT	GAA	AGC	AGG	CCC	CAG	ACA	GCT	GAC	CAG	835
Gln	Arg	Ala	Gly	Ala	Thr	Ser	Glu	Ser	Arg	Pro	Gln	Thr	Ala	Asp	Gln	

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230	235							240							245		
CAC	AGC	ACA	CAT	CGC	ATG	CGG	ACA	GAG	ACC	AAA	GCA	GCC	AAG	ACT	TTA		883
His	Ser	Thr	His	Arg	Met	Arg	Thr	Glu	Thr	Lys	Ala	Ala	Lys	Thr	Leu		
				250					255					260			
TGT	GTC	ATC	ATG	GGC	TGC	TTC	TGT	TTC	TGC	TGG	GCC	CCC	TTC	TTT	GTC		931
Cys	Val	Ile	Met	Gly	Cys	Phe	Cys	Phe	Cys	Trp	Ala	Pro	Phe	Phe	Val		
			265					270					275				
ACC	AAT	ATT	GTG	GAC	CCT	TTC	ATA	GAC	TAC	ACT	GTG	CCC	GAG	AAG	GTG		979
Thr	Asn	Ile	Val	Asp	Pro	Phe	Ile	Asp	Tyr	Thr	Val	Pro	Glu	Lys	Val		
		280					285					290					
TGG	ACT	GCT	TTC	CTC	TGG	CTT	GGC	TAT	ATC	AAT	TCA	GGG	TTG	AAC	CCT		1027
Trp	Thr	Ala	Phe	Leu	Trp	Leu	Gly	Tyr	Ile	Asn	Ser	Gly	Leu	Asn	Pro		
	295					300					305						
TTT	CTC	TAT	GCC	TTC	TTG	AAT	AAG	TCT	TTC	AGA	CGT	GCC	TTC	CTT	ATC		1075
Phe	Leu	Tyr	Ala	Phe	Leu	Asn	Lys	Ser	Phe	Arg	Arg	Ala	Phe	Leu	Ile		
310					315					320					325		
ATC	CTC	TGC	TGT	GAT	GAT	GAG	CGC	TAC	AAA	AGA	CCC	CCC	ATT	CTG	GGC		1123
Ile	Leu	Cys	Cys	Asp	Asp	Glu	Arg	Tyr	Lys	Arg	Pro	Pro	Ile	Leu	Gly		
				330					335					340			
CAG	ACT	GTC	CCC	TGT	TCA	ACC	ACA	ACC	ATT	AAT	GGA	TCC	ACT	CAT	GTG		1171
Gln	Thr	Val	Pro	Cys	Ser	Thr	Thr	Thr	Ile	Asn	Gly	Ser	Thr	His	Val		
			345					350					355				
CTA	AGG	TAT	ACA	GTT	TTG	CAT	AGT	GGT	CAA	CAC	CAG	GAA	CTG	GAG	AAG		1219
Leu	Arg	Tyr	Thr	Val	Leu	His	Ser	Gly	Gln	His	Gln	Glu	Leu	Glu	Lys		
		360				365					370						
TTG	CCC	ATA	CAC	AAT	GAC	CCA	GAG	TCC	CTG	GAA	TCA	TGC	TTT				1261
Leu	Pro	Ile	His	Asn	Asp	Pro	Glu	Ser	Leu	Glu	Ser	Cys	Phe				
	375					380					385						
TGATTGAAGA		CGTGGCTTGC		CTTTAGCGGT		TCATCCCATC		TGTGTCTGCA		TGAACAGGTT							1321
ACTATGGAAT		CACCTCTGAC		TCTGGGCATC		ACCAGTGAAG		CATGAGCATG		GTGAGGCAGG							1381
GTCCGGTGAA		GGTGACAGAG		GGACAGCATT		GAGTGGGACC		TGAACCCAGC		ACATTAAGGA							1441
TTTCAGAAC		GTGTGGGGAT		TTGAGATGTC		ATCAGACCCA		GTGTCTTACC		CAGAGCCCAA							1501

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CTGGCACCTC	CCATTCACCG	CTGACATGTG	GTCAGTCTTT	GCTCACACCT	CTCCAGGGGC	1561
AGGAGCTGAC	TACCTCCTAA	TGTGGTGGGG	AGCTCTTAAT	TGTGTGGAAG	TTCAGTCATT	1621
CATTGGTGGA	CAGTCTCGCT	G				1642

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 387 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met	Asp	Arg	Leu	Asp	Ala	Asn	Val	Ser	Ser	Asn	Glu	Gly	Phe	Gly	Ser	1	5	10	15
Val	Glu	Lys	Val	Val	Leu	Leu	Thr	Phe	Phe	Ala	Met	Val	Ile	Leu	Met	20	25	30	
Ala	Ile	Leu	Gly	Asn	Leu	Leu	Val	Met	Val	Ala	Val	Cys	Arg	Asp	Arg	35	40	45	
Gln	Leu	Arg	Lys	Ile	Lys	Thr	Asn	Tyr	Phe	Ile	Val	Ser	Leu	Ala	Phe	50	55	60	
Ala	Asp	Leu	Leu	Val	Ser	Val	Leu	Val	Asn	Ala	Phe	Gly	Ala	Ile	Glu	65	70	75	80
Leu	Val	Gln	Asp	Ile	Trp	Phe	Tyr	Gly	Glu	Met	Phe	Cys	Leu	Val	Arg	85	90	95	
Thr	Ser	Leu	Asp	Val	Leu	Leu	Thr	Thr	Ala	Ser	Ile	Phe	His	Leu	Cys	100	105	110	
Cys	Ile	Ser	Leu	Asp	Arg	Tyr	Tyr	Ala	Ile	Cys	Cys	Gln	Pro	Leu	Val	115	120	125	
Tyr	Arg	Asn	Lys	Met	Thr	Pro	Leu	Arg	Ile	Ala	Leu	Met	Leu	Gly	Gly	130	135	140	

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Cys	Trp	Val	Ile	Pro	Met	Phe	Ile	Ser	Phe	Leu	Pro	Ile	Met	Gln	Gly	145	150	155	160
Trp	Asn	Asn	Ile	Gly	Ile	Val	Asp	Val	Ile	Glu	Lys	Arg	Lys	Phe	Asn	165	170	175	
His	Asn	Ser	Asn	Ser	Thr	Phe	Cys	Val	Phe	Met	Val	Asn	Lys	Pro	Tyr	180	185	190	
Ala	Ile	Thr	Cys	Ser	Val	Val	Ala	Phe	Tyr	Ile	Pro	Phe	Leu	Leu	Met	195	200	205	
Val	Leu	Ala	Tyr	Tyr	Arg	Ile	Tyr	Val	Thr	Ala	Lys	Glu	His	Ala	Gln	210	215	220	
Gln	Ile	Gln	Met	Leu	Gln	Arg	Ala	Gly	Ala	Thr	Ser	Glu	Ser	Arg	Pro	225	230	235	240
Gln	Thr	Ala	Asp	Gln	His	Ser	Thr	His	Arg	Met	Arg	Thr	Glu	Thr	Lys	245	250	255	
Ala	Ala	Lys	Thr	Leu	Cys	Val	Ile	Met	Gly	Cys	Phe	Cys	Phe	Cys	Trp	260	265	270	
Ala	Pro	Phe	Phe	Val	Thr	Asn	Ile	Val	Asp	Pro	Phe	Ile	Asp	Tyr	Thr	275	280	285	
Val	Pro	Glu	Lys	Val	Trp	Thr	Ala	Phe	Leu	Trp	Leu	Gly	Tyr	Ile	Asn	290	295	300	
Ser	Gly	Leu	Asn	Pro	Phe	Leu	Tyr	Ala	Phe	Leu	Asn	Lys	Ser	Phe	Arg	305	310	315	320
Arg	Ala	Phe	Leu	Ile	Ile	Leu	Cys	Cys	Asp	Asp	Glu	Arg	Tyr	Lys	Arg	325	330	335	
Pro	Pro	Ile	Leu	Gly	Gln	Thr	Val	Pro	Cys	Ser	Thr	Thr	Thr	Ile	Asn	340	345	350	
Gly	Ser	Thr	His	Val	Leu	Arg	Tyr	Thr	Val	Leu	His	Ser	Gly	Gln	His	355	360	365	
Gln	Glu	Leu	Glu	Lys	Leu	Pro	Ile	His	Asn	Asp	Pro	Glu	Ser	Leu	Glu	370	375	380	
Ser	Cys	Phe																	

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(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1622 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: N

(iv) ANTI-SENSE: N

(vi) ORIGINAL SOURCE:

(F) TISSUE TYPE: brain

(vii) IMMEDIATE SOURCE:

(A) LIBRARY: rat brain

(B) CLONE: S10-95

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 50..1267
 (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

AGGGTCCTGT	GGGCACTGAC	ATCCAACGTA	CTCATGCCCA	TTTCCTGTA	ATG GAC	55
					Met Asp	
					1	
AGA CTT GAT GCT AAT GTG AGT TCC AAC GAG GGT TTC GGG TCT GTG GAG	103					
Arg Leu Asp Ala Asn Val Ser Ser Asn Glu Gly Phe Gly Ser Val Glu						
5 10 15						
AAG GTC GTA CTG CTC ACG TTC TTC GCA ATG GTT ATC CTG ATG GCC ATC	151					
Lys Val Val Leu Leu Thr Phe Phe Ala Met Val Ile Leu Met Ala Ile						
20 25 30						
CTG GGC AAC CTG CTG GTG ATG GTT GCT GTG TGC AGG GAC AGG CAG CTC	199					
Leu Gly Asn Leu Leu Val Met Val Ala Val Cys Arg Asp Arg Gln Leu						

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35					40					45					50	
AGG	AAA	ATA	AAA	ACC	AAT	TAT	TTC	ATT	GTG	TCT	CTT	GCC	TTT	GCT	GAT	247
Arg	Lys	Ile	Lys	Thr	Asn	Tyr	Phe	Ile	Val	Ser	Leu	Ala	Phe	Ala	Asp	
				55					60					65		
CTG	CTG	GTT	TCG	GTG	CTG	GTG	AAT	GCC	TTC	GGT	GCC	ATT	GAG	TTG	GTC	295
Leu	Leu	Val	Ser	Val	Leu	Val	Asn	Ala	Phe	Gly	Ala	Ile	Glu	Leu	Val	
			70					75					80			
CAA	GAC	ATC	TGG	TTT	TAT	GGG	GAG	ATG	TTT	TGC	CTG	GTC	CGG	ACC	TCT	343
Gln	Asp	Ile	Trp	Phe	Tyr	Gly	Glu	Met	Phe	Cys	Leu	Val	Arg	Thr	Ser	
		85					90					95				
CTG	GAT	GTC	CTA	CTC	ACC	ACA	GCA	TCA	ATT	TTT	CAC	CTC	TGC	TGC	CTT	391
Leu	Asp	Val	Leu	Leu	Thr	Thr	Ala	Ser	Ile	Phe	His	Leu	Cys	Cys	Leu	
	100						105				110					
TCC	CTG	GAT	AGG	TAT	TAT	GCC	ATC	TGC	TGT	CAA	CCT	TTG	GTT	TAT	AGA	439
Ser	Leu	Asp	Arg	Tyr	Tyr	Ala	Ile	Cys	Cys	Gln	Pro	Leu	Val	Tyr	Arg	
115					120					125					130	
AAC	AAG	ATG	ACC	CCT	CTA	CGC	ATC	GCA	TTA	ATG	CTG	GGA	GGC	TGC	TGG	487
Asn	Lys	Met	Thr	Pro	Leu	Arg	Ile	Ala	Leu	Met	Leu	Gly	Gly	Cys	Trp	
				135					140					145		
GTC	ATT	CCC	ATG	TTT	ATA	TCT	TTT	CTC	CCC	ATA	ATG	CAA	GGC	TGG	AAC	535
Val	Ile	Pro	Met	Phe	Ile	Ser	Phe	Leu	Pro	Ile	Met	Gln	Gly	Trp	Asn	
			150					155					160			
AAC	ATC	GGC	ATA	GTT	GAT	GTG	ATA	GAG	AAA	AGG	AAA	TTC	AAC	CAC	AAC	583
Asn	Ile	Gly	Ile	Val	Asp	Val	Ile	Glu	Lys	Arg	Lys	Phe	Asn	His	Asn	
		165					170					175				
TCT	AAC	TCT	ACA	TTC	TGT	GTC	TTC	ATG	GTC	AAC	AAG	CCC	TAT	GCC	ATC	631
Ser	Asn	Ser	Thr	Phe	Cys	Val	Phe	Met	Val	Asn	Lys	Pro	Tyr	Ala	Ile	
	180					185					190					
ACC	TGC	TCT	GTG	GTG	GCC	TTC	TAC	ATC	CCG	TTT	CTC	CTC	ATG	GTG	CTG	679
Thr	Cys	Ser	Val	Val	Ala	Phe	Tyr	Ile	Pro	Phe	Leu	Leu	Met	Val	Leu	
195					200				205						210	
GCC	TAT	TAC	CGT	ATC	TAT	GTC	ACT	GCT	AAG	GAG	CAT	GCC	CAG	CAG	ATC	727
Ala	Tyr	Tyr	Arg	Ile	Tyr	Val	Thr	Ala	Lys	Glu	His	Ala	Gln	Gln	Ile	
				215				220					225			

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CAG	ATG	TTA	CAA	CGG	GCA	GGA	GCC	ACC	TCT	GAA	AGC	AGG	CCC	CAG	ACA	775
Gln	Met	Leu	Gln	Arg	Ala	Gly	Ala	Thr	Ser	Glu	Ser	Arg	Pro	Gln	Thr	
			230					235					240			
GCT	GAC	CAG	CAC	AGC	ACA	CAT	CGC	ATG	CGG	ACA	GAG	ACC	AAA	GCA	GCC	823
Ala	Asp	Gln	His	Ser	Thr	His	Arg	Met	Arg	Thr	Glu	Thr	Lys	Ala	Ala	
		245					250					255				
AAG	ACT	TTA	TGT	GTC	ATC	ATG	GGC	TGC	TTC	TGT	TTC	TGC	TGG	GCC	CCC	871
Lys	Thr	Leu	Cys	Val	Ile	Met	Gly	Cys	Phe	Cys	Phe	Cys	Trp	Ala	Pro	
	260					265					270					
TTC	TTT	GTC	ACC	AAT	ATT	GTG	GAC	CCT	TTC	ATA	GAC	TAC	ACT	GTG	CCC	919
Phe	Phe	Val	Thr	Asn	Ile	Val	Asp	Pro	Phe	Ile	Asp	Tyr	Thr	Val	Pro	
275					280					285					290	
GAG	AAG	GTG	TGG	ACT	GCT	TTC	CTC	TGG	CTT	GGC	TAT	ATC	AAT	TCA	GGG	967
Glu	Lys	Val	Trp	Thr	Ala	Phe	Leu	Trp	Leu	Gly	Tyr	Ile	Asn	Ser	Gly	
				295					300					305		
TTG	AAC	CCT	TTT	CTC	TAT	GCC	TTC	TTG	AAT	AAG	TCT	TTC	AGA	CGT	GCC	1015
Leu	Asn	Pro	Phe	Leu	Tyr	Ala	Phe	Leu	Asn	Lys	Ser	Phe	Arg	Arg	Ala	
			310					315					320			
TTC	CTT	ATC	ATC	CTC	TGC	TGT	GAT	GAT	GAG	CGC	TAC	AAA	AGA	CCC	CCC	1063
Phe	Leu	Ile	Ile	Leu	Cys	Cys	Asp	Asp	Glu	Arg	Tyr	Lys	Arg	Pro	Pro	
		325					330					335				
ATT	CTG	GGC	CAG	ACT	GTC	CCC	TGT	TCA	ACC	ACA	ACC	ATT	AAT	GGA	TCC	1111
Ile	Leu	Gly	Gln	Thr	Val	Pro	Cys	Ser	Thr	Thr	Thr	Ile	Asn	Gly	Ser	
	340					345					350					
ACT	CAT	GTG	CTA	AGG	GAT	ACA	GTG	GAA	TGT	GGT	GGC	CAA	TGG	GAG	AGT	1159
Thr	His	Val	Leu	Arg	Asp	Thr	Val	Glu	Cys	Gly	Gly	Gln	Trp	Glu	Ser	
355					360					365					370	
CGG	TGT	CAC	CTC	ACA	GCA	ACT	TCT	CCT	TTG	GTG	GCT	GCT	CAG	CCA	GTG	1207
Arg	Cys	His	Leu	Thr	Ala	Thr	Ser	Pro	Leu	Val	Ala	Ala	Gln	Pro	Val	
				375					380					385		
ATA	CGT	AGG	CCC	CAG	GAC	AAT	GAC	CTA	GAA	GAC	AGC	TGT	AGC	TTG	AAA	1255
Ile	Arg	Arg	Pro	Gln	Asp	Asn	Asp	Leu	Glu	Asp	Ser	Cys	Ser	Leu	Lys	
			390					395					400			
AGA	ACC	CAG	TCC	TAAGCTGCTA		CTTCGGTGTA		TGTGGCTGCC		CCTGGCACTT						1307

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Arg Ser Gln Ser
405

TGTTCTCCAA	GGCTTTCCAA	GAGCATGAGG	CAATCCACCE	TGGACTTCCC	GCCACGATTC	1367
TAGCAGGCGG	TATTAGAGGA	AGTCAGGGGA	GAGAAGGGCT	TCCTCCTTAG	CTTCTGTTT	1427
CTCAACATT	TCTCTTCTG	GAGTCTCCAC	TCTTGCTTGG	TGGTCTCTGA	AGTCCACGAC	1487
CCAGTCCCT	TTTGCTGTCT	CCAGTCTGTC	TTGTAAATGT	TTACCGTGTT	CGATTTTCAG	1547
TTTCCAAACA	TGCTTCTTT	GAAGTGTCT	CTTACGATAC	TGTCAAACA	TGTGCCTGTC	1607
TTGATCACAC	TTCTT					1622

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 406 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met	Asp	Arg	Leu	Asp	Ala	Asn	Val	Ser	Ser	Asn	Glu	Gly	Phe	Gly	Ser	1	5	10	15
Val	Glu	Lys	Val	Val	Leu	Leu	Thr	Phe	Phe	Ala	Met	Val	Ile	Leu	Met	20	25	30	
Ala	Ile	Leu	Gly	Asn	Leu	Leu	Val	Met	Val	Ala	Val	Cys	Arg	Asp	Arg	35	40	45	
Gln	Leu	Arg	Lys	Ile	Lys	Thr	Asn	Tyr	Phe	Ile	Val	Ser	Leu	Ala	Phe	50	55	60	
Ala	Asp	Leu	Leu	Val	Ser	Val	Leu	Val	Asn	Ala	Phe	Gly	Ala	Ile	Glu	65	70	75	80
Leu	Val	Gln	Asp	Ile	Trp	Phe	Tyr	Gly	Glu	Met	Phe	Cys	Leu	Val	Arg	85	90	95	

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Thr	Ser	Leu	Asp	Val	Leu	Leu	Thr	Thr	Ala	Ser	Ile	Phe	His	Leu	Cys
			100					105					110		
Cys	Leu	Ser	Leu	Asp	Arg	Tyr	Tyr	Ala	Ile	Cys	Cys	Gln	Pro	Leu	Val
			115				120					125			
Tyr	Arg	Asn	Lys	Met	Thr	Pro	Leu	Arg	Ile	Ala	Leu	Met	Leu	Gly	Gly
	130					135					140				
Cys	Trp	Val	Ile	Pro	Met	Phe	Ile	Ser	Phe	Leu	Pro	Ile	Met	Gln	Gly
145					150					155					160
Trp	Asn	Asn	Ile	Gly	Ile	Val	Asp	Val	Ile	Glu	Lys	Arg	Lys	Phe	Asn
				165					170					175	
His	Asn	Ser	Asn	Ser	Thr	Phe	Cys	Val	Phe	Met	Val	Asn	Lys	Pro	Tyr
			180					185					190		
Ala	Ile	Thr	Cys	Ser	Val	Val	Ala	Phe	Tyr	Ile	Pro	Phe	Leu	Leu	Met
		195					200					205			
Val	Leu	Ala	Tyr	Tyr	Arg	Ile	Tyr	Val	Thr	Ala	Lys	Glu	His	Ala	Gln
	210					215					220				
Gln	Ile	Gln	Met	Leu	Gln	Arg	Ala	Gly	Ala	Thr	Ser	Glu	Ser	Arg	Pro
225					230					235					240
Gln	Thr	Ala	Asp	Gln	His	Ser	Thr	His	Arg	Met	Arg	Thr	Glu	Thr	Lys
				245					250					255	
Ala	Ala	Lys	Thr	Leu	Cys	Val	Ile	Met	Gly	Cys	Phe	Cys	Phe	Cys	Trp
			260					265					270		
Ala	Pro	Phe	Phe	Val	Thr	Asn	Ile	Val	Asp	Pro	Phe	Ile	Asp	Tyr	Thr
		275					280					285			
Val	Pro	Glu	Lys	Val	Trp	Thr	Ala	Phe	Leu	Trp	Leu	Gly	Tyr	Ile	Asn
	290					295					300				
Ser	Gly	Leu	Asn	Pro	Phe	Leu	Tyr	Ala	Phe	Leu	Asn	Lys	Ser	Phe	Arg
305					310					315					320
Arg	Ala	Ph	Leu	Ile	Ile	Leu	Cys	Cys	Asp	Asp	Glu	Arg	Tyr	Lys	Arg
				325					330					335	

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Pro	Pro	Ile	Leu	Gly	Gln	Thr	Val	Pro	Cys	Ser	Thr	Thr	Thr	Ile	Asn
			340					345					350		
Gly	Ser	Thr	His	Val	Leu	Arg	Asp	Thr	Val	Glu	Cys	Gly	Gly	Gln	Trp
		355					360					365			
Glu	Ser	Arg	Cys	His	Leu	Thr	Ala	Thr	Ser	Pro	Leu	Val	Ala	Ala	Gln
		370				375					380				
Pro	Val	Ile	Arg	Arg	Pro	Gln	Asp	Asn	Asp	Leu	Glu	Asp	Ser	Cys	Ser
385					390					395					400
Leu	Lys	Arg	Ser	Gln	Ser										
					405										

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 536 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: N

(iv) ANTI-SENSE: N

(vi) ORIGINAL SOURCE:

(F) TISSUE TYPE: brain

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: human brain
- (B) CLONE: S10 PCR

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..534
- (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

TTG GTC TAT AGG AAC AAG ATG ACC CCT CTG CGC ATC GCA TTA ATG CTG

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Leu	Val	Tyr	Arg	Asn	Lys	Met	Thr	Pro	Leu	Arg	Ile	Ala	Leu	Met	Leu	
1				5					10					15		
GGA	GGC	TGC	TGG	GTC	ATC	CCC	ACG	TTT	ATT	TCT	TTT	CTC	CCT	ATA	ATG	96
Gly	Gly	Cys	Trp	Val	Ile	Pro	Thr	Phe	Ile	Ser	Phe	Leu	Pro	Ile	Met	
			20					25					30			
CAA	GGC	TGG	AAT	AAC	ATT	GGC	ATA	ATT	GAT	TTG	ATA	GAA	AAG	AGG	AAG	144
Gln	Gly	Trp	Asn	Asn	Ile	Gly	Ile	Ile	Asp	Leu	Ile	Glu	Lys	Arg	Lys	
			35				40					45				
TTC	AAC	CAG	AAC	TCT	AAC	TCT	ACG	TAC	TGT	GTC	TTC	ATG	GTC	AAC	AAG	192
Phe	Asn	Gln	Asn	Ser	Asn	Ser	Thr	Tyr	Cys	Val	Phe	Met	Val	Asn	Lys	
	50					55					60					
CCC	TAC	GCC	ATC	ACC	TGC	TCT	GTG	GTG	GCC	TTC	TAC	ATC	CCA	TTT	CTC	240
Pro	Tyr	Ala	Ile	Thr	Cys	Ser	Val	Val	Ala	Phe	Tyr	Ile	Pro	Phe	Leu	
	65				70					75					80	
CTC	ATG	GTG	CTG	GCC	TAT	TAC	CGC	ATC	TAT	GTC	ACA	GCT	AAG	GAG	CAT	288
Leu	Met	Val	Leu	Ala	Tyr	Tyr	Arg	Ile	Tyr	Val	Thr	Ala	Lys	Glu	His	
				85				90						95		
GCC	CAT	CAG	ATC	CAG	ATG	TTA	CAA	CGG	GCA	GGA	GCC	TCC	TCC	GAG	AGC	336
Ala	His	Gln	Ile	Gln	Met	Leu	Gln	Arg	Ala	Gly	Ala	Ser	Ser	Glu	Ser	
			100					105					110			
AGG	CCT	CAG	TGG	GCA	GAC	CAG	CAT	AGC	ACT	CAT	CCG	ATG	AGG	ACA	GAG	384
Arg	Pro	Gln	Ser	Ala	Asp	Gln	His	Ser	Thr	His	Pro	Met	Arg	Thr	Glu	
		115					120					125				
ACC	AAA	GCA	GCC	AAG	ACC	CTG	TGC	ATC	ATC	ATG	GGT	TGC	TTC	TGC	CTC	432
Thr	Lys	Ala	Ala	Lys	Thr	Leu	Cys	Ile	Ile	Met	Gly	Cys	Phe	Cys	Leu	
	130					135					140					
TGC	TGG	GCA	CCA	TTC	TTT	GTC	ACC	AAT	ATT	GTG	GAT	CCT	TTC	ATA	GAC	480
Cys	Trp	Ala	Pro	Phe	Phe	Val	Thr	Asn	Ile	Val	Asp	Pro	Phe	Ile	Asp	
	145				150				155						160	
TAC	ACT	GTC	CCT	GGG	CAG	GTG	TGG	ACT	GCT	TTC	CTC	TGG	CTC	GGC	TAT	528
Tyr	Thr	Val	Pro	Gly	Gln	Val	Trp	Thr	Ala	Phe	Leu	Trp	Leu	Gly	Tyr	
				165				170						175		
ATC	AAT	TC														536
Ile	Asn															

(A) LENGTH: 178 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: Linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Leu	Val	Tyr	Arg	Asn	Lys	Met	Thr	Pro	Leu	Arg	Ile	Ala	Leu	Met	Leu
1				5					10					15	

Gly Gly Cys Trp Val Ile Pro Thr Phe Ile Ser Phe Leu Pro Ile Met
20 25 30

Gln	Gly	Trp	Asn	Asn	Ile	Gly	Ile	Ile	Asp	Leu	Ile	Glu	Lys	Arg	Lys
		35					40					45			

Phe Asn Gln Asn Ser Asn Ser Thr Tyr Cys Val Phe Met Val Asn Lys
50 55 60

Pro	Tyr	Ala	Ile	Thr	Cys	Ser	Val	Val	Ala	Phe	Tyr	Ile	Pro	Phe	Leu
65					70					75					80

Leu Met Val Leu Ala Tyr Tyr Arg Ile Tyr Val Thr Ala Lys Glu His
85 90 95

Ala	His	Gln	Ile	Gln	Met	Leu	Gln	Arg	Ala	Gly	Ala	Ser	Ser	Glu	Ser
			100					105					110		

Arg	Pro	Gln	Ser	Ala	Asp	Gln	His	Ser	Thr	His	Pro	Met	Arg	Thr	Glu
		115					120					125			

Thr	Lys	Ala	Ala	Lys	Thr	Leu	Cys	Ile	Ile	Met	Gly	Cys	Phe	Cys	Leu
	130					135					140				

Cys	Trp	Ala	Pro	Phe	Phe	Val	Thr	Asn	Ile	Val	Asp	Pro	Phe	Ile	Asp
145					150					155					160

Tyr	Thr	Val	Pro	Gly	Gln	Val	Trp	Thr	Ala	Phe	Leu	Trp	Leu	Gly	Tyr
				165					170					175	

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Ile Asn

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1316 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(F) TISSUE TYPE: Brain

(vii) IMMEDIATE SOURCE:

- (A) LIBRARY: Human Brain
 (B) CLONE: S10-87

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 7..1170

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CCTGTA	ATG	GAC	AAA	CTT	GAT	GCT	AAT	GTG	AGT	TCT	GAG	GAG	GGT	TTC	48	
Met	Asp	Lys	Leu	Asp	Ala	Asn	Val	Ser	Ser	Glu	Glu	Gly	Phe			
1				5				10								
GGG	TCA	GTG	GAG	AAG	GTG	GTG	CTG	CTC	ACG	TTT	CTC	TCG	ACG	GTT	ATC	96
Gly	Ser	Val	Glu	Lys	Val	Val	Leu	Leu	Thr	Phe	Leu	Ser	Thr	Val	Ile	
15				20				25						30		
CTG	ATG	GCC	ATC	TTG	GGG	AAC	CTG	CTG	GTG	ATG	GTG	GCT	GTG	TGC	TGG	144
Leu	Met	Ala	Ile	Leu	Gly	Asn	Leu	Leu	Val	Met	Val	Ala	Val	Cys	Trp	
				35				40						45		
GAC	AGG	CAG	CTC	AGG	AAA	ATA	AAA	ACA	AAT	TAT	TTC	ATT	GTA	TCT	CTT	192
Asp	Arg	Gln	Leu	Arg	Lys	Ile	Lys	Thr	Asn	Tyr	Phe	Ile	Val	Ser	Leu	
				50				55					60			

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GCT	TTT	GCG	GAT	CTG	CTG	GTT	TCG	GTG	CTG	GTG	ATG	CCC	TTT	GGT	GCC	240
Ala	Phe	Ala	Asp	Leu	Leu	Val	Ser	Val	Leu	Val	Met	Pro	Phe	Gly	Ala	
		65					70					75				
ATT	GAG	CTG	GTT	CAA	GAC	ATC	TGG	ATT	TAT	GGG	GAG	GTG	TTT	TGT	CTT	288
Ile	Glu	Leu	Val	Gln	Asp	Ile	Trp	Ile	Tyr	Gly	Glu	Val	Phe	Cys	Leu	
	80					85					90					
GTT	CGG	ACA	TCT	CTG	GAC	GTC	CTG	CTC	ACA	ACG	GCA	TCG	ATT	TTT	CAC	336
Val	Arg	Thr	Ser	Leu	Asp	Val	Leu	Leu	Thr	Thr	Ala	Ser	Ile	Phe	His	
	95				100					105					110	
CTG	TGC	TGC	ATT	TCT	CTG	GAT	AGG	TAT	TAC	GCC	ATC	TGC	TGC	CAG	CCT	384
Leu	Cys	Cys	Ile	Ser	Leu	Asp	Arg	Tyr	Tyr	Ala	Ile	Cys	Cys	Gln	Pro	
				115					120					125		
TTG	GTC	TAT	AGG	AAC	AAG	ATG	ACC	CCT	CTG	CGC	ATC	GCA	TTA	ATG	CTG	432
Leu	Val	Tyr	Arg	Asn	Lys	Met	Thr	Pro	Leu	Arg	Ile	Ala	Leu	Met	Leu	
			130					135					140			
GGA	GGC	TGC	TGG	GTC	ATC	CCC	ACG	TTT	ATT	TCT	TTT	CTC	CCT	ATA	ATG	480
Gly	Gly	Cys	Trp	Val	Ile	Pro	Thr	Phe	Ile	Ser	Phe	Leu	Pro	Ile	Met	
		145					150					155				
CAA	GGC	TGG	AAT	AAC	ATT	GGC	ATA	ATT	GAT	TTG	ATA	GAA	AAG	AGG	AAG	528
Gln	Gly	Trp	Asn	Asn	Ile	Gly	Ile	Ile	Asp	Leu	Ile	Glu	Lys	Arg	Lys	
	160					165					170					
TTC	AAC	CAG	AAC	TCT	AAC	TCT	ACG	TAC	TGT	GTC	TTC	ATG	GTC	AAC	AAG	576
Phe	Asn	Gln	Asn	Ser	Asn	Ser	Thr	Tyr	Cys	Val	Phe	Met	Val	Asn	Lys	
	175				180					185					190	
CCC	TAC	GCC	ATC	ACC	TGC	TCT	GTG	GTG	GCC	TTC	TAC	ATC	CCA	TTT	CTC	624
Pro	Tyr	Ala	Ile	Thr	Cys	Ser	Val	Val	Ala	Phe	Tyr	Ile	Pro	Phe	Leu	
				195					200					205		
CTC	ATG	GTG	CTG	GCC	TAT	TAC	CGC	ATC	TAT	GTC	ACA	GCT	AAG	GAG	CAT	672
Leu	Met	Val	Leu	Ala	Tyr	Tyr	Arg	Ile	Tyr	Val	Thr	Ala	Lys	Glu	His	
			210					215					220			
GCC	CAT	CAG	ATC	CAG	ATG	TTA	CAA	CGG	GCA	GGA	GCC	TCC	TCC	GAG	AGC	720
Ala	His	Gln	Ile	Gln	Met	Leu	Gln	Arg	Ala	Gly	Ala	Ser	Ser	Glu	Ser	
		225					230					235				
AGG	CCT	CAG	TCG	GCA	GAC	CAG	CAT	AGC	ACT	CAT	CGC	ATG	AGG	ACA	GAG	768

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Arg	Pro	Gln	Ser	Ala	Asp	Gln	His	Ser	Thr	His	Arg	Met	Arg	Thr	Glu	
	240					245					250					
ACC	AAA	GCA	GCC	AAG	ACC	CTG	TGC	ATC	ATC	ATG	GGT	TGC	TTC	TGC	CTC	816
Thr	Lys	Ala	Ala	Lys	Thr	Leu	Cys	Ile	Ile	Met	Gly	Cys	Phe	Cys	Leu	
255					260					265					270	
TGC	TGG	GCA	CCA	TTC	TTT	GTC	ACC	AAT	ATT	GTG	GAT	CCT	TTC	ATA	GAC	864
Cys	Trp	Ala	Pro	Phe	Phe	Val	Thr	Asn	Ile	Val	Asp	Pro	Phe	Ile	Asp	
				275					280					285		
TAC	ACT	GTC	CCT	GGG	CAG	GTG	TGG	ACT	GCT	TTC	CTC	TGG	CTC	GGC	TAT	912
Tyr	Thr	Val	Pro	Gly	Gln	Val	Trp	Thr	Ala	Phe	Leu	Trp	Leu	Gly	Tyr	
			290					295					300			
ATC	AAT	TCC	GGG	TTG	AAC	CCT	TTT	CTC	TAC	GCC	TTC	TTG	AAT	AAG	TCT	960
Ile	Asn	Ser	Gly	Leu	Asn	Pro	Phe	Leu	Tyr	Ala	Phe	Leu	Asn	Lys	Ser	
		305					310					315				
TTT	AGA	CGT	GCC	TTC	CTC	ATC	ATC	CTC	TGC	TGT	GAT	GAT	GAG	CGC	TAC	1008
Phe	Arg	Arg	Ala	Phe	Leu	Ile	Ile	Leu	Cys	Cys	Asp	Asp	Glu	Arg	Tyr	
	320					325					330					
CGA	AGA	CCT	TCC	ATT	CTG	GGC	CAG	ACT	GTC	CCT	TGT	TCA	ACC	ACA	ACC	1056
Arg	Arg	Pro	Ser	Ile	Leu	Gly	Gln	Thr	Val	Pro	Cys	Ser	Thr	Thr	Thr	
335					340					345					350	
ATT	AAT	GGA	TCC	ACA	CAT	GTA	CTA	AGG	GAT	GCA	GTG	GAG	TGT	GGT	GGC	1104
Ile	Asn	Gly	Ser	Thr	His	Val	Leu	Arg	Asp	Ala	Val	Glu	Cys	Gly	Gly	
				355					360					365		
CAG	TGG	GAG	AGT	CAG	TGT	CAC	CCG	CCA	GCA	ACT	TCT	CCT	TTG	GTG	GCT	1152
Gln	Trp	Glu	Ser	Gln	Cys	His	Pro	Pro	Ala	Thr	Ser	Pro	Leu	Val	Ala	
			370					375					380			
GCT	CAG	CCC	AGT	GAC	ACT	TAGGCCCTG		GGACAATGAC		CCAGAAGACA						1200
Ala	Gln	Pro	Ser	Asp	Thr											
		385														
GCCATGCCTC		CGAAAGAGGG		CCAGGTCTA		AGCTGCTGCT		TGTGCGCGAC		TGCACCCGGC						1260
ATTCTCTTCA		CCTGAGGCTT		TCCGTCCGCC		AGTGCAGGAA		CCCGGTGCTC		GCTGGG						1316

(2) INFORMATION FOR SEQ ID NO:8:

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(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 388 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met	Asp	Lys	Leu	Asp	Ala	Asn	Val	Ser	Ser	Glu	Glu	Gly	Phe	Gly	Ser	
1				5					10					15		
Val	Glu	Lys	Val	Val	Leu	Leu	Thr	Phe	Leu	Ser	Thr	Val	Ile	Leu	Met	
			20					25					30			
Ala	Ile	Leu	Gly	Asn	Leu	Leu	Val	Met	Val	Ala	Val	Cys	Trp	Asp	Arg	
		35					40					45				
Gln	Leu	Arg	Lys	Ile	Lys	Thr	Asn	Tyr	Phe	Ile	Val	Ser	Leu	Ala	Phe	
	50					55					60					
Ala	Asp	Leu	Leu	Val	Ser	Val	Leu	Val	Met	Pro	Phe	Gly	Ala	Ile	Glu	
	65				70					75					80	
Leu	Val	Gln	Asp	Ile	Trp	Ile	Tyr	Gly	Glu	Val	Phe	Cys	Leu	Val	Arg	
			85					90						95		
Thr	Ser	Leu	Asp	Val	Leu	Leu	Thr	Thr	Ala	Ser	Ile	Phe	His	Leu	Cys	
			100					105					110			
Cys	Ile	Ser	Leu	Asp	Arg	Tyr	Tyr	Ala	Ile	Cys	Cys	Gln	Pro	Leu	Val	
		115					120					125				
Tyr	Arg	Asn	Lys	Met	Thr	Pro	Leu	Arg	Ile	Ala	Leu	Met	Leu	Gly	Gly	
	130					135						140				
Cys	Trp	Val	Ile	Pro	Thr	Phe	Ile	Ser	Phe	Leu	Pro	Ile	Met	Gln	Gly	
	145				150					155					160	
Trp	Asn	Asn	Ile	Gly	Ile	Ile	Asp	Leu	Ile	Glu	Lys	Arg	Lys	Phe	Asn	
			165						170					175		
Gln	Asn	Ser	Asn	Ser	Thr	Tyr	Cys	Val	Phe	Met	Val	Asn	Lys	Pro	Tyr	
			180					185					190			

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[illegible]

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(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

(A) NAME/KEY: modified_base

(B) LOCATION: 14

(ix) FEATURE:

(A) NAME/KEY: modified_base

(B) LOCATION: 20

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

TGGAATTCTG YGYAATHKCA CTGGAYMGST A

31

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

(A) NAME/KEY: modified_base

(B) LOCATION: 4

(ix) FEATURE:

(A) NAME/KEY: modified_base

(B) LOCATION: 7

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(ix) FEATURE:

(A) NAME/KEY: modified_base

(B) LOCATION: 9..10

(ix) FEATURE:

(A) NAME/KEY: modified_base

(B) LOCATION: 13

(ix) FEATURE:

(A) NAME/KEY: modified_base

(B) LOCATION: 25

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CATAAVARAA ARAGGDATRW ARAAAGC

27

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 45 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

TCAAAAGCAT GATTCAGGG ACTCTGGGTC ATTGTGTATG GGCAA

45

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

CCTCAATCAG AAGCATGATT CCAGG

25

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

TTGGTCTATA GGAACAAGAT GACCC

25

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 792 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

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(vi) ORIGINAL SOURCE:

(F) TISSUE TYPE: Brain

(vii) IMMEDIATE SOURCE:

(A) LIBRARY: HUMAN BRAIN

(B) CLONE: S10-87

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..783

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

TTG	GTC	TAT	AGG	AAC	AAG	ATG	ACC	CCT	CTG	CGC	ATC	GCA	TTA	ATG	CTG	48
Leu	Val	Tyr	Arg	Asn	Lys	Met	Thr	Pro	Leu	Arg	Ile	Ala	Leu	Met	Leu	
1				5					10					15		
GGA	GGC	TGC	TGG	GTC	ATC	CCC	ACG	TTT	ATT	TCT	TTT	CTC	CCT	ATA	ATG	96
Gly	Gly	Cys	Trp	Val	Ile	Pro	Thr	Phe	Ile	Ser	Phe	Leu	Pro	Ile	Met	
			20					25					30			
CAA	GGC	TGG	AAT	AAC	ATT	GGC	ATA	ATT	GAT	TTG	ATA	GAA	AAG	AGG	AAG	144
Gln	Gly	Trp	Asn	Asn	Ile	Gly	Ile	Ile	Asp	Leu	Ile	Glu	Lys	Arg	Lys	
			35				40					45				
TTC	AAC	CAG	AAC	TCT	AAC	TCT	ACG	TAC	TGT	GTC	TTC	ATG	GTC	AAC	AAG	192
Phe	Asn	Gln	Asn	Ser	Asn	Ser	Thr	Tyr	Cys	Val	Phe	Met	Val	Asn	Lys	
	50					55					60					
CCC	TAC	GCC	ATC	ACC	TGC	TCT	GTG	GTG	GCC	TTC	TAC	ATC	CCA	TTT	CTC	240
Pro	Tyr	Ala	Ile	Thr	Cys	Ser	Val	Val	Ala	Phe	Tyr	Ile	Pro	Phe	Leu	
65					70				75						80	
CTC	ATG	GTG	CTG	GCC	TAT	TAC	CGC	ATC	TAT	GTC	ACA	GCT	AAG	GAG	CAT	288
Leu	Met	Val	Leu	Ala	Tyr	Tyr	Arg	Ile	Tyr	Val	Thr	Ala	Lys	Glu	His	
				85				90						95		
GCC	CAT	CAG	ATC	CAG	ATG	TTA	CAA	CGG	GCA	GGA	GCC	TCC	TCC	GAG	AGC	336
Ala	His	Gln	Ile	Gln	Met	Leu	Gln	Arg	Ala	Gly	Ala	Ser	Ser	Glu	Ser	
			100					105					110			
AGG	CCT	CAG	TCG	GCA	GAC	CAG	CAT	AGC	ACT	CAT	CGC	ATG	AGG	ACA	GAG	384
Arg	Pro	Gln	Ser	Ala	Asp	Gln	His	Ser	Thr	His	Arg	Met	Arg	Thr	Glu	
			115					120				125				

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ACC	AAA	GCA	GCC	AAG	ACC	CTG	TGC	ATC	ATC	ATG	GGT	TGC	TTC	TGC	CTC	432
Thr	Lys	Ala	Ala	Lys	Thr	Leu	Cys	Ile	Ile	Met	Gly	Cys	Phe	Cys	Leu	
	130					135					140					
TGC	TGG	GCA	CCA	TTC	TTT	GTC	ACC	AAT	ATT	GTG	GAT	CCT	TTC	ATA	GAC	480
Cys	Trp	Ala	Pro	Phe	Phe	Val	Thr	Asn	Ile	Val	Asp	Pro	Phe	Ile	Asp	
145					150					155					160	
TAC	ACT	GTC	CCT	GGG	CAG	GTG	TGG	ACT	GCT	TTC	CTC	TGG	CTC	GGC	TAT	528
Tyr	Thr	Val	Pro	Gly	Gln	Val	Trp	Thr	Ala	Phe	Leu	Trp	Leu	Gly	Tyr	
				165					170					175		
ATC	AAT	TCC	GGG	TTG	AAC	CCT	TTT	CTC	TAC	GCC	TTC	TTG	AAT	AAG	TCT	576
Ile	Asn	Ser	Gly	Leu	Asn	Pro	Phe	Leu	Tyr	Ala	Phe	Leu	Asn	Lys	Ser	
			180					185					190			
TTT	AGA	CGT	GCC	TTC	CTC	ATC	ATC	CTC	TGC	TGT	GAT	GAT	GAG	CGC	TAC	624
Phe	Arg	Arg	Ala	Phe	Leu	Ile	Ile	Leu	Cys	Cys	Asp	Asp	Glu	Arg	Tyr	
		195					200					205				
CGA	AGA	CCT	TCC	ATT	CTG	GGC	CAG	ACT	GTC	CCT	TGT	TCA	ACC	ACA	ACC	672
Arg	Arg	Pro	Ser	Ile	Leu	Gly	Gln	Thr	Val	Pro	Cys	Ser	Thr	Thr	Thr	
	210					215					220					
ATT	AAT	GGA	TCC	ACA	CAT	GTA	CTA	AGG	TAC	ACC	GTT	CTG	CAC	AGG	GGA	720
Ile	Asn	Gly	Ser	Thr	His	Val	Leu	Arg	Tyr	Thr	Val	Leu	His	Arg	Gly	
225					230					235					240	
CAT	CAT	CAG	GAA	CTC	GAG	AAA	CTG	CCC	ATA	CAC	AAT	GAC	CCA	GAA	TCC	768
His	His	Gln	Glu	Leu	Glu	Lys	Leu	Pro	Ile	His	Asn	Asp	Pro	Glu	Ser	
				245					250					255		
CTG	GAA	TCA	TGC	TTC	TGATTGAGG											792
Leu	Glu	Ser	Cys	Phe												
				260												

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 261 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

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(xi)			SEQUENCE	DESCRIPTION:				SEQ	ID	NO:15:						
Leu	Val	Tyr	Arg	Asn	Lys	Met	Thr	Pro	Leu	Arg	Ile	Ala	Leu	Met	Leu	
1				5					10					15		
Gly	Gly	Cys	Trp	Val	Ile	Pro	Thr	Phe	Ile	Ser	Phe	Leu	Pro	Ile	Met	
			20					25					30			
Gln	Gly	Trp	Asn	Asn	Ile	Gly	Ile	Ile	Asp	Leu	Ile	Glu	Lys	Arg	Lys	
		35					40					45				
Phe	Asn	Gln	Asn	Ser	Asn	Ser	Thr	Tyr	Cys	Val	Phe	Met	Val	Asn	Lys	
	50					55					60					
Pro	Tyr	Ala	Ile	Thr	Cys	Ser	Val	Val	Ala	Phe	Tyr	Ile	Pro	Phe	Leu	
65					70					75					80	
Leu	Met	Val	Leu	Ala	Tyr	Tyr	Arg	Ile	Tyr	Val	Thr	Ala	Lys	Glu	His	
			85					90						95		
Ala	His	Gln	Ile	Gln	Met	Leu	Gln	Arg	Ala	Gly	Ala	Ser	Ser	Glu	Ser	
			100					105					110			
Arg	Pro	Gln	Ser	Ala	Asp	Gln	His	Ser	Thr	His	Arg	Met	Arg	Thr	Glu	
		115					120					125				
Thr	Lys	Ala	Ala	Lys	Thr	Leu	Cys	Ile	Ile	Met	Gly	Cys	Phe	Cys	Leu	
	130					135					140					
Cys	Trp	Ala	Pro	Phe	Phe	Val	Thr	Asn	Ile	Val	Asp	Pro	Phe	Ile	Asp	
145					150					155					160	
Tyr	Thr	Val	Pro	Gly	Gln	Val	Trp	Thr	Ala	Phe	Leu	Trp	Leu	Gly	Tyr	
			165					170						175		
Ile	Asn	Ser	Gly	Leu	Asn	Pro	Phe	Leu	Tyr	Ala	Phe	Leu	Asn	Lys	Ser	
			180					185					190			
Phe	Arg	Arg	Ala	Phe	Leu	Ile	Ile	Leu	Cys	Cys	Asp	Asp	Glu	Arg	Tyr	
		195					200					205				
Arg	Arg	Pro	Ser	Ile	Leu	Gly	Gln	Thr	Val	Pro	Cys	Ser	Thr	Thr	Thr	
	210					215					220					
Ile	Asn	Gly	Ser	Thr	His	Val	Leu	Arg	Tyr	Thr	Val	Leu	His	Arg	Glu	

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225					230					235					240	
His	His	Gln	Glu	Leu	Glu	Lys	Leu	Pro	Ile	His	Asn	Asp	Pro	Glu	Ser	
				245					250					255		
Leu	Glu	Ser	Cys	Phe												
			260													

What is claimed is:

1. An isolated nucleic acid molecule encoding a mammalian 5-HT₄ receptor.
2. A nucleic acid molecule of claim 1, wherein the nucleic acid molecule encodes a human 5-HT₄ receptor.
3. A nucleic acid molecule of claim 1, wherein the nucleic acid molecule is a DNA molecule.
4. A DNA molecule of claim 3, wherein the DNA molecule is a cDNA molecule.
5. A DNA molecule of claim 3 wherein the DNA molecule is genomic DNA.
6. A nucleic acid molecule of claim 2, wherein the nucleic acid molecule is a DNA molecule.
7. An isolated DNA molecule encoding a mammalian 5-HT₄ receptor having the sequence H₂N-Y-X-COOH wherein Y is the amino acid sequence beginning at amino acid 1 and ending at amino acid 359 of Figure 1 (SEQ ID NO. 2) and wherein X is an amino acid sequence encoding the carboxy terminal region of the receptor.
8. An isolated nucleic acid molecule of claim 7, wherein X is the amino acid sequence beginning at amino acid 360 and ending at amino acid 387 of Figure 1 (SEQ ID NO. 2).
9. An isolated nucleic acid molecule of claim 7, wherein X is the amino acid sequence beginning at

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amino acid 360 and ending at amino acid 406 of
Figur 2 (SEQ ID NO. 4).

- 5 10. An isolated nucleic acid molecule of claim 7,
wherein Y is encoded by the nucleotide sequence
from nucleotide 101 to nucleotide 1177 of Figure
1 (SEQ ID NO. 1).
- 10 11. An isolated nucleic acid molecule of claim 8,
wherein X is encoded by the nucleotide sequence
from nucleotide 1178 to nucleotide to 1261 of
Figure 1 (SEQ ID NO. 1).
- 15 12. An isolated nucleic acid molecule of claim 9,
wherein X is encoded by the nucleotide sequence
from nucleotide 1127 to nucleotide 1267 of Figure
2 (SEQ ID NO. 3).
- 20 13. A vector comprising a cDNA molecule of claim 4.
14. A plasmid vector of claim 13.
- 25 15. A vector of claim 13 adapted for expression in a
bacterial cell which comprises regulatory elements
necessary for expression of the cDNA encoding a 5-
HT₁ receptor in the bacterial cell operatively
linked to the cDNA encoding the 5-HT₁ receptor as
to permit expression thereof.
- 30 16. A vector of claim 13 adapted for expression in a
yeast cell which comprises the regulatory elements
necessary for the expression of the cDNA encoding
a 5-HT₁ receptor in the yeast cell operatively
linked to the cDNA encoding the 5-HT₁ receptor as
35 to permit expression thereof.
17. A vector of claim 13 adapted for expression in an

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insect cell which comprises the regulatory elements necessary for expression of the cDNA encoding a 5-HT₁ receptor in the insect cell operatively linked to the cDNA encoding the 5-HT₁ receptor as to permit expression thereof.

18. A vector of claim 13 adapted for expression in a mammalian cell which comprises the regulatory elements necessary for expression of the cDNA encoding a 5-HT₁ receptor in the mammalian cell operatively linked to the cDNA encoding the 5-HT₁ receptor as to permit expression thereof.

19. A plasmid of claim 14 adapted for expression in a mammalian cell which comprises the regulatory elements necessary for expression of the DNA in the mammalian cell operatively linked to the DNA encoding a 5-HT₁ receptor as to permit expression thereof.

20. A plasmid of claim 19 designated pcEXV-S10-87 (ATCC Accession No. 75390).

21. A plasmid of claim 19 designated pcEXV-S10-95 (ATCC Accession No. 75391).

22. A plasmid of claim 19 designated pBluescript-hS10 (ATCC Accession No. 75392).

23. A mammalian cell comprising the plasmid of claim 14.

24. A mammalian cell of claim 23, wherein the mammalian cell is an LM (tk-) cell.

25. A nucleic acid probe comprising a nucleic acid molecule of at least 15 nucleotides capable of

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specifically hybridizing with a unique sequence included within the sequence of a nucleic acid molecule encoding a mammalian 5-HT₁ receptor.

- 5 26. A nucleic acid probe comprising a nucleic acid molecule of at least 15 nucleotides capable of specifically hybridizing with a unique sequence included within the sequence of a nucleic acid molecule encoding a human 5-HT₁ receptor.
- 10 27. The nucleic acid probe of claim 25, wherein the nucleic acid is DNA.
- 15 28. The nucleic acid probe of claim 26, wherein the nucleic acid is DNA.
- 20 29. A mixture of nucleic acid probes in accordance with claim 25, such probes having sequences which differ from one another at predefined positions.
- 25 30. An antisense oligonucleotide having a sequence capable of specifically binding to a mRNA molecule encoding a mammalian 5-HT₁ receptor so as to prevent translation of the mRNA molecule.
- 30 31. An antisense oligonucleotide capable of specifically binding to a mRNA molecule encoding a human 5-HT₁ receptor so as to prevent translation of the mRNA molecule.
- 35 32. An antisense oligonucleotide of claim 30 comprising chemical analogs of nucleotides.
33. A mixture of antisense oligonucleotides according to claim 30, such oligonucleotides having sequences which differ from one another at

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predefined positions.

- 5 34. A method for detecting expression of a mammalian 5-HT₁ receptor, which comprises obtaining RNA from cells or tissue, contacting the RNA so obtained with a nucleic acid probe of claim 25 under hybridizing conditions, detecting the presence of any mRNA hybridized to the probe, the presence of mRNA hybridized to the probe indicating expression of the mammalian 5-HT₁ receptor, and thereby detecting the expression of the mammalian 5-HT₁ receptor.
- 10
- 15 35. A method for detecting expression of a human 5-HT₁ receptor, which comprises obtaining RNA from cells or tissue, contacting the RNA so obtained with a nucleic acid probe of claim 26 under hybridizing conditions, detecting the presence of any mRNA hybridized to the probe, the presence of mRNA hybridized to the probe indicating expression of the human 5-HT₁ receptor, and thereby detecting the expression of the human 5-HT₁ receptor.
- 20
- 25 36. A method of detecting expression of a mammalian 5-HT₁ receptor in a cell or tissue by in situ hybridization, which comprises contacting the cell or tissue with a nucleic acid probe of claim 25 under hybridizing conditions, detecting the presence of any mRNA hybridized to the probe, the presence of mRNA hybridized to the probe indicating expression of a mammalian 5-HT₁ receptor, and thereby detecting the expression of a mammalian 5-HT₁ receptor.
- 30
- 35 37. A method of detecting expression of a human 5-HT₁ receptor in a cell or tissue by in situ hybridization, which comprises contacting the cell

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or tissue with a nucleic acid probe of claim 26 under hybridizing conditions, detecting the presence of any mRNA hybridized to the probe, the presence of mRNA hybridized to the probe indicating expression of a human 5-HT₁ receptor, and thereby detecting the expression of the human 5-HT₁ receptor.

38. A method of isolating from a gene library a gene encoding a receptor other than the 5-HT₁ receptor which comprises contacting the library under hybridizing conditions with a probe of claim 27 and isolating any gene to which the probe hybridizes.
39. A method of claim 38, which additionally comprises simultaneously contacting the DNA comprising the library under hybridizing conditions with a second nucleic acid probe comprising a sequence capable of hybridizing to a DNA sequence of the complementary strand of the DNA of the gene to which the first probe hybridizes, treating any gene sequence to which both probes hybridized so as to produce multiple copies of the gene sequence, isolating the amplified gene sequence and using the isolated gene sequence as a probe to isolate from a gene library the gene to which the amplified DNA sequence hybridizes.
40. The gene isolated by the method of claim 38 or 39.
41. A synthetic gene which comprises the isolated nucleic acid molecule of claim 1 and at least one regulatory element attached thereto so as to increase the number of RNA molecules transcribed from the synthetic gene.

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the resulting cell.

- 5 49. An antibody directed to a mammalian 5-HT₁ receptor or to a protein fragment of the mammalian 5-HT₁ receptor.
- 10 50. An antibody directed to a human 5-HT₁ receptor or to a protein fragment of the human 5-HT₁ receptor.
- 15 51. An antibody of claim 49, wherein the antibody is a monoclonal antibody.
- 20 52. An antibody of claim 50, wherein the antibody is a monoclonal antibody.
- 25 53. A monoclonal antibody of claim 51, wherein the antibody is directed to an epitope of a mammalian cell-surface 5-HT₁ receptor and having an amino acid sequence substantially the same as the amino acid sequence of a cell-surface epitope of the mammalian 5-HT₁ receptor.
- 30 54. A monoclonal antibody of claim 52, wherein the antibody is directed to an epitope of a human cell-surface 5-HT₁ receptor and having an amino acid sequence substantially the same as the amino acid sequence for a cell-surface epitope of the human 5-HT₁ receptor.
- 35 55. A pharmaceutical composition comprising an amount of a substance effective to alleviate the abnormalities resulting from overexpression of a human 5-HT₁ receptor and a pharmaceutically acceptable carrier.
56. A pharmaceutical composition comprising an amount of a substance effective to alleviate

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abnormalities resulting from underexpression of a human 5-HT₁ receptor and a pharmaceutically acceptable carrier.

- 5 57. A pharmaceutical composition comprising an effective amount of an oligonucleotide of claim 31 effective to reduce expression of a human 5-HT₁ receptor by passing through a cell membrane and specifically binding with mRNA encoding a human 5-HT₁ receptor in the cell so as to prevent its translation and a pharmaceutically acceptable hydrophobic carrier capable of passing through a cell membrane.
- 10
- 15 58. A pharmaceutical composition claim 57, wherein the nucleotide is coupled to a substance which inactivates mRNA.
- 20 59. A pharmaceutical composition of claim 58, wherein the substance which inactivates the mRNA is a ribozyme.
- 25 60. A pharmaceutical composition of claim 58, wherein the pharmaceutically acceptable hydrophobic carrier capable of passing through a cell membrane comprises a structure which binds to a transporter specific for a selected cell type and is thereby taken up by the cells of the selected cell type.
- 30 61. A pharmaceutical composition which comprises an amount of the antibody of claim 50 effective to block binding of naturally occurring substrates to a human 5-HT₁ receptor and a pharmaceutically acceptable carrier.
- 35 62. A transgenic nonhuman mammal which comprises a nucleic acid molecule of claim 1.

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- 5 63. A transgenic nonhuman mammal whose genome comprises a nucleic acid molecule of claim 1 so placed as to be transcribed into antisense mRNA complementary to mRNA encoding a human 5-HT₁ receptor and which hybridizes to mRNA encoding a human 5-HT₁ receptor thereby reducing its translation.
- 10 64. The transgenic nonhuman mammal of claim 62, wherein the nucleic acid molecule further comprises an inducible promoter.
- 15 65. The transgenic nonhuman mammal of claim 62 or 64 wherein the nucleic acid molecule additionally comprises tissue specific regulatory elements.
- 20 66. The transgenic non-human mammal of 62 wherein the transgenic non-human mammal is a mouse.
- 25 67. A method for determining the physiological effects of varying the levels of expression of a human 5-HT₁ receptor which comprises producing a transgenic non-human mammal whose levels of expression of a human 5-HT₁ receptor can be varied by use of an inducible promoter.
- 30 68. A method for determining the physiological effects of expressing varying levels of a human 5-HT₁ receptor which comprises producing a panel of transgenic non-human mammals each expressing a different amount of a human 5-HT₁ receptor.
- 35 69. A method for determining whether a compound not known to be capable of specifically binding to a human 5-HT₁ receptor can specifically bind to the human 5-HT₁ receptor, which comprises contacting a mammalian cell comprising a plasmid adapted for

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5 expression in a mammalian cell which plasmid
further comprises DNA which expresses a human 5-
HT₁ receptor on the cell's surface with the
compound under conditions permitting binding of
ligands known to bind to a human 5-HT₁ receptor,
detecting the presence of any compound bound to
the human 5-HT₁ receptor, the presence of bound
compound indicating that the compound is capable
of specifically binding to the human 5-HT₁
10 receptor.

70. The method of claim 70, wherein the mammalian cell
is a non-neuronal cell.

15 71. A method of screening compounds to identify drugs
which interact with, and specifically bind to, a
human 5-HT₁ receptor on the surface of a cell,
which comprises contacting a mammalian cell which
comprises a plasmid adapted for expression in a
20 mammalian cell which plasmid further comprises DNA
which expresses a human 5-HT₁ receptor on the
cell's surface with a plurality of compounds,
determining those compounds which bind to the
human 5-HT₁ receptor expressed on the cell surface
25 of the mammalian cell, and thereby identifying
compounds which interact with, and specifically
bind to, the human 5-HT₁ receptor.

72. The method of claim 71, wherein the mammalian cell
30 is a non-neuronal cell.

73. A method for determining whether a compound not
known to be capable of specifically binding to a
human 5-HT₁ receptor can specifically bind to a
human 5-HT₁ receptor, which comprises preparing a
cell extract from mammalian cells, which comprise
35 a plasmid adapted for expression in a mammal,

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- 5 which plasmid further comprises DNA which expresses a human 5-HT₄ receptor on the cell's surface, isolating a membrane fraction from the cell extract, incubating the compound with the membrane fraction under conditions permitting binding of ligands known to bind to the human 5-HT₄ receptor, detecting the presence of any bound compound, and thereby determining whether the compound is capable of specifically binding to the human 5-HT₄ receptor.
- 10
74. The method of claim 73, wherein the mammalian cell is a non-neuronal cell.
- 15
75. A method for screening compounds to identify drugs that interact with, and specifically bind to, a human 5-HT₄ receptor, which comprises preparing a cell extract from mammalian cells, which comprise a plasmid adapted for expression in a mammalian cell which plasmid further comprises DNA which expresses a human 5-HT₄ receptor on the cell's surface, isolating a membrane fraction from the cell extract, incubating the membrane fraction with a plurality of compounds, determining those compounds which interact with and bind to the human 5-HT₄ receptor, and thereby identifying compounds which interact with, and specifically bind to, the human 5-HT₄ receptor.
- 20
- 25
- 30
76. The method of claim 75, wherein the mammalian cell is a non-neuronal cell.
- 35
77. A method for identifying a compound which is not known to be capable of binding to the human 5-HT₄ receptor activates the human 5-HT₄ receptor on the surface of a mammalian cell or prevents a ligand which does so, which comprises contacting the

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mammalian cell which cell comprises a plasmid adapted for expression in the mammalian cell such plasmid further comprising DNA which expresses the human 5-HT₂ receptor on the cell surface of the mammalian cell with the compound under conditions permitting activation or blockade of a functional response, determining whether the compound activates the human 5-HT₂ receptor or prevents a ligand which does so, and thereby identifying the compound as a compound which interacts with, and activates the human 5-HT₂ receptor or prevents the activation of the human 5-HT₂ receptor by a ligand which does so.

78. The method of claim 77, wherein the mammalian cell is a non-neuronal cell comprising the cellular components necessary to produce a second messenger and wherein the determination of whether the compound activates or blocks the activation of the human 5-HT₂ comprises detecting the change in the concentration of the second messenger.

79. The method of claim 78, wherein the second messenger is cyclic AMP (cAMP).

80. The method of claim 78, wherein the non-neuronal cell is a COS-7 cell.

81. A method of claim 78, wherein the second messenger is an inositol phosphate metabolite.

82. The method of claim 78, wherein the second messenger is intracellular calcium.

83. A compound identified by the method of claim 69, 73 or 77.

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84. A pharmaceutical composition of a drug identified by the method of claim 71 or 75.
- 5 85. A method for detecting the presence of a human 5-HT₄ receptor on the surface of a cell, which comprises contacting the cell with an antibody of claim 50, under conditions that permit binding of the antibody to the receptor, detecting the presence of any of the antibody bound to the cell, and thereby the presence of a human 5-HT₄ receptor on the surface of the cell.
- 10
86. A method for treating an abnormal condition related to an excess of activity of a human 5-HT₄ receptor, which comprises administering a patient an amount of a pharmaceutical composition of claim 84, effective to reduce 5-HT₄ activity as a result of naturally occurring substrate binding to and activating the 5-HT₄ receptor.
- 15
- 20 87. The method of treating abnormalities which are alleviated by an increase in the activity of a 5-HT₄ receptor, which comprises administering a patient an amount of a pharmaceutical composition of claim 84, effective to increase the activity of the 5-HT₄ receptor thereby alleviating abnormalities resulting from abnormally low receptor activity.
- 25
- 30 88. A method for diagnosing a predisposition to a disorder associated with the expression of a human 5-HT₄ receptor allele which comprises:
- 35 a. obtaining DNA from subjects suffering from a disorder;
- b. performing a restriction digest of the DNA

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with a panel of restriction enzymes;

c. electrophoretically separating the resulting DNA fragments on a sizing gel;

5

d. contacting the gel with a nucleic acid probe capable of specifically hybridizing to DNA encoding a human 5-HT₄ receptor and labelled with a detectable marker;

10

e. detecting the labelled bands which have hybridized to the DNA encoding 5-HT₄ receptor, labelled with the detectable marker to create a unique band pattern specific to the DNA of subjects suffering with the disorder;

15

f. preparing DNA for diagnosis by steps a-e;

20

g. comparing the unique band pattern specific to the DNA of patients suffering from the disorder from step e and DNA obtained for diagnosis from step f to determine whether the patterns are the same or different and to diagnose thereby predisposition to the disorder if the patterns are the same.

25

89. The method of claim 88, wherein a disorder is associated with the expression of a specific human 5-HT₄ receptor allele is diagnosed.

30

90. A method of identifying a substance capable of alleviating the abnormalities resulting from overexpression of a human 5-HT₄ receptor which comprises administering a substance to the transgenic non-human mammal of claim 67 or 68, and determining whether the substance alleviates the

35

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physical and behavioral abnormalities displayed by the transgenic nonhuman mammal as a result of overexpression of the human 5-HT₁ receptor.

5 91. A method of identifying a substance capable of
alleviating the abnormalities resulting from
underexpression of a human 5-HT₁ receptor, which
comprises administering a substance to the
10 transgenic mammal of claim 62, and determining
whether the substance alleviates the physical and
behavioral abnormalities displayed by the
transgenic nonhuman mammal as a result of
underexpression of the human 5-HT₁ receptor.

15 92. A method of treating abnormalities in a subject,
wherein the abnormality is alleviated by the
reduced expression of a human 5-HT₁ receptor which
comprises administering to a subject an effective
20 amount of the pharmaceutical composition of claim
55, 57, 83 or 84 effective to reduce expression of
the human 5-HT₁ receptor.

25 93. A method of treating abnormalities resulting from
underexpression of a human 5-HT₁ receptor which
comprises administering to a subject an amount of
a pharmaceutical composition of claim 56, 83 or 84
effective to alleviate abnormalities resulting
from underexpression of the human 5-HT₁ receptor.

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FIGURE 1-1

FIGURE 1-1, 1/32

FIGURE 1-2, 2/32

1	AGCCTGCCGAGCCTGGCTTGGTTGGAAGGAGGAGGATGCTCGGTGCCAGGGTCCIG	60
61	TGGGCACIGACATCCAACGTACTCATGCCCATTTCTGTATGGACAGACTTGAIGCTAA	120
	M D R L D A N	7
121	TGTGAGTTCCAACGAGGGTTTCGGGTCIGTGGAGAAGGTCGTACIGTCACGTTCITCGC	180
8	V S S N E G F G S V E K V V L L T F F A	27
181	AATGGTATCCIGATGGCCATCCCTGGGCAACCCTGCTGGTGATGGTTGCTGTGTCAGGGA	240
28	M V I L M A I L G N L L V M V A V C R D	47
241	CAGGCAGCTCAGGAAAATAAAACCAATTATTCATTTGTCITCTTGCCITTCIGATCT	300
48	R Q L R K I K T N Y F I V S L A F A D L	67
301	GCTGGTTTCGGIGCTGGIGAATGCCCTTCGGTGCCATTGAGTTGGTCCAGACATCTGGTT	360
68	L V S V L V N A F G A I E L V Q D I W F	87
361	TIATGGGgagatgtttgcctggtgctcgacccctctctggtgctcctactcaccacagcatc	420
88	Y G E M F C L V R T S L D V L L T T A S	107
421	batcttcacccctgctgcatctccctGGATAGGATTAATGCCATCTGCTGTCACCTTT	480
108	I F H L C C I S L D R Y Y A I C C Q P L	127
481	GGTTTATAGAAACAAGATGACCCCTCTACGCATCGCATTAAATGCTGGGAGGCTGCTGGGT	540
128	V Y R N K M T P L R I A L M L G G C W V	147
541	CATCCCAATGTTATATCTTTCTCCCCATAATGCAAGGCTGGAACAACATCGGCATAGT	600
148	I P M F I S F L P I M Q G W N N I G I V	167
601	TGATGTAGAGAAAGGAATTCACCAACTCTACTCTACATTTCTGTCTTCAT	660
168	D V I E K R K F N H N S N S T F C V F M	187
661	GGTCAACAAGCCCTATGCCATCACCTGCTCIGTGGGCTTCTACATCCCGTTTCICCT	720
188	V N K P Y A I T C S V V A F Y I P F L L	207

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FIGURE 1-2

721	CATGGTGGCCATTACCGTATCTATGCTACTGCTAAGGAGCATGCCCGCAGATCCA	780
208	M V L A Y Y R I Y V T A K E H A Q Q I Q	227
781	GATGTIACAACGGGCGAGGCCACCTCTGAAGCAGGCCCCAGACAGCTGACCCAGCACAG	840
228	M L Q R A G A T S E S R P Q T A D Q H S	247
841	CACACATCGCATGGGACAGACAGACCAAGCAGCCCAAGACTTATGTGTCATCATGGGCTG	900
248	T H R M R T E T K A A K T L C V I H G C	267
901	CTTCGTTCGTGGGCCCCCTTCCTTGTGTCACCAATATTGTGGACCCCTTTCATAGACTA	960
268	F C F C W A P F F V T N I V D P F I D Y	287
961	CACGTGCCCCGAGAAGGTGTGGACTGCTTCCTCTGGCTTGGCTATATCAATTCAGGGTT	1020
288	T V P E K V W T A F L W L G Y I N S G L	307
1021	GAACCCCTTTCCTATGCTTCCTTGAATAAGTCTTTCAGACGTCCTTCCTTATCATCCT	1080
308	N P F L Y A F L N K S F R R A F L I I L	327
1081	CTGCTGTGATGAGCGCTACAAAGACCCCTTCCTGGCCAGACTGTCCCCTGTTC	1140
328	C C D D E R Y K R P P I L G Q T V P C S	347
1141	AACCACAACCATTAATGGATCCACTCACTGCTAAGGTATACAGTTTGCATAGIGTCA	1200
348	T T T I N G S T H V L R Y T V L H S G Q	367
1201	ACACGAGAACTGGAGAAGTTGCCCATACACAAATGACCCAGAGTCCCTGGAAATCATGCTT	1260
368	H Q E L E K L P I H N D P E S L E S C F	387
1261	TTGATTGAAGACGTGGCTTGGCTTAGCGGTTTCATCCCATCTGTGTGTCATGAACAGGT	1320
	★	
1321	TACTATGGAATCACCTCTGACTCTGGGCATCACCCAGTGAAGCATGAGCATGGTGAGGCAG	1380
1381	GGTCCGGTGAAGGTGCACAGAGGACAGCATTGAGTGGACCTGAACCCAGCACATTAAAGG	1440
1441	ATTTCAGAACCGTGTGGGATTGAGATGTATCAGACCCAGTGTCTTACCCAGAGCCCA	1500
1501	ACTGGCACCTCCCATTCACGCTGACATGTGGTCAGTCTTGTCTCACACCTCTCCAGGGG	1560
1561	CAGGAGCTGACTACCTCCTAATGTGGTGGGAGCTCTTAATTGTGTGGAAGTTCAGTCAT	1620
1621	TCATTGGTGGACAGTCTCGCTG	1642

FIGURE 2-1, 3/32

FIGURE 2-2, 4/32

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FIGURE 2-1

1	CAGCGCAGCAGCGGCAGAGCAGGGCTCTGGCTAAAGGTGGCCGACGGAATCTGCC	60
61	AGGTGCEGCTCTCTTAGCCINCGCAGCCGGCTGGGCTGGAGTGCokGCCACGAGA	120
121	TCAGAACCCAGGAGCCACGTGTGAAGGTCCCTGTGGCACTGACATCCAACGTACTCA	180
181	TGCCCATTTCCIGTAAIGGACAGACTTGATGCTAATGTGAGTTCACACGAGGTTTCGGG	240
	M D R L D A N V S S N E G F G	15
241	TCGTGGAGAGGTGCTACTGCTCAGCTTCTTCGCAATGGTATCCIGATGGCCATCCTG	300
16	S V E K V V L L T F F A M V I L M A I L	35
301	GGCAACCTGCTGGTGAITGGTTCGTGTGTCAGGACAGGCAGCTCAGGAAATAAAACC	360
36	G N L L V M V A V C R D R Q L R K I K T	55
361	AATTATTTTCATTTGCTCTCTTGCCTTTGCTGATCIGCTGGTTCGGTGGTGAATGCC	420
56	N Y F I V S L A F A D L L V S V L V N A	75
421	TTCGGTGCCATIGAGTIGGTCGAAGACATCTGGTTTATGGGAGATGTTTGGCTGGTC	480
76	F G A I E L V Q D I W F Y G E M F C L V	95
481	CGGACCTCTCTGGATGCTCTACTCACCACGATCAATTTTACCCTCTGCTGCCCTTCC	540
96	R T S L D V L L T T A S I F H L C C I S	115
541	CTGGATAGGTATTATGCCATCTGCTGTCACCTTTGGTTTATAGAAACAAGATGACCCCT	600
116	L D R Y Y A I C C Q P L V Y R N K M T P	135
601	CTACGCATCGCATTAAATGCTGGAGGCTGCTGGGTCATTCCTCATGTTTATCTTTCTC	660
136	L R I A L M L G G C W V I P M F I S F L	155
661	CCCATATGCAAGGCTGGAACAACATCGGCATAGTGTGATAGAGAAAGGAAATTC	720
156	P I M Q G W N N I G I V D V I E K R K F	175
721	AACCACACTTAACCTACATCTGCTGCTTTCATGGTCAACAGGCCCTATGCCATCACC	780
176	N H N S N S T F C V F M V N K P Y A I T	195
781	TGCTCTGTGGCTTCTACATCCCGTTTCTCTCATGGTGGCTGCTATACCGTATC	840
196	C S V V A F Y I P F L L M V L A Y Y R I	215

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FIGURE 2-2

841	TATGTCAC	TGCTAAGG	AGCATGCC	CCAGCAG	ATCCAG	ATGTTACA	ACGGG	CAGG	AGCCACC	900
216	Y V T A	K E H A	Q Q I Q	M L Q	R A G	A T				235
901	TCTGAAG	CAGGCC	CCAGCAG	CTGACC	AGCAGC	ACACAT	CGCATC	GGGAC	GAGAGCC	960
236	S E S R	P Q T A	D Q H S	T H R M	R T E T					255
961	AAAGCAG	CCAAGAC	TTAIGT	GTCA	TATGGG	CTGCTT	CTGTTT	CTGCTG	GGCCCCC	1020
256	K A A K	T L C V	I M G C	F C F C	W A P F					275
1021	TTTGTCA	CCAATAT	TGTGGAC	CCCTT	CATAGAC	TAACAT	GTGCCC	GAGAGG	GTGGACT	1080
276	F V T N	I V D P	F I D Y	T V P E	K V W T					295
1081	GCITTC	CTCTGG	CTATG	CAATTC	AGGGT	TGAACC	CTTTCT	CTATGC	CTTCTTG	1140
296	A F L W	L G Y I	N S G L	N P F L	Y A F L					315
1141	AATAAG	CTTTCAG	ACGTC	CTTCTT	ATCATC	CTCTG	CTGIG	ATGATG	AGCGCTAC	1200
316	N K S F	R R A F	L I I L	C C D D	E R Y K					335
1201	AGACCC	CCCAT	TTCTGG	CCAGAC	TGTC	CCCTGT	CAACCA	CAACCA	TTAATG	1260
336	R P P I	L G Q T	V P C S	T T I T	I N G S	T				355
1261	CATGTG	CTAAGG	GATAC	AGTGG	AATGTG	TGGCCA	ATGGG	AGAGT	CGGTGTC	1320
356	H V L R	D T V E	C G G Q	W E S R	C H L T					375
1321	GCAACT	CTCTCT	TGGTGG	CTGCTC	AGCCAG	TGATAC	GTAGG	CCCCC	AGGACA	1380
376	A T S P	L V A A	Q P V I	R R P Q	D N D L					395
1381	GAAGAC	AGCTGT	AGCTTGA	AAAGAG	CCAGTC	CTAAGC	TGCTAC	TTCCG	GTATG	1440
396	E D S C	S L K R	S Q S *							406
1441	GCCCCI	GGCAC	TTGTTC	CCAAGG	CTTCCA	AGAGC	ATGAGG	CAATCC	ACCCCTG	1500
1501	CCCGCC	ACGAT	CTAGC	AGGCG	TATTAG	AGGAAG	TCAGG	GAGAG	AGGGCTC	1560
1561	TAGCTT	CTGTT	CTCAAC	ATTTCT	CTTCC	TGGAG	CTCCAC	CTCTG	TTGGTGG	1620
1621	TGAAGT	CCAGC	ACCCAG	TCCCC	TTTGC	TGCTC	CCAGT	CTGCTG	TAAATG	1680
1681	GTTGAT	TTTCAG	TTTCCA	AACATG	CCCTT	CTTTGA	AGTGT	CACTCT	TACGATA	1740
1741	ACATGT	CCCTG	CTGCTG	ATCAC	ACTTCT					1768

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FIGURE 3

1 MDRLDANVSSNEGFGSVEKVLLTFFAMVILMAILGNLLVMVAVCRDRQL 50
|||||
1 MDRLDANVSSNEGFGSVEKVLLTFFAMVILMAILGNLLVMVAVCRDRQL 50

51 RKIKTNYFIVSLAFADLLVSVLVNAFGAIELVQDIWFGEMFCLVRTSLD 100
|||||
51 RKIKTNYFIVSLAFADLLVSVLVNAFGAIELVQDIWFGEMFCLVRTSLD 100

101 VLLTTASIFHLCCISLDRYYAICCPVYRNKMTPLRIALMLGGCWVIPM 150
|||||
101 VLLTTASIFHLCCISLDRYYAICCPVYRNKMTPLRIALMLGGCWVIPM 150

151 FISFLPIMQGWNIGIVDVEIKRKFNHNSNSTFCVFMVNKPYAITCSVVA 200
|||||
151 FISFLPIMQGWNIGIVDVEIKRKFNHNSNSTFCVFMVNKPYAITCSVVA 200

201 FYIPFLLMVLAYRYIYVTAKEHAQQIQHLQAGATSESRPQTADQHSTHR 250
|||||
201 FYIPFLLMVLAYRYIYVTAKEHAQQIQHLQAGATSESRPQTADQHSTHR 250

251 MRTETKAAKTLCVINGCFCFCWAPFFVTNIVDPFIDYTVPEKVWTAFLWL 300
|||||
251 MRTETKAAKTLCVINGCFCFCWAPFFVTNIVDPFIDYTVPEKVWTAFLWL 300

301 GYINSGLNPFLYAFLNKSFRRAFLIILCCDDERYKRPPILGQTVPCSTTT 350
|||||
301 GYINSGLNPFLYAFLNKSFRRAFLIILCCDDERYKRPPILGQTVPCSTTT 350

351 INGSTHVLRYTVLHSGQ.....HQELEKLPIHNDPESLES 385
||||| :|| :|| :|| :|| :|| :||
351 INGSTHVL RDTVECGGQWESRCHLTATSPLVAAQPVI RRPQDNLE..DS 398

386 CF 387
|
399 CSLKRSQS 406

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FIGURE 4-1

FIGURE 4-1, 6/32
FIGURE 4-2, 7/32
FIGURE 4-3, 8/32

S10-95	MMDVNSSGRPDLYGHLRSFLLPEVGRGLPDLSPDGGADPPVAGSWAPHLLS	0
S10-87	MEILCEDNISLSSSIPNSLMQLGCGPRLYHNDFNSR	0
Hp78		50
5-HT2 Rat	MVNLGNNAVRSLLMHLIGL	35
5-HT1C Rat		18
Hist2 Dog		0
S10-95	MDRLDANVSSMEGFGSVE	28
S10-87	MDRLDANVSSMEGFGSVE	28
Hp78	EVTASPAPTWDAPPDNASGCEQIN YGRVE	90
5-HT2 Rat	DANTSEASNVTIDAENRTMLSCCEGYLPPTCLSLHLQE	83
5-HT1C Rat	LVWQFDISISPVAA	63
Hist2 Dog	MISMGITGSSSFCLD\$PPC	27
S10-95	VILMAIILBHLVLVMVAVCGRRQLRKKIKTIN YFFIVSLAFADLLVSVLVNAFQA	78
S10-87	VILMAIILGNLLVMVAVCGRRQLRKKIKTIN YFFIVSLAFADLLVSVLVNAFQA	78
Hp78	ITLTLIAQNCILVIMAVSVCFVKKLRQ.PSNHYLLIVSLALADLSVAVAVMPPVVS	139
5-HT2 Rat	VILTLIAQNCILVIMAVSVCFVKKLRQ.ATNHYFLMGLAIADMLVGLLVMPVSM	132
5-HT1C Rat	ILITMTIGGNILVIMAVSVCFVKKLRQ.ATNHYFLMGLAIADMLVGLLVMPVSM	112
Hist2 Dog	LILITIAQNCILVIMAVSVCFVKKLRQ.ATNHYFLMGLAIADMLVGLLVMPVSM	76

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FIGURE 4-2

S10-95	I . E L V Q Q D I W F Y G E M F C L V R T S L D V L L T T A S I F H L C C I S L D R Y Y A I C C Q P L	127
S10-87	I . E L V Q Q D I W F Y G E M F C L V R T S L D V L L T T A S I F H L C C I S L D R Y Y A I C C Q P L	127
Hp78	V T D L I G G K W I F G H F F C N Y F I A M D V M C C T A S I M T L C V I S I D R Y L G I . T R P L	188
5-HT2 Rat	L T I L Y G Y R W P L P S K L C A I W I Y L D V L F S T A S I M H L C A I S L D R Y V A I Q . N P I	181
5-HT1C Rat	L A I L Y D Y V W P L P R Y L C P Y W I S L D V L F S T A S I M H L C A I S L D R Y V A I R . N P I	161
Hist2 Dog	F . Y Q L S C R W S F G K V F C N I Y T G L D V M L C T A S I L N L F M I S L D R Y C A V . T D P L	124
III		
S10-95	V Y R N K M T P L R I A L M L B G C U V I P M F I S F L P I M Q Q W H N I B I V D V I E K R K F H H	177
S10-87	V Y R N K M T P L R I A L M L B G C U V I P M F I S F L P I M Q Q W H N I B I V D V I E K R K F H H	177
Hp78	T Y P V R Q N G K C H A K M I L S V H L L S A S I T L P P L F . G W A Q N	224
5-HT2 Rat	H H S R F N S R T K A F L K I I A V W T I S V G I S M P I P V F G L Q D S K V F . K E G	225
5-HT1C Rat	E H S R F N S R T K A I M K I A I V W A I S I G V S S V P I P V I G L R D E S K V F V M N T	206
Hist2 Dog	R Y P V L I T P V R V A V S L V L I W V I S I T L G F L S T H L G W H S R N E T S S F H H	169
IV		
S10-95	N S N S T F C V F M V N K P Y A I T C S V V A F Y I P F L L M V L A Y Y R I Y Y T A K E H A	223
S10-87	N S N S T F C V F M V N K P Y A I T C S V V A F Y I P F L L M V L A Y Y R I Y Y T A K E H A	223
Hp78	V N D D K V C L I S Q D F G Y T I Y S T A V A F Y I P M S V M L F M Y Y Q I Y K A A R K S A	270
5-HT2 Rat S C L L A D D N F V L I G S F V A F F I P L T I M V I T Y F L T I K S L Q K E A T L C V	269
5-HT1C Rat T C V L N D P N F V L I G S F V A F F I P L T I M V I T Y F L T I Y V L R R Q I L M L L	250
Hist2 Dog	T I P K . . C K V Q V N L V Y G L V D G L V T F Y L P L L V M C I T Y Y R I F K I A R D Q A	213
V		
S10-95 Q Q I Q M . . L Q R A G A T S E S R P Q T A D Q	245
S10-87 Q Q I Q M . . L Q R A G A T S E S R P Q T A D Q	245
Hp78 A K H K F P G F P R V E P D S V I A L N G I V K . . L Q K E V E E C A N L S R L L K H E R	313
5-HT2 Rat	S D L S T R A K L A . . S F S F L P Q S S L S S E K L F Q R S I H R E P G S Y A G R R	310
5-HT1C Rat	R G . H T E E E L A N M S L N F L N C C C K K N G G E E E N A P N P D Q K P R R K K E K R P R	299
Hist2 Dog K R I H H . M G S W K A A T I G E	229
VI		

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FIGURE 4-3

S10-95	H S T H R R M R Y E T K A A K T L C V I M G C F C F C H A P F F V T M I V D P F I D Y T V P E K V . .	293
S10-87	H S T H R R M R Y E T K A A K T L C V I M G C F C F C H A P F F V T M I V D P F I D Y T V P E K V . .	293
Hp78	K N I S I F K R E Q K A A T T L G I I V G A F T V C W L P F F L L S T A R P F I C G T S C S C I P L	363
S-HT2 Rat	. T M Q S I S M E Q K A C K V L G I V F F L F V V M W C P F F I T M I M A V I C K E S C M E N V I G	359
S-HT1C Rat	G T M Q A I N E F K A S K V L G I V F F F L I M W C P F F I T M I L S V L C G K A C M Q K L M E	349
Hist2 Dog	H K A T V T L A A V N G A F I I C W F P Y F T V F Y R G L K G D A I N E A . .	268

	VII																
S10-95	WTA	FLWL	GYIN	SGLN	PF	LYA	FLN	KSFR	RAFL	IL	CCDD	ERYK	RPPI	LG	341		
S10-87	WTA	FLWL	GYIN	SGLN	PF	LYA	FLN	KSFR	RAFL	IL	CCDD	ERYK	RPPI	LG	341		
Hp78	WVER	TF	WL	GL	YAN	SLIN	PF	IYA	FFNR	DL	RTTY	RS	SL	LC	402		
5-HT2 Rat	ALLN	VFW	WIG	YLS	SAVN	PLV	YTL	LFNK	TYRS	SA	FS	RYI	QC	QYKE	NRKPLQLI	409	
5-HT1C Rat	KLLN	VFW	WIG	YVCS	GIN	PLV	YTL	LFNK	IYRR	RA	FS	SKY	LR	CDYK	PPDXXKP	398	
Hist2 Dog	FEA	VVL	WL	GL	YAN	SLIN	PI	LYAT	ILNR	DF	RTAY	QQ	LF	...	RCRPA	SHMA	312

	S10-95	S10-87	Hp78	5-HT2 Rat	5-HT1C Rat	Hist2 Dog																																								
Q	T	V	P	C	S	T	T	I	N	G	S	T	H	V	L	..	R	D	T	V	E	C	G	G	R	W	E	S	R	C	H	L	T	A	T	S	P	L	V	A	Q	P	V	I	R	388
Q	T	V	P	C	S	T	T	I	N	G	S	T	H	V	L	..	R	D	T	V	E	C	G	G	R	W	E	S	R	C	H	L	T	A	T	S	P	L	V	A	Q	P	V	I	R	376
Q	T	V	P	C	S	T	T	I	N	G	S	T	H	V	L	..	R	D	T	V	E	C	G	G	R	W	E	S	R	C	H	L	T	A	T	S	P	L	V	A	Q	P	V	I	R	440
Q	T	V	P	C	S	T	T	I	N	G	S	T	H	V	L	..	R	D	T	V	E	C	G	G	R	W	E	S	R	C	H	L	T	A	T	S	P	L	V	A	Q	P	V	I	R	459
Q	T	V	P	C	S	T	T	I	N	G	S	T	H	V	L	..	R	D	T	V	E	C	G	G	R	W	E	S	R	C	H	L	T	A	T	S	P	L	V	A	Q	P	V	I	R	448
Q	T	V	P	C	S	T	T	I	N	G	S	T	H	V	L	..	R	D	T	V	E	C	G	G	R	W	E	S	R	C	H	L	T	A	T	S	P	L	V	A	Q	P	V	I	R	354

S10-95	R P Q D N D L E D G C S L K R S Q S .	406
S10-87	L P I H N D P E S L E S C F	387
Hp78	K G H D S	445
5-HT2 Rat	N I E Y V N E K V S C V	471
5-HT1C Rat	P S N V V S E R I S S V	460
Hist2 Dog	G A T D R	359

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FIGURE 5

574	TTGGTCTATAGGAACAAGATGACCCCTCTGGCGATCGCATTAATGCTGGAGGCTGCTGG	633
127	L V Y R N K M T P L R I A L M L G G C W	146
634	GTCATCCCCACGTTTATTCTTTCTCCCTATAATGCAAGGCTGGAATAACATTGGCATA	693
147	V I P T F I S F L P I M Q G W N N I G I	166
694	ATTGATTGTAGAAAGAGGAAGTTCAACCAGAACTCTAACTCTACGTACTGTGTCTTC	753
167	I D L I E K R K F N Q N S N S T Y C V F	186
754	ATGGTCAACAAGCCCTACGCCATCACCTGCTGTGTGGCCTTCTACATCCCATTTCTC	813
187	M V N K P Y A I T C S V V A F Y I P F L	206
814	CTCATGGTGGCCCTATTACCGCATCTATGTCACAGCTAAGGAGCATGCCCATCAGATC	873
207	L M V L A Y Y R I Y V T A K E H A H Q I	226
874	CAGATGTTACAACGGGAGGAGCCCTCCCGAGAGCAGGCCCTCAGTCGGCAGACCAGCAT	933
227	Q M L Q R A G A S S E S R P Q S A D Q H	246
934	AGCACTCATCCGATGAGGACAGAGACCAAGCAGCACCCTGTGCATCATCATGGGT	993
247	S T H P M R T E T K A A K T L C I I M G	266
994	TGCTTCTGCCCTCTGCTGGGCACCATTTCTTGTCAACCAATATTGTGGATCCTTTCATAGAC	1053
267	C F C L C W A P F F V T N I V D P F I D	286
1054	TACACTGTCCCTGGGCAGGTGTGGACTGCTTCTCTGGCTCGGCTATATCAATTC	1109
287	Y T V P G Q V W T A F L W L G Y I N	304

FIGURE 6

574 TTGGTCTATAGGAACAAGATGACCCCTCTGCGCATCGCATTAATGCTGGAGGCTGCTGG 633
 T A A
 634 GTCATCCCCACGTTATTCTTTCTCCCTATAATGCAAGGCTGGAATAACATTGGGCATA 693
 T T A C C
 694 ATTGATTGTAGAAAAGAGGAAGTTCAACCAGAACTCTAACTCTACGTACTGTGTCTTC 753
 G G A A C A T
 754 ATGGTCAACAAGCCCTACGCCATCACCTGCTCTGTGGTGGCCCTTCTACATCCCATTCTC 813
 T G
 814 CTCATGGTGTGGCCCTATTACCGCATCTATGTCAACAGCTAAGGAGCATGCCCATCAGATC 873
 T T G
 874 CAGATGTTACAACGGGCAGGAGCCCTCCTCCGAGAGCAGGCCCTCAGTCGGCAGACCAGCAT 933
 A T A C A A T C
 934 AGCACTCATCCGATGAGGACAGAGACCACCAAGCAGCCAGACCCCTGTGCATCATCATGGGT 993
 A GC C TT A TG C
 994 TGCTTCTGCCCTCTGCTGGGCACCATTTCTTGTCAACCAATATTGTGGATCCTTTCATAGAC 1053
 TT C C
 1054 TACACTGTCCCTGGGCAGGTGTGGACTGCTTCCCTCTGGCTCGGCTATATCAATTC 1109
 G C A A T

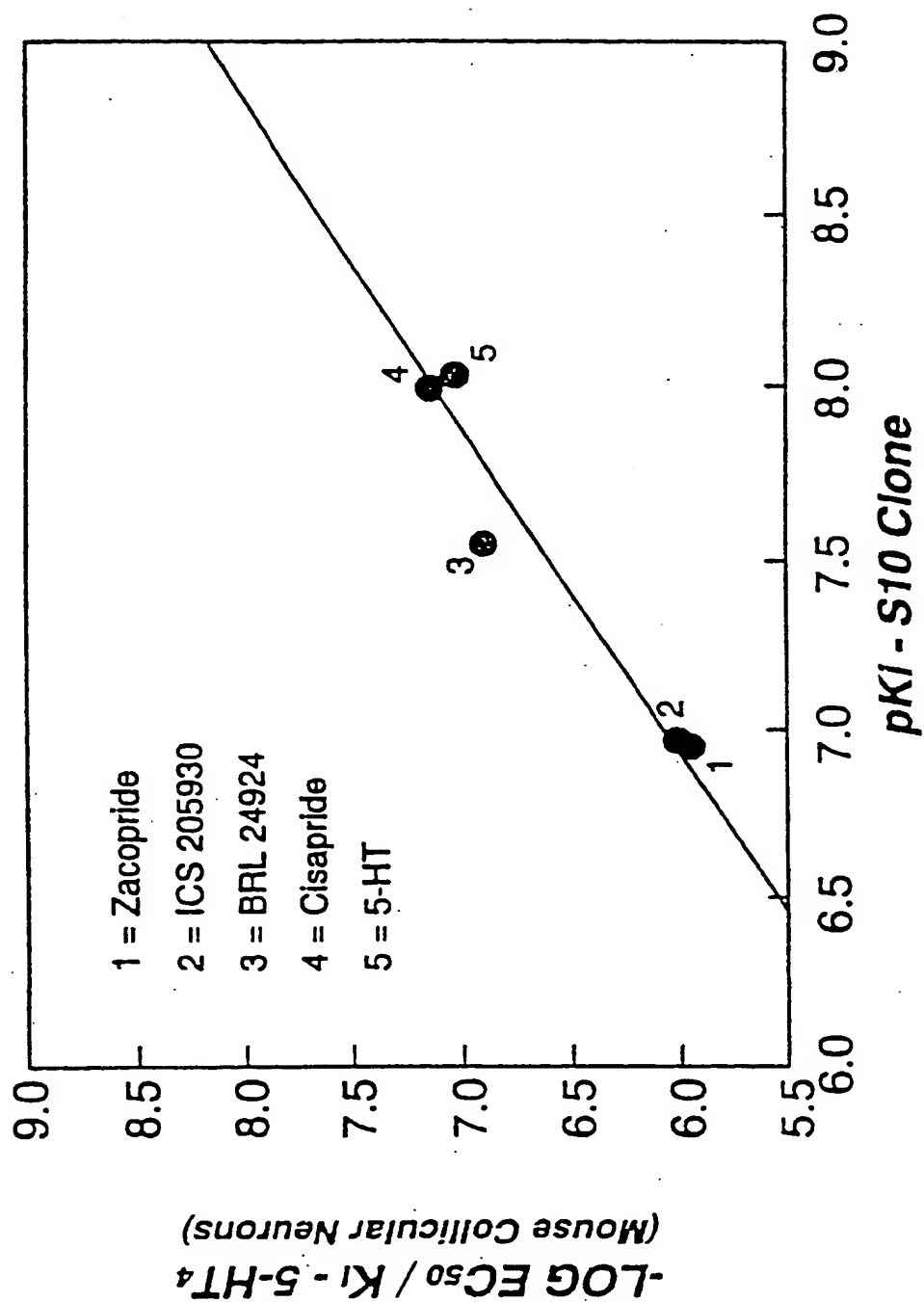
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1127	LVYRNKMTPLRIALMLGGCWV	IPTFISFLPIMQGWNNIGI	IDLIEKRKFQNSNSTYCVF	186
		M	V V H F	
1187	MVNKPYAITCSVVAFYIPFL	MLVLA YYRIYVTAKEHAHQ	IQLQRACASSESRPQSAQHQ	246
		Q	T T	
247	STHPMRTEKAAKTLCIIMGC	FCLCWAPFFVTNIVDPFIDY	TVPGQVWTAFLWLGYIN	304
	R	V F	EK	

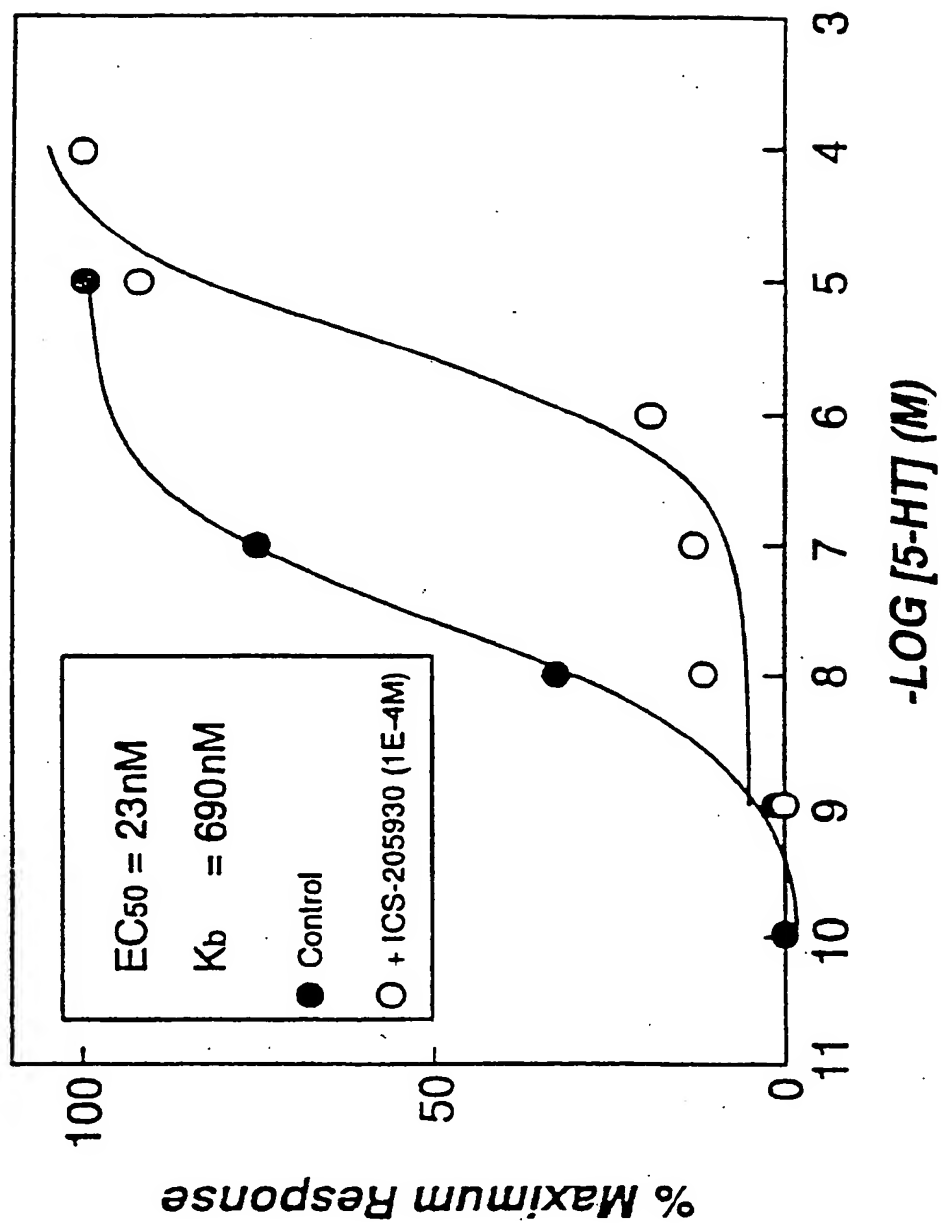
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FIGURE 8



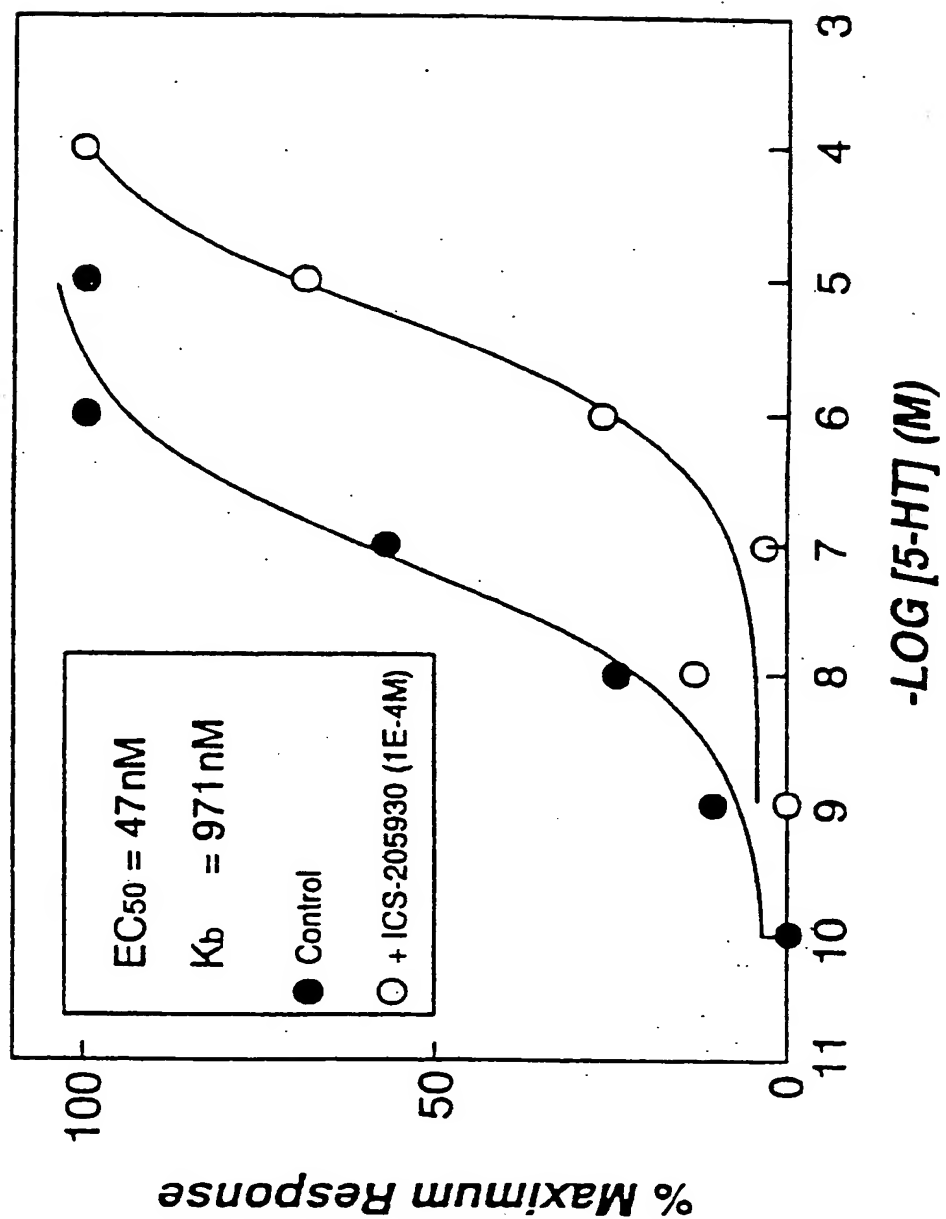
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FIGURE 9



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FIGURE 10



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FIGURE 11A

1	TTGGTCTATAGGAACAAGATGACCCCTCTGCGCATCGCATTAAATGCTGGAGGCTGCTGG	60
61	GTCATCCCCACGTTTATTCTCTCCCTATAATGCAAGGCTGGAATAACATTTGGCATA	120
121	ATTGATTTGATAGAAAAGAGGAAGTTCAACCAAGAACTCTAACTCTACGTACTGTGTCTTC	180
181	ATGGTCAACAAGCCCTACGCCATCACCTGTCTGTGTGGCTTCTACATCCCATTTCTC	240
241	CTCATGGTGTGGCTATTACCGCATCTATGTCACAGCTAAGGAGCATGCCCATCAGATC	300
301	CAGATGTTACAACGGGAGGAGCTCTCCGAGAGCAGGCTCAGTCGGCAGACCAGCAT	360
361	AGCACTCATCGCATGAGGACAGAGACCAAGCAGCAAGACCTGTGCATCATATGGGT	420
421	TGCTTCTGCCCTGTCTGGGCACCATTTCTTGTCAACCAATATTGTGGATCCTTTCATAGAC	480
481	TACACTGTCCCTGGGAGGTGTGACTGTCTTCTCTGGCTCGGCTATATCAATTCCGGG	540
541	TTGAACCCCTTTCTCTACGCCCTTCTTGAATAAGTCTTTTAGACGTGCCTTCTCATCATC	600
601	CTCTGTGTGATGATGAGCGCTACCGAAGACCTTCCATTCTGGGCCAGACTGTCCCTTGT	660
661	TCAACCACAACCATTAATGATCCACACATGTAAGGTACACCGTTCTGTGCACAGGGA	720
721	CATCATCAGGAACCTCGAGAACTGCCCATACACAATGACCCAGAATCCCTGGAATCATGC	780
781	TTCTGATTGAGG	

FIGURE 11B

1	L	V	I	D	V	M	M	T	F	T	N	C	Y	L	L	S	H	F
21	V	I	D	V	M	M	T	F	T	N	C	Y	L	L	S	H	F	
41	I	M	L	Q	S	C	Y	L	L	S	H	F						
61	M	L	Q	S	C	Y	L	L	S	H	F							
81	L	Q	S	C	Y	L	L	S	H	F								
101	Q	S	C	Y	L	L	S	H	F									
121	S	C	Y	L	L	S	H	F										
141	C	Y	L	L	S	H	F											
161	Y	L	L	S	H	F												
181	L	L	S	H	F													
201	L	L	S	H	F													
221	L	L	S	H	F													
241	S	H	F															
261	F																	

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FIGURE 12-1

FIGURE 12-1, 16/32
FIGURE 12-2, 17/32
FIGURE 12-3, 18/32

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1 .....TTGGTCTATAGGAACAAGATGA 22
   ||||| ||||| ||||| |||||
451 TAGGTATTATGCCATCTGCTGTCAACCTTTGGTTTATAGAAACAAGATGA 500

23 CCCCTCTGCGCATCGCATTAAATGCTGGGAGGCTGCTGGGTCAATCCCCACG 72
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
501 CCCCTCTACGCATCGCATTAAATGCTGGGAGGCTGCTGGGTCAATCCCATG 550

73 TTTATTTCTTTTCTCCCTATAATGCAAGGCTGGAATAACATTGGCATAAT 122
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
551 TTTATATCTTTTCTCCCATATAATGCAAGGCTGGAACAACATCGGCATAGT 600

123 TGATTTGATAGAAAAGAGGAAGTTCAACCAGAACTCTAACTCTACGTACT 172
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
601 TGATGTGATAGAGAAAAGGAAATTCAACCACAACCTCTAACTCTACATTCT 650

173 GTGTCTTCATGGTCAACAAGCCCTACGCCATCACCTGCTCTGTGGTGGCC 222
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
651 GTGTCTTCATGGTCAACAAGCCCTATGCCATCACCTGCTCTGTGGTGGCC 700

223 TTCTACATCCCATTTTCTCCTCATGGTGCTGGCCCTATTACCGCATCTATGT 272
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
701 TTCTACATCCCGTTTCTCCTCATGGTGCTGGCCCTATTACCGTATCTATGT 750
```


FIGURE 12-3

573	GTCTTTTAGACGTGCCTTCCTCATCTCATCTCTGTGTGATGATGAGCGCT	622
1051	GTCTTTTCAGACGTGCCTTCCTTATCATCTCTCTGTGTGATGATGAGCGCT	1100
623	ACCGAAGACCTTCCATTCTGGGCCAGACTGTCCCTTGTTCAACCAACAAC	672
1101	ACAAAGACCCCCATTCTGGGCCAGACTGTCCCTGTTCAACCAACAAC	1150
673	ATTAATGGATCCACACATGTACTAAGGTACACCGTTCTGCACAGGGGACA	722
1151	ATTAATGGATCCACTCATGTGCTAAGGTATACAGTTTTCATAGTGGTCA	1200
723	TCATCAGGAACTCGAGAACTGCCCCATACACAATGACCCAGAAATCCCCTGG	772
1201	ACACCAGGAACTGGAGAAAGTTGCCCATACACAATGACCCAGAGTCCCCTGG	1250
773	AATCATGCTTCTGATTGAGG.....	792
1251	AATCATGCTTTTGATTGAAGACGTGGCTTGCCCTTAGCGGTTTCATCCCAT	1300

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FIGURE 13

1 LVYRNKMTPLRIALMLGGCWVPT 24
|||
101 VLL'TTASIFHLCCISLDRYYAICCCQLVYRNKMTPLRIALMLGGCWVPM 150
|||
25 FISFLPIMQGWNNIGIIDLIEKRKFENQNSNSTYCVFMVNKPYAITCSVVA 74
|||
151 FISFLPIMQGWNNIGIVDIEKRKFENHNSNSTFCVFMVNKPYAITCSVVA 200
|||
75 FYIPFLLMVLAYYRIYVTAKEHAHQIQMLQRAGASSESRPQADQHS THR 124
|||
201 FYIPFLLMVLAYYRIYVTAKEHAQQIQMLQRAGATSESRPQTADQHS THR 250
|||
125 MRTETKAAKTLCIIMGCFCLCWAPFFVTNIVDPFIDYTVPGQVWTAFLWL 174
|||
251 MRTETKAAKTLCVIMGCCFCFCWAPFFVTNIVDPFIDYTVPEKVWTAFLWL 300
|||
175 GYINSGLNPFLYAFLNKSFRRAFLIILCCDDERYRRPSILGQTVPCSTTT 224
|||
301 GYINSGLNPFLYAFLNKSFRRAFLIILCCDDERYKRPPILGQTVPCSTTT 350
|||
225 INGSTHVLRYTVLHRGHQHELEKLPINHDPESLESCF* 262
|||
351 INGSTHVLRYTVLHSGQHQHELEKLPINHDPESLESCF* 387
|||

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FIGURE 14A, 20/32
FIGURE 14B, 21/32

FIGURE 14A

1	CCTGTAATGGACAAACTTGATGCTAATGTGAGTTCTGAGGAGGGTTTCGGGTCAAGTGGAG	60
61	AAGGTGGTGCTGCTACGTTTCTCTCGACGGTTATCCTGATGGCCATCTTTGGGGAACCTG	120
121	CTGGTGATGGTGGCTGTGCTGGACAGGCAGCTCAGGAAATAAAACAATTAATTC	180
181	ATTGTAATCTCTTGTCTTTTGGCGATCTGCTGGTTTTCGGTGCTGGTGATGCCCTTTGGTGCC	240
241	ATTGAGCTGGTTCAAGACATCTGGATTATGCGGAGGTGTTTGTCTTGTTCGGACATCT	300
301	CTGGACGTCCTGCTCACAAACGGCATCGATTTTTCACCTGTGCTGCATTTCTCTGGATAGG	360
361	TATTACGCCATCTGCTGCCAGCCTTTTGGTCTATAGGAACAAGATGACCCCTCTGCGCATC	420
421	GCATTAAATGCTGGGAGGCTGCTGGGTCAATCCCAACGTTTATTTCTTTTCTCCCTATAATG	480
481	CAAGGCTGGAATAACATTGGCATAATTGATTGATAGAAAGAGGAAGTTCAACCAGAAC	540
541	TCTAACTCTACGTACTGTGTCTTCAATGGTCAACAAGCCCTACGCCATCACCTGCTCTGTG	600
601	GTGGCCTTCTACATCCCATTCTCCTCATGGTGCTGGCCTATTACCGCATCTATGTCACA	660
661	GCTAAGGAGCATGCCCATCAGATCCAGATGTTACAACGGCAGGAGCCTCCTCCGAGAGC	720
721	AGGCCCTCAGTCGGCAGACCATAGCACTCATCGCATGAGGACAGAGACCAAGCAGCC	780
781	AAGACCCCTGTGCATCATATGGGTGCTTCTGCTCTGCTGGCACCATTTCTTTGTCAAC	840
841	AATATTGTGGATCCCTTTCATAGACTACACTGTCCCTGGCAGGTGTGACTGCTTTCCCTC	900
901	TGGCTCGGCTATATCAATTCCGGGTTGAACCCCTTTTCTCTACGCCCTTCTTGAATAAGTCT	960
961	TTTAGACGTGCCCTTCTCATATCCTCTGCTGTGATGATGAGCGCTACCGAAGACCTTCC	1020
1021	ATTCTGGGCCAGACTGTCCCTTGTCAACCACCAACCATTAATGGATCCACACATGTACTA	1080
1081	AGGGATGCAGTGGAGTGTGTGGTGGCAGTGGAGAGTCAAGTGTACCCGCCAGCAACTTCT	1140
1141	CCTTTGGTGCTGCTCAGCCCAAGTGACACTTAGGCCCTTGGGACAAATGACCCAGAAAGACA	1200
1201	GCCATGCCCTCCGAAAGAGGGCCAGGTCCCTAAGCTGCTGCTTGTGCGGACTGCACCCGGC	1260
1261	ATTCTCTTACCTGAGGCTTTCCTGTCGCCAGTGCAAGAACCCGGTGTCTGCTGGG	1316

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FIGURE 14B

20	E	L	F	A	S	R	I	M	N	V	T	S	A	T	L	S	S	L	S
40	V	N	Y	G	T	D	R	I	Q	S	V	E	A	V	F	K	P	V	T
60	S	G	N	F	R	L	L	P	N	C	Y	S	K	F	A	N	R	H	A
80	G	L	T	P	V	S	P	L	F	T	I	S	T	F	T	L	R	T	P
100	F	I	K	M	L	I	T	F	K	I	R	A	E	P	W	F	Y	S	P
120	G	A	I	V	C	C	M	S	R	A	Y	G	T	A	V	A	R	G	H
140	E	M	K	L	F	C	K	I	K	Y	Y	A	R	W	Q	Y	E	N	C
160	E	L	R	V	V	L	N	F	E	P	A	R	M	C	G	L	D	I	Q
180	S	I	L	S	E	H	R	T	I	K	L	Q	R	L	P	F	D	T	S
200	S	V	Q	V	G	F	Y	P	L	N	V	L	H	C	V	P	C	T	E
220	V	T	R	L	Y	I	V	I	D	V	M	M	T	F	T	N	C	T	W
240	N	S	D	L	I	S	L	V	I	M	L	Q	S	C	Y	L	L	S	Q
260	A	L	W	D	W	A	P	W	I	F	L	I	H	G	D	G	I	C	G
280	D	F	C	A	I	T	Q	C	G	V	F	Q	Q	M	I	S	I	P	G
300	L	T	V	F	D	T	C	G	I	C	P	H	D	I	F	N	L	V	C
320	K	L	A	A	Q	L	C	G	N	Y	I	A	A	I	P	I	F	T	E
340	D	L	V	L	V	L	I	L	N	T	Y	H	S	C	D	Y	A	Q	V
360	M	V	M	S	L	V	A	M	W	S	F	E	Q	L	V	G	R	G	A
380	V	V	V	E	D	Y	L	G	N	A	K	P	T	I	L	R	L	D	L
400	K	L	I	I	L	Y	A	Q	S	V	A	R	K	N	W	F	I	R	P

FIGURE 15-1

FIGURE 15-1, 22/32
FIGURE 15-2, 23/32
FIGURE 15-3, 24/32
FIGURE 15-4, 25/32

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```
7  ATGGACAAACTTGA TGCTAA TG TGAGTTCTGAGGAGGGTTTCGGGTCAGT 56
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
152 ATGGACAGACTTGA TGCTAA TG TGAGTTCC AACGAGGGTTTCGGGTCGTGT 201
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

57  GGAGAAGGTGGT GCTGCTCAGCTTCTCTCGACGGTTATCCTGATGGCCA 106
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
202 GGAGAAGGTGCTACTGCTCAGCTTCTTCGCAATGGTTATCCTGATGGCCA 251
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

107 TCTTGGGGAACCTGCTGGTGATGGTGGCTGTGTGCTGGGACAGGCAGCTC 156
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
252 TCC TGGCAACCTGCTGGTGATGGTGTGCTGTGTCAGGACAGGCAGCTC 301
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

157 AGGAAAAATAAAACAAATTATTTCA TTGTATCTCTTGCTTTTGCGGATCT 206
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
302 AGGAAAAATAAAACCAATTATTTCA TTGTGTCTCTTGCCCTTTGCTGATCT 351
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

207 GCTGGTTTCGGTGCTGGTGATGCCCTTTGGTGCCATTGAGCTGGTTCAAG 256
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
352 GCTGGTTTCGGTGCTGGTGAA TGCCCTTCGGTGCCATTGAGTTGGTCCCAAG 401
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

257 ACATCTGGATTATGGGAGGTGTTTGTCTTTGTTCGGACATCTCTTGAC 306
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
402 ACATCTGGTTTATGGGGagatgttttgccctgggtccggacctctctggat 451
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

307 GTCCTGCTCACAACGGCATCGATTTTTCACCTGTGCTGCCATTTCTCTGGA 356
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
452 gtcctactcaccacagcatcaatttttcacctctgctgcatttccctGGA 501
   ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
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FIGURE 15-2

357 TAGGTATTACGCCATCTGCTGCCAGCCTTTGGTCTATAGGAACAAGATGA 406
|||||
502 TAGGTATTATGCCATCTGCTGTCAACCTTTGGTTTATAGAAACAAGATGA 551
|||||
407 CCCCTCTGCGCATCGCATTAATGCTGGGAGGCTGCTGGTCAATCCCCACG 456
|||||
552 CCCCTCTACGCATCGCATTAATGCTGGGAGGCTGCTGGTCAATCCCCATG 601
|||||
457 TTATATTCTTTTCTCCCTATAATGCAAGGCTGGAATAACATTGGCATAAT 506
|||||
602 TTATATATCTTTTCTCCCTATAATGCAAGGCTGGAACAACATCGGCATAGT 651
|||||
507 TGATTTGATAGAAAGAGGAAGTTCAACCAGAACTCTAACTCTACGTACT 556
|||||
652 TGATGTGATAGAGAAAAGGAAATTCAACCACAACTCTAACTCTACATTCT 701
|||||
557 GTGCTTTCATGGTCAACAAGCCCTACGCCATCACCTGCTCTGTGGTGCC 606
|||||
702 GTGCTTTCATGGTCAACAAGCCCTATGCCATCACCTGCTCTGTGGTGCC 751
|||||
607 TTCTACATCCCATTCTCTCATGGTGTGGCCCTATTACCGCATCTATGT 656
|||||
752 TTCTACATCCCCTTTCTCTCATGGTGTGGCCCTATTACCGTATCTATGT 801
|||||

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FIGURE 15-3

657 CACAGCTAAGGAGCATGCCCATCAGATCCAGATGTTACAACGGGCAGGAG 706
|||||
802 CACTGCTAAGGAGCATGCCCCAGCAGATCCAGATGTTACAACGGGCAGGAG 851
|||||
707 CCTCCTCCGAGAGCAGGCCCTCAGTCGGCAGACCAGCATAAGCATTATCGC 756
|||||
852 CCACCTCTGAAGCAGGCCCCAGACAGCTGACCAGCACACATCGC 901
|||||
757 ATGAGGACAGAGACCAAGCAGCCAAAGACCCTGTGCATCATCATGGGTTG 806
|||||
902 ATGCGGACAGAGACCAAGCAGCCAAAGACTTATGTGTATCATGGGCTG 951
|||||
807 CTTCTGCCCTCTGCTGGGCACCATTTCTTGTACCAATAATTGTGGATCCTT 856
|||||
952 CTTCTGTTCTGCTGGGCCCTTCTTTGTACCAATAATTGTGGACCTT 1001
|||||
857 TCATAGACTACACTGTCTCCCTGGGCAGGTGTGGACTGCTTTCCCTCTGGCTC 906
|||||
1002 TCATAGACTACACTGTGCCCCGAGAAGGTGTGGACTGCTTTCCCTCTGGCTT 1051
|||||
907 GGCTATATCAATTCCGGGTTGAACCCCTTTTCTCTACGCCCTTCTTGAATAA 956
|||||
1052 GGCTATATCAATTCCAGGGTTGAACCCCTTTTCTCTATGCCCTTCTTGAATAA 1101
|||||

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FIGURE 15-4

```
957  GTCTTTTAGACGTGCCCTTCCTCATCATCCTCTGCTGTGATGATGAGCGCT 1006
      ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
1102  GTCTTTCAGACGTGCCCTTCCTTATCATCCTCTGCTGTGATGATGAGCGCT 1151

1007  ACCGAAGACCTTCCATTCTGGGCCAGACTGTCCCTTGTTCAACCACAACC 1056
      || ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
1152  ACAAAAGACCCCCCATTTCTGGGCCAGACTGTCCCTTGTTCAACCACAACC 1201

1057  ATTAATGGATCCACACATGTACTAAGGGATGCAGTGGAGTGTGGTGGCCA 1106
      ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
1202  ATTAATGGATCCACTCATGTGCTAAGGGATACAGTGGAAATGTGGTGGCCA 1251

1107  GTGGGAGAGTCAGTGTACCCGCCAGCAACTTCTCCTTTGGTGGCTGCTC 1156
      ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
1252  ATGGGAGAGTCGGTGTACCTCACCCTCACAGCAACTTCTCCTTTGGTGGCTGCTC 1301

1157  AGCCCAGTGACACTTAG..... 1173
      || ||||| || |||||

1302  AG.CCAGTGATACGTAGGCCCCAGGACAATGACCTAGAAGACAGCTGTAG 1350
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FIGURE 16-1, 26/32
FIGURE 16-2, 27/32

FIGURE 16-1

```
1 MDKLDANVSSEEGFSGVEKVLLTFLSTVILMAILGNLLVMVAVCWRQL 50
  :::::::::::::::::::::
1 MDRLDANVSSNEGFGSVEKVLLTFFAMVILMAILGNLLVMVAVCRDRQL 50
  :::::::::::::::::::::

51 RKIKTNYFIVSLAFADLLVSVLMPFGAIELVQDIWIYGEVFCCLVRTSLD 100
  :::::::::::::::::::::
51 RKIKTNYFIVSLAFADLLVSVLVNAFGAIELVQDIWFYGEVFCCLVRTSLD 100
  :::::::::::::::::::::

101 VLLTTASIFHLCCISLDRYYAICCCQPLVYRNKMTPLRIALMLGGCWVPT 150
  :::::::::::::::::::::
101 VLLTTASIFHLCCISLDRYYAICCCQPLVYRNKMTPLRIALMLGGCWVPM 150
  :::::::::::::::::::::

151 FISFLPIMQGWNIGIIDLIEKRKFNSNSTYCVFMVNKPYAITCSVVA 200
  :::::::::::::::::::::
151 FISFLPIMQGWNIGIVDVEKRKFNSNSTFCVFMVNKPYAITCSVVA 200
  :::::::::::::::::::::

201 FYIPFLMLVAYRIYVTAKEHAHQIQMLQRAGASSESRPQADQHSRTHR 250
  :::::::::::::::::::::
201 FYIPFLMLVAYRIYVTAKEHAQQIQMLQRAGATSESRPQTADQHSRTHR 250
  :::::::::::::::::::::
```

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FIGURE 16-2

```
251 MRTETKAAKTLICIIMGCFCCLWAPFFVTNIVDPFIDYTVPGQVWTAFLWL 300
|||||:|||||:|||||:|||||:|||||:|||||:|||||:|||||:
251 MRTETKAAKTLCVIMGCFCFCWAPFFVTNIVDPFIDYTVPEKVWTAFLWL 300

301 GYINSGLNPFLYAFLNKSFRRAFLIILCCDDERYRRPSILGQTVPCSTTT 350
|||||:|||||:|||||:|||||:|||||:|||||:|||||:|||||
301 GYINSGLNPFLYAFLNKSFRRAFLIILCCDDERYKRPPILGQTVPCSTTT 350

351 INGSTHVLRDAVECGGQWESQCHPPATSPLVAAQPSDT*..... 389
|||||:|||||:|||||:|||||:|||||:|||||:|||||:|||||
351 INGSTHVLRDVECGGQWESRCHLTATSPLVAAQPVIRRPQDNDLEDSCS 400

401 LKRSQS*..... 406
```

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FIGURE 17-1

FIGURE 17-1, 28/32
FIGURE 17-2, 29/32
FIGURE 17-3, 30/32

357 TAGGTATTACGCCATCTGCTGCCAGCCTTTGGTCTATAGGAACAAGATGA 406
|||||
1 TTGGTCTATAGGAACAAGATGA 22

407 CCCCTCTGCGCATCGCAATTAATGCTGGAGGCTGCTGGTCAATCCCCACG 456
|||||
23 CCCCTCTGCGCATCGCAATTAATGCTGGAGGCTGCTGGTCAATCCCCACG 72

457 TTTATTCTTTTCTCCCTATAATGCAAGGCTGGAATAACATTGGCATAAT 506
|||||
73 TTTATTCTTTTCTCCCTATAATGCAAGGCTGGAATAACATTGGCATAAT 122

507 TGATTGTAGAAAAGAGGAAGTTCAACCAGAACTCTAACTCTACGTACT 556
|||||
123 TGATTGTAGAAAAGAGGAAGTTCAACCAGAACTCTAACTCTACGTACT 172

557 GTGTCTTCAATGGTCAACAAGCCCTACGCCATCACCTGCTCTGTGTGGCC 606
|||||
173 GTGTCTTCAATGGTCAACAAGCCCTACGCCATCACCTGCTCTGTGTGGCC 222

607 TTCTACATCCCATTCTCCTCATGGTGCTGGCCCTATTACCGCATCTATGT 656
|||||
223 TTCTACATCCCATTCTCCTCATGGTGCTGGCCCTATTACCGCATCTATGT 272

657 CACAGCTAAGGAGCATGCCCATCAGATCCAGATGTTACAACGGGCAGGAG 706
|||||
273 CACAGCTAAGGAGCATGCCCATCAGATCCAGATGTTACAACGGGCAGGAG 322

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FIGURE 17-2

707 CCTCCTCCGAGAGCAGGCCCTCAGTCGGCAGACCAGCATAGCACTCATCGC 756
|||||
323 CCTCCTCCGAGAGCAGGCCCTCAGTCGGCAGACCAGCATAGCACTCATCGC 372
757 ATGAGGACAGAGACCAAGCAGCCAGACCCTGTGCATCATATGGGTTG 806
|||||
373 ATGAGGACAGAGACCAAGCAGCCAGACCCTGTGCATCATATGGGTTG 422
807 CTTCTGCCCTCTGCTGGGCACCATCTTTGTACCAATAATTGTGGATCCTT 856
|||||
423 CTTCTGCCCTCTGCTGGGCACCATCTTTGTACCAATAATTGTGGATCCTT 472
857 TCATAGACTACACTGTCCCTGGGCAGGTGTGGACTGCTTTCCCTCTGGCTC 906
|||||
473 TCATAGACTACACTGTCCCTGGGCAGGTGTGGACTGCTTTCCCTCTGGCTC 522
907 GGCTATATCAATTCCGGGTGAACCCCTTTTCTCTACGCCCTTCTTGAATAA 956
|||||
523 GGCTATATCAATTCCGGGTGAACCCCTTTTCTCTACGCCCTTCTTGAATAA 572
957 GTCTTTTAGACGTGCCCTTCCCTCATCATCCCTCTGCTGTGATGATGAGCGCT 1006
|||||
573 GTCTTTTAGACGTGCCCTTCCCTCATCATCCCTCTGCTGTGATGATGAGCGCT 622

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FIGURE 17-3

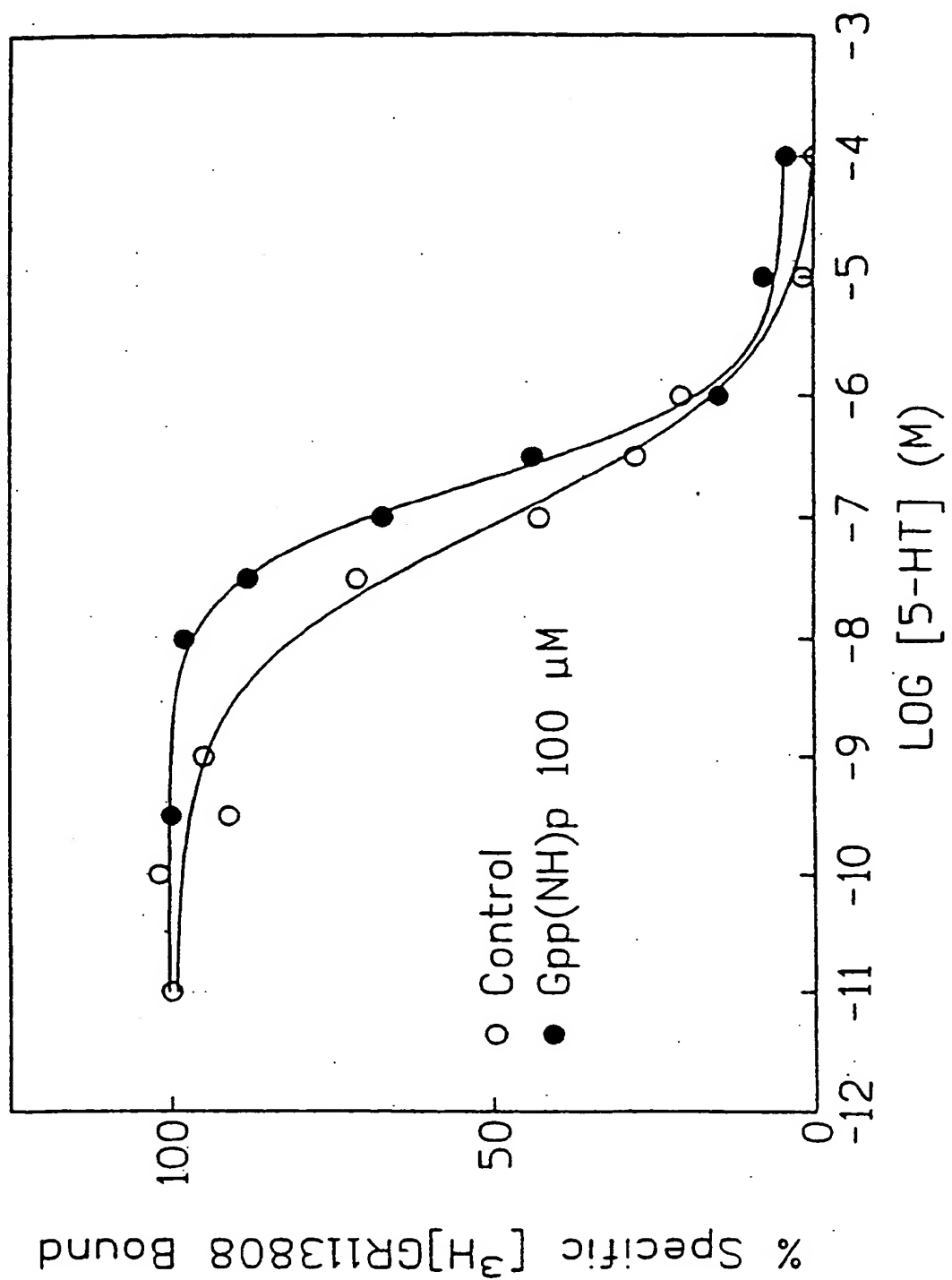
1007 ACCGAAGACCTTCCATTCTGGGCCAGACTGTCCCTTGTTCAACCACAACC 1056
|||||
623 ACCGAAGACCTTCCATTCTGGGCCAGACTGTCCCTTGTTCAACCACAACC 672
|||||
1057 ATTAATGGATCCACACACATGTACTAAGGGATGCAGTGGAGTGTGGTGGCCA 1106
|||||
673 ATTAATGGATCCACACACATGTACTAAGGTACACCGTTCTGCACAGGGGACA 722
|||||
1107 GTGGGAGAGTCAGTGTCAACCCGCCAGCAACTTCTCCTTTGGTGGCTGCTC 1156
|||
723 TCATCAGGAACCTCGAGAAACTGCCCCATACACAATGACCCAGAAATCCCTGG 772
|||
1157 AGCCCAGTGACACTTAG 1173
|||
773 AATCATGCTTCTGA... 786

FIGURE 18

1101	VLLTTASIFHLCCISLDRYYAICCCQPLVYRNKMTPLRIALMLGCVIPT	150
1LVYRNKMTPLRIALMLGCVIPT	24
1151	FISFLPIMQGWNNGIIDLIEKRKFQNSNSTYCVFMVNKPYAITCSVVA	200
25	FISFLPIMQGWNNGIIDLIEKRKFQNSNSTYCVFMVNKPYAITCSVVA	74
201	FYIPFLLMVLAYYRIYVTAKEHAHQIQMLQAGASSESRPQSAHQHSTHR	250
75	FYIPFLLMVLAYYRIYVTAKEHAHQIQMLQAGASSESRPQSAHQHSTHR	124
251	MRTETKAAKTLCIIMGCFCLCWAPFFVTNIVDPFIDYTPVGQVWTAFLWL	300
1125	MRTETKAAKTLCIIMGCFCLCWAPFFVTNIVDPFIDYTPVGQVWTAFLWL	174
301	GYINSGLNPFLYAFLNKSFRAFLIILCCDDERYRRPSILGQTVPCSTTT	350
1175	GYINSGLNPFLYAFLNKSFRAFLIILCCDDERYRRPSILGQTVPCSTTT	224
351	INGSTHVLRDAVECGGWESQCHPPATSPLVAAQPSDT*	389
225	INGSTHVLRYTVLHRGHHQEKLPIHNDPESLESCF*	262

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FIGURE 19



INTERNATIONAL SEARCH REPORT

International application No.

PCT/US93/12586

A. CLASSIFICATION OF SUBJECT MATTER

IPC(S) : C12N 1/15, 1/21, 5/10, 15/00, 15/12

US CL : 435/6, 172.3, 240.2, 252.3, 255.1, 320.1; 536/23.5, 24.3, 24.31

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/6, 172.3, 240.2, 252.3, 255.1, 320.1; 536/23.5, 24.3, 24.31

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS and DIALOG (files 5, 155, 351, 357, 358) search terms: serotonin, receptor, DNA, 5-HT

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US, A, 4,985,352 (JULIUS ET AL) 15 January 1991, see Figure 1.	25-28
A, P	US, A, 5,242,822 (MARULLO ET AL) 07 September 1993, see abstract and claims.	1-37, 40-42
A	FEBS, Volume 312, Number 2,3, issued November 1992, Loric et al, "New mouse 5-HT ₂ -like receptor", pages 203-207, see entire document.	1-37 and 40-42

☐ Further documents are listed in the continuation of Box C.☐ See patent family annex.

* Special categories of cited documents:	* T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
* A* document defining the general state of the art which is not considered to be part of particular relevance	* X	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
* E* earlier document published on or after the international filing date	* Y	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
* L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	* Z	document member of the same patent family
* O* document referring to an oral disclosure, use, exhibition or other means		
* P* document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

07 June 1994

Date of mailing of the international search report

JUN 20 1994

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Telephone No. (703) 308-0196

Form PCT/ISA/210 (second sheet)(July 1992)*

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US93/12586

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
1-37 and 40-42

Remark on Protest

☐
☐

- The additional search fees were accompanied by the applicant's protest.
No protest accompanied the payment of additional search fees.

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

I. Claims 1-37 and 40-42, drawn to nucleic acid sequences for the 5-HT₄ receptor, vectors, host cells, probes, antisense nucleic acids, and a method for detecting expression of a mammalian 5-HT₄ receptor, classified in at least, for example, Class 536, subclass 23.5.

II. Claims 38-39, drawn to a method of isolating a gene, classified in at least, for example, Class 435, subclass 172.3.

III. Claims 43-44, drawn to a 5-HT₄ receptor protein, classified in at least, for example, Class 530, subclass 350.

IV. Claims 45-48, drawn to a method of production of a mammalian 5-HT₄ receptor, classified in at least, for example, Class 435, subclass 69.1.

V. Claims 49-54 and 61, drawn to an antibody to a 5-HT₄ receptor protein, classified in at least, for example, Class 530, subclass 387.1.

VI. Claim 55, drawn to a pharmaceutical composition to alleviate abnormalities resulting from overexpression of a 5-HT₄ receptor, classified in at least, for example, Class 514, subclass 2.

VII. Claim 56, drawn to a pharmaceutical composition to alleviate abnormalities resulting from underexpression of a 5-HT₄ receptor, classified in at least, for example, Class 514, subclass 2.

VIII. Claims 57-60, drawn to pharmaceutical compositions containing oligonucleotides, classified in at least, for example, Class 514, subclass 44.

IX. Claims 62-66, drawn to transgenic animals, classified in at least, for example, Class 800, subclass 2.

X. Claims 67-68, drawn to a method for determining physiological effects of varying levels of 5-HT₄ receptors using transgenic animals, classified in at least, for example, Class 514, subclass 44.

XI. Claims 69-70, drawn to a method for determining specific binding using whole cells, classified in at least, for example, Class 435, subclass 7.2.

XII. Claims 71-72, drawn to a method of screening compounds to identify drugs using whole cells, classified in at least, for example, Class 435, subclass 7.2.

XIII. Claims 73-74, drawn to a method for determining specific binding using cell extracts, classified in at least, for example, Class 435, subclass 7.1.

XIV. Claims 75-76, drawn to a method of screening compounds to identify drugs using membrane fractions, classified in at least, for example, Class 435, subclass 7.1.

XV. Claims 77-82, drawn to a method for identifying a compound using whole cells and activation blockade, classified in at least, for example, Class 435, subclass 7.2.

XVI. Claims 83-84, drawn to compounds and pharmaceutical compositions, classified in at least, for example, Class 514, subclass 2.

XVII. Claim 85, drawn to a method for detecting 5-HT₄ receptor on the cell surface using an antibody, classified in at least, for example, Class 435, subclass 7.2.

XVIII. Claim 86, drawn to a method for treating an abnormal condition related to excess activity of 5-HT₄ receptor, classified in at least, for example, Class 514, subclass 2.

XIX. Claim 87, drawn to a method for treating an abnormal condition alleviated by increasing the activity of 5-HT₄ receptor, classified in at least, for example, Class 514, subclass 2.

XX. Claims 88-89, drawn to a method for diagnosing a predisposition to a disorder associated with expression of 5-HT₄ receptor alleles, classified in at least, for example, Class 435, subclass 6.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US93/12586

XXI. Claim 90, drawn to a method of identifying a substance that alleviates abnormalities from overexpression of 5-HT₄ receptors using transgenic animals, classified in at least, for example, Class 424, subclass 9.

XXII. Claim 91, drawn to a method of identifying a substance that alleviates abnormalities from underexpression of 5-HT₄ receptors using transgenic animals, classified in at least, for example, Class 424, subclass 9.

XXIII. Claim 92, drawn to a method of treating abnormalities alleviated by reduced expression of 5-HT₄, classified in at least, for example, Class 514, subclass 44.

XXIV. Claim 93, drawn to a method of treating abnormalities resulting from underexpression of 5-HT₄ receptor, classified in at least, for example, Class 514, subclass 2.





INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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With international search report.

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claims and to be republished in the event of the receipt of
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18 August 1994 (18.08.94)(54) Title: DNA ENCODING 5-HT₄ SEROTONIN RECEPTORS AND USES THEREOF

(57) Abstract

This invention provides an isolated nucleic acid molecule encoding a mammalian 5-HT₄ receptor and an isolated nucleic acid molecule encoding a human 5-HT₄ receptor, an isolated protein which is a mammalian 5-HT₄ receptor, an isolated protein which is a human 5-HT₄ receptor, vectors comprising an isolated nucleic acid molecule encoding a mammalian 5-HT₄ receptor, vectors comprising an isolated nucleic acid molecule encoding a human 5-HT₄ receptor, mammalian cells comprising such vectors, antibodies directed to the 5-HT₄ receptor, nucleic acid probes useful for detecting nucleic acid encoding a mammalian or human 5-HT₄ receptor, antisense oligonucleotides complementary to any sequences of a nucleic acid molecule which encodes a mammalian or human 5-HT₄ receptor, pharmaceutical compounds related to the human 5-HT₄ receptor, and nonhuman transgenic animals which express DNA encoding a normal or a mutant mammalian or human 5-HT₄ receptor. This invention further provides methods for determining ligand binding, detecting expression, drug screening, and treatments for alleviating abnormalities associated with a human 5-HT₄ receptor.

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FR	France			VN	Viet Nam
GA	Gabon				

A. CLASSIFICATION OF SUBJECT MATTER IPC(5) : C12N 1/15, 1/21, 5/10, 15/01, 5/12 US C6 : 435/6, 172.3, 240.2, 252.3, 320.1; 536/23.5, 24.3, 24.31 According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) U.S. : 435/6, 172.3, 240.2, 252.3, 255.1, 320.1; 536/23.5, 24.3, 24.31 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) APS and DIALOG (files 5, 155, 351, 357, 358) search terms: serotonin, receptor, DNA, 5-HT		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US, A, 4,985,352 (JULIUS ET AL) 15 January 1991, see Figure 1.	25-28
A, P	US, A, 5,242,822 (MARULLO ET AL) 07 September 1993, see abstract and claims.	1-37, 40-42
A	FEBS, Volume 312, Number 2,3, issued November 1992, Loric et al, "New mouse 5-HT ₂ -like receptor", pages 203-207, see entire document.	1-37 and 40-42
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
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Date of the actual completion of the international search 07 June 1994	Date of mailing of the international search report JUN 20 1994	
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230	Authorized officer Marianne Porta Allen <i>M. Porta Allen</i> Telephone No. (703) 308-0196	

Form PCT/ISA/210 (second sheet)(July 1992)*

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
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1-37 and 40-42

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

Form PCT/ISA/210 (continuation of first sheet(1))(July 1992)*

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